Tick Spit Shines a Light on the Initiation of Coagulation

Running title: Aleman et al.; Ticks illuminate the initiation of coagulation

Maria M. Aleman, BS¹; Alisa S. Wolberg, PhD¹,²

¹Dept of Pathology and Laboratory Medicine; ²McAllister Heart Institute, University of North Carolina at Chapel Hill, Chapel Hill, NC

Address for Correspondence:
Alisa S. Wolberg, PhD
Department of Pathology and Laboratory Medicine
University of North Carolina at Chapel Hill
815 Brinkhous-Bullitt Building, CB #7525
Chapel Hill, NC 27599-7525
Tel: 919-966-8430
Fax: 919-966-6718
E-mail: alisa_wolberg@med.unc.edu

Journal Subject Codes: Anticoagulants: [159] Anticoagulant mechanisms, Basic science research:[130] Animal models of human disease

Key words: Editorial, anticoagulant, animal model of human disease remodeling
Mammalian blood has numerous essential and well-known functions, including oxygen and nutrient delivery. This elixir is recognized by blood-feeding species of mosquitoes, ticks, fleas, lice, leeches, and bats that rely on blood meals for nutrition, life cycle progression and survival. To obtain these blood meals that require minutes to a week or longer to complete, these blood-sucking creatures must thwart endogenous defense systems contained within blood—immune and procoagulant cells and plasma proteins that rapidly clot (within 3-4 minutes) to provide first-line defense against breaches in vascular integrity. In a fascinating display of evolutionary agility, hemovores have adapted elegant mechanisms to evade detection and prevent blood coagulation by synthesizing an extensive armament of molecules with anesthetic, immunosuppressive, vasodilatory, anticoagulant, and profibrinolytic properties in mammals. Research characterizing the molecules generated by hemovores to bypass mammalian defense pathways has revealed exciting new mechanisms and in some cases, novel therapeutic approaches for anticoagulation.

In particular, ticks have received considerable attention for their remarkable evolutionary adaptations to life as obligate hemovores. Briefly, the typical tick lifecycle includes four stages: egg, six-legged larva, eight-legged nymph, and adult (Figure 1), and takes 1-3 years to complete this full cycle. Ticks must consume blood at the larval, nymph and adult stages to survive, and die if they do not find a host. Interestingly, while neither larva nor nymphs have overt sexual differentiation, adults are fully differentiated into males and females, and compared to males, longer nymph feeding is required for expression of female characteristics. Consequently, tick saliva contains multiple proteins that maintain blood fluidity to enable feeding, and therefore tick survival.

Using yeast surface display, Schuijt and colleagues previously identified several novel
tick salivary proteins that promote feeding. Characterization of these proteins revealed anti-complement (P8, later termed Tick Salivary Lectin Pathway Inhibitor [TSLPI])\textsuperscript{6} and anti-coagulant (P23) activities. Interestingly, when rabbits were immunized against a cocktail containing these recombinant proteins, nymph feeding was reduced, and this reduction had fascinating consequences. Compared to tick nymphs fed on control rabbits, nymphs fed on rabbits immunized against tick salivary proteins were significantly smaller. Further, this reduction in weight had a profound effect on the sexual maturation of ticks into adults. Schuijt et al\textsuperscript{6} observed that nymphs that reached 3.4 mg or greater molted into female adults; whereas, nymphs 3.3 mg and smaller molted into male adults.\textsuperscript{6} Notably, the smaller “male” nymphs were composed of two distinct populations, prompting speculation that the larger of these populations were “failed females” unable to reach sexual maturation. These data suggest that altering feeding in a way that even subtly decreases mean nymph size could profoundly alter adult sex ratios and decrease tick numbers in subsequent generations. These findings suggest anti-complement and anti-coagulant proteins in tick saliva are potential vaccine candidates for reducing tick populations, as well as reducing transmission of tick-borne illnesses.

In the current issue of Circulation, Schuijt et al\textsuperscript{7} have now extended their work by characterizing the biological target of the anticoagulant protein P23 (now termed Tick Inhibitor of factor Xa towards factor V [TIX-5]), revealing additional features in this already intriguing story. Recombinant TIX-5 (rTIX-5) delays thrombin generation in human plasma in which clotting is initiated via intrinsic or extrinsic activators, and in factor VIII- or factor XI-deficient plasmas, suggesting its molecular target lies within the common pathway. Accordingly, rTIX-5 is unable to inhibit thrombin generation in the presence of pre-activated factor V (FV), indicating rTIX-5 inhibits clotting by delaying FV activation. Unexpectedly, however, the inhibitory
effects of rTIX-5 on FV activation were not observed in reactions triggered by thrombin or meizothrombin, the hypothesized activators of FV. Rather, rTIX-5 inhibits factor Xa/phospholipid-mediated generation of factor Va (FVa). Schuijt et al\(^7\) further interrogated the nature of this mechanism and showed that rTIX-5 does not inhibit factor Xa substrate cleavage or even bind directly to factor Xa, but instead specifically blocks factor Xa-mediated activation of FV in a FV B-domain-dependent fashion. Thus, this study has not only identified a novel anticoagulant protein in ticks, but importantly, the identification of TIX-5’s biological target reveals the physiological importance of a pathway not previously described during mammalian coagulation in vivo.

This work has important implications for several hot topics in coagulation research. First, the identification of the molecular target of TIX-5 sheds light on a long-standing question regarding the origin of FVa during the initiation of coagulation. Although several proteases including \(\alpha\)-thrombin\(^8\), meizothrombin\(^9\), calpain\(^10\), plasmin\(^11\), elastase and cathepsin G\(^12\), and factor Xa\(^13\) can cleave FV to generate the active cofactor (FVa) in purified systems, the primary activator of FV during the initiation of coagulation has remained elusive. This activity has primarily been attributed to trace amounts of meizothrombin and/or \(\alpha\)-thrombin generated by the extrinsic activation of factor Xa. Since rTIX-5 specifically delays factor Xa-, but not thrombin- or meizothrombin-mediated activation of FV, this protein provides a unique tool to study the role of factor Xa-mediated activation of FV during coagulation. Schuijt et al\(^7\) show that rTIX-5 prolongs the lag time to thrombin generation in both human and rabbit plasma, as well as in human whole blood. Importantly, Schuijt et al show that when rabbits are immunized with rTIX-5, post-feeding weights of adult ticks are significantly reduced compared to controls, indicating that inhibiting TIX-5 prevents the natural anticoagulation mechanism needed for
optimal feeding. This simple, but elegant, assay shows for the first time, the relevance of this pathway during coagulation in vivo.

Second, the identification of rTIX-5 yields a valuable new tool to characterize the molecular mechanisms that maintain the procofactor state of circulating FV and the conversion of FV to active cofactor FVa. During coagulation, proteolytic removal of the large central B-domain of FV eliminates steric constraints provided by the B-domain that block factor V(a) activity; however, the nature of the steric inhibition mechanism has been elusive because FV(a) does not require proteolysis to acquire its activity.\textsuperscript{14} Bos and Camire\textsuperscript{15} recently identified two evolutionarily-conserved sequences in the B-domain, one acidic and one basic, that define minimal sequence requirements for FV’s autoinhibitory function. The ability of rTIX-5 to inhibit FV molecules with mutations in the cleavage sites, but not inhibit B-domain-deleted FV, demonstrates that TIX-5 does not simply block proteolysis, but instead interferes with this B-domain-dependent inhibitory mechanism. Notably, rTIX-5 can bind to both the basic and acidic regions of the FV B-domain, as well as phospholipids, and the inhibitory activity of rTIX-5 is supported by the presence of both the B-domain acidic and basic regions. These data suggest rTIX-5 forms a complex with the FV B-domain and the phospholipid surface that blocks the accessibility of factor Xa to FV. Further, since factor Xa activates the FV QIQQQ variant that lacks all specific factor Xa activation sites, other arginine residues in the B-domain or heavy and light chains must also support partial activation of FV. Future studies to model these interactions on the molecular level promise critical information regarding the role of the B-domain in mediating the procofactor to cofactor transition during FV activation, and potentially, a means to modulate this transition and control coagulation.

Third, it is interesting that ticks utilize both anti-complement and anti-coagulant strategies
to facilitate feeding and maturation over their life cycles. Coagulation and complement pathways are both ancient serine protease defense mechanisms, components of which have been in existence since the divergence of lamprey eels from jawed vertebrates over 600 million years ago, and several studies have linked these pathways in modern physiology. For example, factor XIIa can activate complement factor C1r, a subcomponent of C1 which initiates the classical complement pathway, and both thrombin and factor Xa can activate complement factors C3 and C5, members of the common complement pathway. The lectin pathway, which is inhibited by the tick salivary protein TSLP, has been shown to promote prothrombin activation (reviewed in [19]). These observations suggest multiple levels of cross-talk between these systems, such that ticks must inhibit these pathways at several points during certain life stages to accomplish their blood meal. Indeed, nymphs can inhibit either coagulation or complement to obtain a blood meal and mature to adult size; whereas, adults must block the coagulation pathway for sufficient feeding (Figure 1). The use of these and other tick salivary proteins may therefore yield additional information about cross-talk between complement and coagulation, with important implications for both thrombotic and inflammatory disorders.

Identification of TIX-5 anticoagulant activity and procoagulant pathway gives rise to exciting questions regarding the role of this pathway in coagulation and future studies to evaluate its potential as a therapeutic target. First, in contrast to ticks fed on TIX-5-immunized rabbits, the mean weight of ticks fed on TIX-5-immunized mice is not reduced, suggesting mice do not make antibodies against TIX-5, or that the coagulation pathway inhibited by TIX-5 is less important in mice than in humans or rabbits. Since mice are a primary go-to model for human coagulation studies, these data suggest caution in the interpretation of murine studies evaluating initiating events in coagulation. Second, recent studies have shown that in contrast to platelets,
when prothrombinase is assembled on the surface of erythrocytes, thrombin generation proceeds via the meizothrombin intermediate\textsuperscript{21}, indicating that procoagulant pathways are determined by both plasma protein composition and the nature of the cell surface. The observation that rTIX-5 can bind directly to phospholipids suggests the composition of the cell surface could also influence TIX-5’s ability to block factor Xa activation of FV. It will be interesting to determine the role of the factor Xa/FV pathway on different cells and under different initiating circumstances.

Finally, and perhaps most attractive, by demonstrating the physiological relevance of factor Xa-mediated activation of FV in vivo and the ability of rTIX-5 to inhibit this reaction, this work has yielded a potential new antithrombotic approach. Both hemovores and clinicians share a goal with regard to anticoagulation – the need to reduce clotting in a highly-controlled way that does not cause excessive bleeding. Indeed, anticoagulant mechanisms developed by nature have previously been exploited for the development of other drugs, including hirudin and analogs from leeches and defibrinating enzymes from snakes.\textsuperscript{3} In this case, ticks have identified and inhibited a procoagulant pathway that researchers had not yet characterized in vivo. Can we once again exploit nature’s efforts to regulate coagulation in the clinic? This exciting possibility warrants studies to test the antithrombotic potential of TIX-5 and similar molecules in “traditional” thrombosis models that have shown clinical relevance during the development of other antithrombotic drugs. Since the inhibitory effect of rTIX-5 is reduced in plasmas with reduced anticoagulant levels, targeting this pathway may be less effective in certain types of coagulopathies stemming from deficiency in anticoagulant pathways (e.g. protein C deficiency). It will be critical to explicitly test the ability of TIX-5 and similar molecules to prevent thrombosis in specific models of plasma hypercoagulability and vascular dysfunction in future studies.
Funding Sources: This work was partially supported by National Institutes of Health grants R01HL094740 (to ASW) and F31HL112608 (to MMA).

Conflict of Interest Disclosures: None.

References:


**Figure Legend:**

**Figure 1.** Lifecycle of a typical tick. Larvae, nymphs, and adults require blood meals for survival, and secrete anti-complement (Tick Salivary Lectin Pathway Inhibitor, TSLPI) and anti-
coagulant (Tick Inhibitor of factor Xa towards factor V, TIX-5) proteins to accomplish the blood meal. Schuijt et al.\textsuperscript{7} have now described the function of TIX-5 as an inhibitor of factor Xa-mediated factor V activation during the initiation of coagulation.
Figure 1
Tick Spit Shines a Light on the Initiation of Coagulation
Maria M. Aleman and Alisa S. Wolberg

Circulation. published online July 1, 2013;
Circulation is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2013 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circ.ahajournals.org/content/early/2013/06/25/CIRCULATIONAHA.113.003800

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Circulation can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Circulation is online at:
http://circ.ahajournals.org/subscriptions/