Vascular Endothelial Adrenomedullin-RAMP2 System is Essential for Vascular Integrity and Organ Homeostasis

Running title: Koyama et al.; Adrenomedullin-RAMP2 system in vascular integrity

Teruhide Koyama1; Laura Ochoa-Callejero, PhD2; Takayuki Sakurai, PhD1; Akiko Kamiyoshi, PhD1; Yuka Ichikawa-Shindo, MD, PhD1; Nobuyoshi Inuma, MD, PhD1; Takuma Arai, MD, PhD1; Takahiro Yoshizawa1; Yasuhiro Iesato, MD1; Yang Lei1; Ryuichi Uetake1; Ayano Okimura1; Akihiro Yamauchi1; Megumu Tanaka1; Kyoko Igarashi1; Yuichi Toriyama, MD1; Hisaka Kawate1; Ralf H. Adams, PhD3; Hayato Kawakami, MD, PhD4; Naoki Mochizuki, MD, PhD5; Alfredo Martínez, PhD2; Takayuki Shindo MD, PhD1

1Dept of Cardiovascular Research, Shinshu University Graduate School of Medicine, Matsumoto, Japan; 2Oncology Area, Center for Biomedical Research of La Rioja, Logroño, Spain; 3Dept of Tissue Morphogenesis, University of Münster, Münster, Germany; 4Dept of Anatomy, Kyorin University School of Medicine, Mitaka, Japan; 5Dept of Structural Analysis, National Cardiovascular Center Research Institute, Osaka, Japan

Address for Correspondence:
Takayuki Shindo, MD, PhD
Department of Cardiovascular Research
Shinshu University Graduate School of Medicine
Asahi 3-1-1, Matsumoto, Nagano, 390-8621, Japan
Tel: +81-263-37-3192
Fax: +81-263-37-3437
E-mail: tshindo@shinshu-u.ac.jp

Abstract:

Background—Revealing the mechanisms underlying the functional integrity of the vascular system could make available novel therapeutic approaches. We previously showed that knocking out the widely expressed peptide adrenomedullin (AM) or receptor activity-modifying protein 2 (RAMP2), an AM-receptor accessory protein, causes vascular abnormalities and is embryonically lethal. Our aim was to investigate the function of the vascular AM-RAMP2 system directly.

Methods and Results—We generated endothelial cell-specific RAMP2 and AM knockout mice (E-RAMP2-/-, E-AM-/-). Most E-RAMP2-/- mice died perinatally. In surviving adults vasculitis occurred spontaneously. With aging, E-RAMP2-/- mice showed severe organ fibrosis with marked oxidative stress and accelerated vascular senescence. Later, liver cirrhosis, cardiac fibrosis and hydronephrosis developed. We next used a line of drug-inducible E-RAMP2-/- mice (DI-E-RAMP2-/-) to induce RAMP2-deletion in adults, which enabled us to analyze the initial causes of the aforementioned vascular and organ damage. Early after the induction, pronounced edema with enhanced vascular leakage occurred. In vitro analysis revealed the vascular leakage to be caused by actin disarrangement and detachment of endothelial cells. We found that the AM-RAMP2 system regulates the Rac1-GTP/RhoA-GTP ratio and cortical actin formation, and that a defect in this system causes the disruption of actin formation, leading to vascular and organ damage at the chronic stage after the gene deletion.

Conclusions—Our findings show that the AM-RAMP2 system is a key determinant of vascular integrity and homeostasis from prenatal stages through adulthood. Furthermore, our models demonstrate how endothelial cells regulate vascular integrity and how their dysregulation leads to organ damage.

Key words: arteriosclerosis, vasculature, endothelium, edema, aging
Introduction

Chronic organ dysfunction is the most common cause of morbidity among aging individuals. Generally, the process of organ dysfunction begins slowly, but it is progressive and intractable in its chronic phase. From a mechanistic viewpoint, chronic organ dysfunction can be explained as a disruption of physiological homeostasis and the processes responsible for organ maintenance and repair. This is particularly true for the vascular system, which plays central roles in the maintenance of organ homeostasis\(^1,2\). Revealing the mechanisms underlying the functional integrity of the vascular system could make available novel therapeutic approaches to the treatment of organ dysfunction.

Vascular endothelial cells (ECs) and vasoactive molecules play key roles in the maintenance of vascular homeostasis\(^3,4\). ECs actively secrete a variety of bioactive molecules, including nitric oxide, atrial natriuretic peptide (ANP), prostacyclin and adrenomedullin (AM). Originally identified as a vasodilating peptide isolated from human pheochromocytoma\(^5\), AM is now known to be widely secreted from a variety of organs and tissues, including ECs, and to be involved in a number of biological functions\(^6-13\). Plasma levels of AM are elevated in patients with such cardiovascular diseases as hypertension and congestive heart failure\(^14,15\). Moreover, it was recently reported that plasma AM levels are a highly sensitive marker of chronic kidney disease (CKD) that may be predictive of its prognosis\(^16\). We previously showed that homozygotic AM knockout (KO) (AM\(^{-/-}\)) mice die at mid-gestation due to edema and hemorrhage, and clarified the critical role of AM in angiogenesis\(^17\). Heterozygotic AM KO (AM\(^{+/\cdot}\)) mice grow to adulthood with no apparent deficits, but show accelerated cardiac hypertrophy, fibrosis, renal failure, and arteriosclerosis upon cardiovascular injury. Given these observations, the clinical application of AM has been much anticipated\(^18-22\); however, AM is a peptide with a short half-life in the
bloodstream, which limits its usefulness for the treatment of chronic diseases.

To overcome that limitation, we have been focusing on AM’s receptor system. AM is a member of the calcitonin superfamily and acts via a G protein-coupled seven transmembrane domain receptor, calcitonin-receptor-like receptor (CLR)\(^{23,24}\). The specificity of CLR for its ligands is regulated by a group of three receptor-activity-modifying proteins, RAMP1, -2 and -3. We have shown that homozygotic RAMP2 KO (RAMP2\(^{-/-}\)) mice die in utero. Interestingly, among the RAMP KO mice, RAMP2\(^{-/-}\) mice die from vascular abnormalities similar to those observed in AM\(^{-/-}\) mice\(^{25}\). This suggests RAMP2 is the key determinant of AM’s vascular function.

Our aim in the present study was to clarify the pathophysiological function of the vascular AM-RAMP2 system directly. To accomplish that, we generated and utilized four KO models: EC-specific (E)-RAMP2\(^{-/-}\), aged conventional RAMP2\(^{+/-}\), drug-inducible (DI)-E-RAMP2\(^{-/-}\) and EC-specific (E)-AM\(^{-/-}\) mice. With these models, we were able to determine both the acute and chronic effects of RAMP2 deletion, and demonstrate the contribution made by the AM-RAMP2 system to the maintenance of vascular integrity and organ homeostasis.

**Methods**

**Mouse models**

Vascular endothelial cadherin (VE-cadherin) Cre transgenic mice and mice expressing tamoxifen-inducible Cre-recombinase (Cre-ERT2) under the regulation of VE-cadherin promoter were crossed with floxed RAMP2 mice (RAMP2\(^{flox/flox}\)) to create vascular EC-specific RAMP2 conditional KO mice (E- RAMP2\(^{-/-}\)) and tamoxifen drug-inducible (DI) vascular EC-specific RAMP2 KO mice (DI-E-RAMP2\(^{-/-}\)), respectively.
Quantitative real-time RT-PCR analysis

The primers and probes used are listed in Supplemental Table 1. For other experimental procedures, please refer to Supplemental File.

Results

Generation of vascular EC-specific RAMP2 KO mice

Vascular endothelial cadherin (VE-cadherin) Cre transgenic mice were crossed with floxed RAMP2 mice (RAMP2flox/flox) to create vascular EC-specific RAMP2 conditional KO mice (E-RAMP2-/-) (Fig. 1a). Conventional RAMP2-/- embryos died at mid-gestation (around 14.5 dpc), as we reported previously25. By contrast, EC-specific RAMP2-/- (E-RAMP2-/-) embryos survived until later in development, though most died during the perinatal period (Fig. 1b).

Among the RAMP2flox/flox Cre+/- intercrosses, the estimated survival rate of E-RAMP2-/- mice at 4 weeks of age was less than 5% (Supplemental Table 2). Perinatal stage E-RAMP2-/- mice exhibited systemic edema (Fig. 1c, d), as well as interstitial edema within the intestinal villi (Fig. 1e) and lung (Fig. 1f), and severe hemorrhagic changes within the liver (Fig. 1g). As expected, E-RAMP2-/- mice showed vascular abnormalities, including malformation of aortic ECs (Fig. 1h) and partial detachment of ECs from the basement membrane (Fig. 1i, j).

Abnormalities in the vascular structure of surviving E-RAMP2-/- adults

In the 5% of E-RAMP2-/- mice that survived until adulthood, RAMP2 expression in aortic ECs was 20% of that in their littermate controls (Cont) (data not shown). The surviving E-RAMP2-/- adults also exhibited thinning of the aortic wall and enlarged aortic diameters (Fig. 1k, l). Both systolic and diastolic blood pressures (SBP and DBP) were lower in E-RAMP2-/- than Cont mice (Fig. 1m). Electron microscopic observation revealed that the aortic smooth muscle layer was in
disarray in E-RAMP2-/- mice (Fig. 1o), and that the ECs were detached from the basement membrane and severely deformed (Fig. 1n). These morphological changes in the vascular cells may partially explain the enlargement of the aorta and reduction in blood pressure. Given these observations, we speculated that EC viability was diminished in E-RAMP2-/- mice, which was confirmed by the severe disruption of angiogenesis seen in ex vivo aortic ring assays (Fig. 1p).

**Severe vascular inflammation and organ fibrosis in E-RAMP2-/- adults**

Vasculitic lesions developed spontaneously in surviving E-RAMP2-/- adults, beginning when they were about 6-month-old, and severe infiltration and accumulation of inflammatory cells was observed in blood vessels within the major organs, including the liver (Fig. 2a), kidneys (Fig. 2b) and lungs (Fig. 2c). The infiltrating cells were CD3+ or F4/80+, which is indicative of T cells and macrophages, respectively (Fig. 2d, e). In addition, the expression of inflammatory cytokines and macrophage markers was also upregulated (Fig. 2f).

We speculated that the vascular damage in the surviving adult E-RAMP2-/- mice was the primary cause of the vasculitis. In that regard, it was recently suggested that vascular damage may be the cause of vascular senescence. To test whether these phenomena indicate accelerated senescence, we next evaluated aged conventional RAMP2 KO mice (RAMP2+/-). At 2 years of age, RAMP2+/- mice showed inflammatory cell infiltration of the main organs (Fig. 2g-i).

Senescence-associated (SA)-β-gal staining, which is commonly used to assess accelerated aging, was more intense in the aortas of the aged RAMP2+/- than WT mice (Fig. 2j). Moreover, the SA-β-gal staining could be detected at a much younger age (6 months) in E-RAMP2-/- mice (Fig. 2k).

The presence of activated Akt and p53 in the aorta is another recognized marker of vascular senescence. Levels of Akt and p53 activation were much greater in aged RAMP2+/- than WT
mice (Fig. 2l). No difference was seen between the levels of Akt and p53 activation in younger RAMP2+/− and WT mice, though the inflammatory cell adhesion markers ICAM-1 and VCAM-1 were already upregulated in the young RAMP2+/− animals. This is consistent with the idea that vascular senescence is preceded by vascular damage and inflammation.

More interestingly, organ damage developed spontaneously in E-RAMP2−/− adults. In particular, the livers of E-RAMP2−/− mice appeared cirrhotic by the time the animals were 6 months old (Fig. 3a). Pathological analysis revealed that the cirrhosis-like changes were not caused by damage to parenchymal hepatocytes, but by severe fibrosis along the vasculature within the liver, most likely due to dysfunction of the sinusoidal ECs (Fig. 3b, c). Cardiac enlargement with interstitial fibrosis was also seen in 6-month-old E-RAMP2−/− mice (Fig. 3d, e). In addition, quantitative real-time PCR showed elevated expression of ANP, brain natriuretic peptide (BNP) and transforming growth factor-β (TGF-β), which is indicative of heart failure and fibrosis (Fig. 3f). In addition, spontaneous hydronephrosis (Fig. 3g) and polycystic changes (Fig. 3h) were noted in the kidneys of E-RAMP2−/− mice, as was glomerular enlargement and sclerotic changes (Fig. 3i). Electron microscopy revealed podocyte fusion, hyperplasia of the basal membrane, and partial dehiscence between the glomerular ECs and basal membrane (Fig. 3j). Renal expression of TGF-β and collagen was also upregulated in E-RAMP2−/− mice (Fig. 3k).

**EC-specific AM KO mice showed vascular inflammation and glomerulosclerosis**

We next compared EC-specific AM KO (E-AM−/−) with E-RAMP2−/−. In contrast to E-RAMP2−/− neonates, E-AM−/− neonates were healthy and presented no overtly pathophysiological phenotypes (Supplemental Table 3). We suggest that in E-AM−/− neonates, circulating and/or paracrine AM from sources other than ECs likely attenuate the phenotypes observed in E-RAMP2−/− neonates. In adults, however, the vascular inflammation and organ damage were similar to those seen in
E-RAMP2-/- mice. E-AM-/- mice showed severe infiltration and accumulation of inflammatory cells around the vasculature (Fig. 4a, b). They also showed glomerulosclerotic changes with mesangial expansion and upregulation of inflammatory molecules (Fig. 4c-e).

**RAMP2 deficiency enhances inflammatory reactions in both ECs and leukocytes**

We speculated that the observed vasculitic lesions began with the cellular adhesion of inflammatory cells to ECs. To test that idea, we initially examined the effect of endothelial RAMP2-deficiency on the interaction between ECs and macrophages in vitro. After isolating and culturing LSECs from RAMP2+/- and WT mice, we evaluated the attachment of macrophages isolated from GFP-mice (provided by Prof. Sato of Kagoshima University). We found that GFP+ macrophages attached more readily to RAMP2+/- LSECs than to WT LSECs (Fig. 5a, b), and that VCAM-1 expression was higher in RAMP2+/- LSECs than in WT cells (Fig. 5c).

To then assess the effect of RAMP2-deficiency on the leukocytes, we crossbred DsRed+ (provided by Dr. Tagawa of the Tokyo Institute of Technology) with RAMP2+/- mice to generate a DsRed+RAMP2+/- mouse line. We found that both peripheral leukocytes (Fig. 5d, e) and peritoneal macrophages (Fig. 5f, g) collected from DsRed+RAMP2+/- attached more readily to LSECs than did WT. Phagocytosis of macrophages was not different between WT and RAMP2+/- (Fig. 5h, i), however, expression of the inflammation markers, including monocyte chemotactic protein-1 (MCP-1), interleukin-6 (IL-6), and IL-1β were upregulated in RAMP2+/- macrophages (Fig. 5j). This suggests that RAMP2-deficiency in leukocytes, as well as in ECs, enhances inflammatory reactions, and that both likely contribute to the chronic inflammation observed in aged RAMP2+/- mice.

**RAMP2 deficiency increases oxidative stress, while RAMP2 overexpression leads to resistance to senescence under conditions of oxidative stress in ECs**
It has been reported that AM exerts antioxidant effects. In 6-month-old E-RAMP2-/- mice, we detected elevated levels of oxidative stress in the major organs. Levels of 4-hydroxy-2-nonenal (4HNE) immunostaining, which reveals the presence of peroxides of unsaturated fatty acids, were increased in E-RAMP2-/- (Fig. 5g). Consistent with this finding, expression of three NADPH-oxidase subunits (p22-phox, p47-phox and p67-phox) were elevated (Fig. 5h). Thus RAMP2-deficiency in ECs appears to exacerbate oxidative stress.

We next used EAhy926 ECs to generate EC lines stably overexpressing RAMP2. H2O2-treatment has been shown to induce EC senescence29. We found that after incubation for 24 h in the presence of H2O2 plus AM, control cells were strongly stained by SA-β-gal, but cells overexpressing RAMP2 were resistant to staining (Fig. 5i).

**Initial EC change elicited by RAMP2 deletion in drug-inducible EC-specific RAMP2 KO mice**

As mentioned, most E-RAMP2-/- mice died as neonates and analysis using adult mice was limited. Therefore, we next generated a line of drug-inducible EC-specific RAMP2 KO (DI-E-RAMP2-/-) mice (Fig. 6a). With this model, we are able to selectively delete RAMP2 gene from the ECs of adult mice through administration of tamoxifen. Interestingly, the tamoxifen-treated mice showed remarkable body weight gains with systemic edema (Fig. 6c, d). The cause of the edema was enhanced vascular permeability and plasma leakage, as evidenced by the extravascular leakage of Evans Blue dye (Fig. 6e). In a hind-limb ischemia model, which enables us to evaluate the post-natal angiogenic potency, we also found that blood flow recovery was delayed in DI-E-RAMP2-/- (Fig. 6f).

Next, we evaluated whether we could rescue the phenotype of DI-E-RAMP2-/- by vascular overexpression of RAMP2 gene. We employed the gene delivery method in the hind-limb
ischemia model by electroporation of plasmid, which could overexpress RAMP2 under the control of VE-cadherin promoter (Supplemental Fig. 1). At first, we confirmed the VE-cadherin promoter works only in vascular endothelial cells; when VE-cadherin-EGFP plasmid is electroporated at the femoral region, the signal was detected only at the isolectin-positive endothelial cells (Fig. 6g). In addition, after 24 h of electroporation of VE-cadherin-RAMP2 plasmid, we confirmed that RAMP2 expression was strongly induced in DI-E-RAMP2-/- (Fig. 6h). Actually, the RAMP2 gene delivery in DI-E-RAMP2-/- successfully restored the blood flow recovery in the hind-limb ischemia (Fig. 6i, j). Moreover, the RAMP2 gene delivery enhanced capillary formation and reduced interstitial edema in DI-E-RAMP2-/- (Fig. 6k, l).

Using the DI-E-RAMP2-/- line, we further evaluated the primary cause of the vascular failure. In electron micrographs of aortas obtained 2 weeks after RAMP2-deletion, ECs in DI-E-RAMP2-/- mice appeared deformed and were detached from the basement membrane (Fig. 6m). We speculated that the cause was cytoskeletal abnormality within the ECs. Consistent with that idea, actin polymerization in DI-E-RAMP2-/- ECs appeared in disarray, and there was a loss of actin-bundle formation under the plasma membrane (cortical actin formation) (Fig. 6n). It has been reported that small GTPases, Rac1 and RhoA, play crucial roles in the regulation of ECs’ barrier function by regulating the formation cortical actin and stress fibers30. When we analyzed the activation of Rac1 and RhoA in DI-E-RAMP2-/- ECs, we found that levels of the activated form of Rac1 (Rac1-GTP) were significantly reduced, whereas the activated form of RhoA (RhoA-GTP) was increased (Fig. 6o, p). On the other hand, AM-treatment enhanced cortical actin formation in HUVECs (Fig. 6q). Notably, the Rac1 inhibitor NSC23766 blocked the AM-induced enhancement of cortical action formation, whereas Rho-associated protein kinase (ROCK) inhibitor Y27632 had no effect (Fig. 6r). Taken together, these observations suggest the
AM-RAMP2 system regulates the Rac1-GTP/RhoA-GTP ratio and therefore cortical actin formation, and that a defect in this system will disrupt actin formation.

We also confirmed that at the chronic stage after the gene-deletion, DI-E-RAMP2-/- mice exhibited vascular damage with enhanced perivascular inflammation that was the same as that of adult E-RAMP2-/- (Fig. 7). With DI-E-RAMP2-/-, therefore, we were able to clarify both the acute and chronic effects of RAMP2 deletion in the adult, and demonstrate that RAMP2 is essential for EC viability, vascular integrity, and homeostasis.

Discussion

Our initial finding was that most E-RAMP2-/- mice die perinatally from severe systemic edema. In contrast to conventional RAMP2-/- embryos, which die at mid-gestation, E-RAMP2-/- embryos survived until later stages of development. As we and other groups reported previously, in conventional RAMP2-/- embryos, deformity was detected in both the endothelium and the smooth muscle layer in both blood and lymphatic vessels\(^25,31\). By contrast, in the E-RAMP2-/- embryos used in this study, the lesion was limited in the ECs, which likely accounts for their longer survival. Nonetheless, most E-RAMP2-/- died due to edema, reflecting the endothelial abnormality and vascular leakage. Clearly, endogenous RAMP2 is essential for EC viability and vascular integrity.

About 5% of E-RAMP2-/- mice survived to adulthood. E-RAMP2-/- adults were apparently normal when young, but showed EC deformity and reduced viability later. Interestingly, the cellular deformities in the adult animals was not limited to the endothelium, but also included the smooth muscle cell layer, which suggests the congenital EC abnormalities induced secondary postnatal disorder of other vascular component cells. E-RAMP2-/- mice also had lower blood pressures than WT mice, which seems contradictory, as AM is well known to be a
vasodilator. We speculate that the reduction in blood pressure might be due to the observed morphological changes in the vascular cells. Another possibility is that our findings are consistent with the report that although AM is a vasodilator when injected peripherally, it is a vasoconstrictor when injected intracerebrally\(^\text{32}\); that is, when endothelial RAMP2 deficiency affects both peripheral and central AM signaling, the expected central phenotype is observed.

E-RAMP2-/- adults also showed marked accumulation of inflammatory cells along the blood vessels within major organs, and the chronic vascular damage by such inflammation can accelerate vascular senescence\(^\text{28,33-35}\). We found that activation of aortic Akt and p53, as well as SA-\(\beta\)-gal staining, two indicators of vascular senescence, were enhanced in adult E-RAMP2-/- mice, and that similar vasculitic lesions were present in aged conventional RAMP2+/+ mice. The vascular lesions in E-RAMP2-/- mice led to organ damage, which included liver cirrhosis, cardiac enlargement with fibrosis, hydronephrosis, polycystic changes in the kidney and glomerulosclerosis. Taken together, these findings suggest that endothelial RAMP2 deficiency is the primary cause of subsequent vascular inflammation, accelerated senescence, fibrosis, and chronic organ dysfunction.

Unlike virus-related liver cirrhosis, in which the lesion starts within the hepatocytes, the cirrhotic changes seen in E-RAMP2-/- mice were caused by severe fibrosis that started in the vasculature. This is consistent with idea that vascular damage is the primary cause of this cirrhosis. In the E-RAMP2-/- kidney, both glomerular and interstitial changes were observed, along with epithelial cell damage (podocyte fusion) associated with glomerular EC deformity. And in the heart, cardiac enlargement and fibrosis led to heart failure. Thus loss of EC viability and vascular integrity accelerated vascular senescence and exacerbated interstitial lesions, making it the primary cause of chronic organ dysfunction.
For this study, we also generated EC-specific AM KO mice (E-AM-/-). In contrast to E-RAMP2-/- neonates, the E-AM-/- neonates appeared healthy. It seems the phenotype caused by knocking out the ligand during development is much milder than the one caused by knocking out the receptor. This is likely because AM is secreted from cells other than ECs, and circulating and/or paracrine AM from these other sources compensate for the endothelial AM deficiency during development. In adults, however, the chronic reduction in AM signaling led to vascular inflammation and chronic organ damage in E-AM-/- mice, suggesting that aging is another important factor contributing to the emergence of the pathophysiological phenotype.

To better understand the mechanism by which endothelial RAMP2 deficiency could promote the aforementioned pathological features, we first characterized the inflammatory response triggered in RAMP2-deficient ECs. We recently reported that the expression of adhesion markers was downregulated by AM\textsuperscript{36}. In the absence of AM signaling in RAMP2-deficient ECs in the present study, these inflammatory adhesion molecules were upregulated, facilitating the attachment of macrophages. This inflammatory response, along with the partial detachment of ECs from the basement membrane, likely accelerates the attachment, transmigration and accumulation of inflammatory cells within the vascular wall.

We also found high levels of oxidative stress, including elevation of NADPH-oxidase levels, within the major organs of E-RAMP2-/- mice. It has been reported that AM exerts strong antioxidant effects\textsuperscript{9}, and in the present study AM protected ECs stably overexpressing RAMP2 from cellular senescence induced by oxidative stress. This suggests that AM-RAMP2 signaling suppresses oxidative stress, and that endothelial RAMP2 may be an effective target through which to regulate oxidative stress.

Because most E-RAMP2-/- mice die as neonates, analysis of adult animals has been
limited. To overcome this limitation, we generated a drug-inducible EC-specific RAMP2 KO mouse (DI-E-RAMP2/-). In this model, we can induce RAMP2 gene deletion in adults on demand and then analyze the changes induced by RAMP2 deletion. Interestingly, DI-E-RAMP2/- mice exhibit marked systemic edema caused by increased vascular permeability. It is widely recognized that an increase in intracellular cyclic adenosine monophosphate (cAMP) within ECs strengthens their barrier function and reduces endothelial permeability, both in vitro and in vivo\(^3\),\(^3\). It is therefore not surprising that cAMP-elevating G protein-coupled receptor agonists, such as AM, prostacyclin and prostaglandin E2, reduce endothelial hyperpermeability induced by inflammatory stimuli\(^3\). In addition, small GTPases, especially Rac1 and RhoA, play crucial roles in the regulation of EC barrier function by regulating the formation of cortical actin and stress fibers, respectively\(^3\). Elevation of cAMP leads to Rac1 activation, which in turn strengthens endothelial barrier function by enhancing cortical actin ring formation\(^3\). We found that in DI-E-RAMP2/- ECs, actin polymerization was disrupted and Rac1 activation was reduced. By contrast, AM-stimulated ECs showed cortical actin formation, which disappeared upon treatment with a Rac1 inhibitor. Taken together, these findings suggest that in DI-E-RAMP2/- ECs, downregulation of cAMP production and Rac1 activation reduces cortical actin formation, diminishing endothelial barrier function.

The relationship between the AM receptor and human disease has also been reported. A human SNP study described the relationship between CLR with essential hypertension\(^4\). A human study also revealed the relationship between RAMP1 SNPs with the incidence of cerebral infarction \(^4\). Some human mutations of RAMP3 gene have also been reported \(^4\). On the other hand, there have been no reports on human RAMP2 gene mutation. As we have shown, RAMP2-deletion can cause embryonic lethality at mid-gestation, and this may be one reason that
we cannot see human congenital disease. In this context, the endothelial cell-specific conditional knockout mice model in this study should be recognized as a model of vascular failure and relevant organ dysfunction and should not be recognized as a specific congenital disease model.

In this study, we were able to clarify both the acute and chronic effects of RAMP2 deletion in the adult, and to demonstrate that the AM-RAMP2 system is essential for vascular integrity and organ homeostasis. The illustration in Figure 8 summarizes the series of phenomena stemming from RAMP2 deletion. Early after RAMP2 gene deletion, ECs show morphological changes due to actin filament abnormality (disappearance of the cortical actin ring and disarray of actin polymerization). The resultant EC deformation causes detachment, barrier dysfunction, enhanced vascular permeability and edema, which in turn promote the attachment and infiltration of inflammatory cells. Chronic vascular inflammation induces vascular damage and accelerated senescence, and enhances oxidative stress and organ fibrosis. Finally, the accumulated disorders cause chronic organ dysfunction with aging. The results obtained using these models demonstrate that vascular EC integrity is essential for organ homeostasis. We also suggest that RAMP2, via which we could modulate vascular integrity, is a potentially useful therapeutic target for the treatment of chronic organ dysfunction.

Acknowledgements: The authors thank Dr. Sandra Hervás for her assistance with cell sorting.

Funding Sources: This study was supported by the Funding Program for Next Generation World-Leading Researchers (NEXT Program) from the Cabinet Office, Government of Japan and Innovation Grant from Spain’s Ministry of Science (SAF2009-13240-C02-01).

Conflict of Interest Disclosures: None
References:


**Figure Legends:**

**Figure 1.** E-RAMP2-/- mice showed systemic edema and high perinatal mortality, while surviving adult mice showed vascular abnormalities. (a) Strategy of conditional gene targeting of mouse RAMP2. A RAMP2 floxed mouse was crossbred with a VE-cadherin promoter-driven Cre recombinase-expressing mouse (lower panel) to generate vascular endothelial cell-specific RAMP2-/- mice (E-RAMP2-/-). (b) E-RAMP2-/- survival curve. E-RAMP2-/- embryos generally survived at least until the perinatal period. (c, d) E19.5 E-RAMP2-/- embryos showed systemic edema (e) with increased body weight (d). n = 11 in both Control and E-RAMP2-/-, Bars are means ± SEM. **p<0.01 (e-g) H&E staining of the intestine (e), lung (f) and liver (g) from E19.5 RAMP2*^floxed/floxed* mice (Cont) and E-RAMP2-/- (h-j) Electron micrographs of aortas from E19.5 Cont and E-RAMP2-/- (h-j) Electron micrographs of aortas from E19.5 Cont and E-RAMP2-/-. (h-j) Electron micrographs of aortas from E19.5 Cont and E-RAMP2-/-. (k, l) H&E staining of aortas from adult Cont and E-RAMP2-/-. Scale bars = 300µm (k), 50 µm (l). (m) Blood pressures in adult Control and E-RAMP2-/- mice. SBP systolic blood pressure; DBP diastolic blood pressures. n = 12 in Control and n=9 in E-RAMP2-/-. Bars are means ± SEM. **p<0.01. (n, o) Electron micrographs of adult aortas. E-RAMP2-/-
aortas showed detachment of ECs (n) and a disordered smooth muscle layer (o). Scale bars = 5 μm.

(p) ex vivo analysis of angiogenesis using adult aortic ring specimens after 14 days of culture in collagen gel.

**Figure 2.** Adult E-RAMP2-/− mice showed spontaneous vasculitic lesions and premature vascular senescence. (a-e) Severe infiltration and accumulation of inflammatory cells in blood vessels within major organs of adult E-RAMP2-/−. Histology (H&E staining) of liver (a), kidney (b) and lung (c) from Cont and E-RAMP2-/− is shown. E-RAMP2-/− showed spontaneous vascular lesions, beginning at about 6 months of age. Immunohistological analysis revealed these cells to be CD3+ (d) or F4/80+ (e), which is indicative of T cells and macrophages, respectively. (f) Quantitative real-time PCR analysis of kidney showing upregulation of inflammatory cytokines and macrophage markers in E-RAMP2-/−. n = 6 in Control and n = 3 in E-RAMP2-/−. Bars are means. *p<0.05. (g-i) Vasculitic lesions similar to those in E-RAMP2-/− were confirmed in aged (2-year-old) conventional RAMP2+/− (Scale bars = 100μm). (j, k) SA-β-gal staining of aorta. In aged mice (2 year-old), aortic SA-β-gal staining was more intense in RAMP2+/− than wild-type mice (WT) (j). The SA-β-gal positivity was detected much earlier (6 months of age) in E-RAMP2-/− than Cont (k). (l) Western blot analysis of aortas from RAMP2+/− and WT. p53 and Akt were more activated in aged (2-year-old) RAMP2+/− than WT (right panel), but not in younger (9-week-old) mice (left panel). However, expression of ICAM-1 and VCAM-1 was already upregulated in young RAMP2+/− (left panel).

**Figure 3.** E-RAMP2-/− mice showed fibrosis starting around vessels and progression of organ damage. (a) Livers appeared cirrhotic in 6-month-old E-RAMP2-/− mice. (b, c) Silver (b) and
sirius red (c) staining of E-RAMP2-/ livers showed severe fibrosis along the vasculature. Scale bars = 100μm. (d, e) Cardiac enlargement with interstitial fibrosis in 6-month-old E-RAMP2-/ mice. Gross appearance (d) and histology (Masson trichrome staining) of the heart (e) are shown. Scale bars = 200μm. (f) Quantitative real-time PCR analysis of hearts from Cont and E-RAMP2-/ mice. Expression of heart failure and fibrosis markers was upregulated in E-RAMP2-/ mice. n = 4 in both Control and E-RAMP2-/-. Bars are means ± SEM. **p<0.01. (g-i) Histological abnormalities in the kidneys of E-RAMP2-/-. (g) Section of whole E-RAMP2-/ kidney showing an enlarged renal pelvis; (h) Section of renal cortex showing polycystic changes. Scale bars = 100 μm. (i) Enlargement of the glomerular area in a E-RAMP2-/ kidney. Scale bars = 50 μm. n = 4 in both Control and E-RAMP2-/-. Bars are means ± SEM. **p<0.01. (j) Electron micrographs of glomeruli from Cont and E-RAMP2-/ kidneys. In the E-RAMP2-/ kidney, fusion of podocytes (upper panel) and detachment of ECs from the basement membrane (lower panel) are apparent (arrows). Scale bars = 1 μm. (k) Quantitative real-time PCR analysis of Cont and E-RAMP2-/ kidneys. Fibrotic markers were upregulated in E-RAMP2-/ kidneys. n = 4 in both Control and E-RAMP2-/-. Bars are means ± SEM. *p<0.05.

**Figure 4.** Adult E-AM/- mice showed spontaneous vasculitic lesions and glomerulosclerosis. (a, b) Vasculitic lesion showing accumulation of inflammatory cells in an aged (6-month-old) EC-specific AM/- (E-AM/-) mouse. Histology (H&E staining) of the liver from a Cont and E-AM/- mouse is shown (a). The accumulated cells are CD3+ T cells (b). Scale bars = 100μm. (c, d) Glomerulosclerotic changes in E-AM/- kidneys. Sirius red staining shows excessive collagen deposits among mesangial cells and in a Bowman’s capsule from an E-AM/- kidney (c).
Glomerular area was significantly enlarged in the E-AM-/- kidney (d). Scale bars = 50 μm. n = 10 in both Control and E-AM-/-. Bars are means ± SEM. **p<0.01.

(e) Quantitative real-time PCR analysis showing upregulation of inflammatory and fibrosis markers in E-AM-/- kidneys. n = 10 in both Control and E-AM-/-. Bars are means ± SEM.

Figure 5. RAMP2 deficiency enhances EC inflammatory reactions and oxidative stress. (a, b) Macrophage adhesion to ECs. Macrophage adhesion to LSECs cultured from WT or RAMP2+/- mice was analyzed. RAMP2+/- LSECs showed greater adhesion, which was indicated by a higher cell count. Scale bars = 100 μm. n = 8 in both WT and RAMP2+/- . Bars are means ± SEM.

* p<0.05. (c) Quantitative real-time PCR analysis of LSECs. n = 6 in both WT and RAMP2+/- . Bars are means ± SEM. ** p<0.01, * p<0.05. (d-g) To analyze the effect of RAMP2 deficiency on peripheral leukocytes and macrophages, buffy coat and peritoneal macrophages collected from DsRed/RAMP2+/- or DsRed/RAMP2+/- mice were cultured with LSECs from WT mice. Attached DsRed+ cells were photographed (d, f) and counted (e, g). RAMP2+/- leukocytes (d, e) and macrophages (f, g) showed greater attachment than those of WT. Scale bars – 100 μm. n = 8 in both WT and RAMP2+/- . Bars are means ± SEM. ** p<0.01, *** p<0.001. (h, i) Effect of RAMP2 deficiency on phagocytosis of macrophages. Photographs (h) and dot plot (i) representing phagocytosis of FITC-labeled particles by macrophages (circles in the dot plot) are shown.

Phagocytosis was not different between WT and RAMP2+/- . Scale bars = 50 μm. (j) Quantitative real-time PCR analysis of macrophages from RAMP2+/- and WT mice. n = 8 in both WT and RAMP2+/- . Bars are means ± SEM. ** p<0.01. (k) Enhanced oxidative stress in E-RAMP2/- mice. Immunostaining of unsaturated fatty acid peroxides was enhanced in E-RAMP2/- liver and kidney. Particularly strong staining was detected around central veins in the liver and renal tubules.
in the kidney. Scale bars = 200 μm. (l) Quantitative real-time PCR analysis of NADPH-oxidase subunits in the kidney. n = 6 in Control and n = 3 in E-RAMP2-/- . Bars are means. *p<0.05. (m) Suppression of oxidative stress-induced cellular senescence in ECs stably overexpressing (O/E) RAMP2. After 24 h of treatment with H2O2 (200 μmol/L) and AM (10^{-7} M), SA-β-gal positivity was detected in the control EAhy926 ECs, but not RAMP2-overexpressing cells. Scale bars = 100 μm.

Figure 6. Analysis of vascular endothelial RAMP2 deficiency in adults using drug-inducible EC-specific RAMP2 KO mice (DI-E-RAMP2-/-). (a) Strategy of drug-inducible conditional gene targeting of mouse RAMP2. A RAMP2 floxed mouse was crossbred with a VE-cadherin promoter-driven, tamoxifen-inducible Cre recombinase transgenic mouse to generate drug-inducible EC-specific RAMP2-/- mice (DI-E-RAMP2-/-). (b) Quantitative real-time PCR analysis of LSECs cultured primarily from DI-E-RAMP2-/- mice (2 weeks after the treatment with tamoxifen or corn oil). In tamoxifen-treated mice, endothelial RAMP2 expression was reduced to about 20% of control, while AM expression was upregulated to 200% of control. n – 8 in both Control and DI-E-RAMP2-/- . Bars are means ± SEM. **p<0.01. (c, d) Appearance and body weight changes after induction of RAMP2 deletion in adult (8-week-old) mice. After induction of RAMP2 deletion, the mice showed facial and systemic edema (e) and marked body weight gain (d). n = 8 in Control and n = 12 in DI-E-RAMP2-/- . Bars are means ± SEM. *p<0.05, **p<0.01. (e) Photographs (back skin, ear, hindlimb) showing the extravascular leakage of Evans Blue dye after its intravenous injection. DI-E-RAMP2-/- mice showed much greater vascular leakage. (f) Blood flow ratio obtained from laser Doppler analysis in unilateral hind-limb ischemia model of control and DI-E-RAMP2-/- mice. n = 7 in both Control and DI-E-RAMP2-/- . Bars are means ±
SEM. *p<0.05, **p<0.01. (g) Section of the femoral muscle electroporated with VE-cadherin-EGFP plasmid. EGFP-signaling and red fluorescence of isolectin-staining are shown. Scale bars = 50 μm (h) Quantitative real-time PCR analysis of RAMP2-expression in the femoral muscle of control and DI-E-RAMP2-/- mice electroporated with the indicated plasmids. n = 4 in each group. Bars are means ± SEM. *p<0.05 vs. control mice + control plasmid, †† p < 0.01 vs. DI-E-RAMP2-/- mice + control plasmid. (i-l) Unilateral hind-limb ischemia model used to evaluate angiogenic potency by electroporation of the indicated plasmids. (i) Laser Doppler imaging. (j) Blood flow ratio of ischemic limb/control limb. n = 7-8 in each group. Bars are means ± SEM. **p<0.01 vs. control mice + control plasmid, †† p < 0.01 vs. DI-E-RAMP2-/- mice + control plasmid. (k) Sections showing the neovascularization (isolectin staining) and interstitial edema (H&E staining) of the femoral muscle. Scale bars = 100 μm. (l) Bar graphs showing the capillary density and interstitial edema area quantified from each section. n = 4-5 in each group. Bars are means ± SEM. *p<0.05, **p<0.01 vs. control mice + control plasmid, †† p < 0.01 vs. DI-E-RAMP2-/- mice + control plasmid. (m) Electron micrographs of aortas obtained 2 weeks after gene deletion. DI-E-RAMP2-/- vessels showed detachment of ECs (arrows). Scale bars = 1 μm. (n) LSECs cultured from control and DI-E-RAMP2-/- mice were stained with rhodamine-phalloidin to visualize F-actin. Scale bars = 50 μm. (o) Western blots of Rac1-GTP, total Rac1, Rho-GTP and total Rho from ECs. (p) Densitometric analysis of the Western blots. n = 3 in both Control and DI-E-RAMP2-/- . Data are show as the relative ratio to Control. Bars are means. *p<0.05. (q) Effect of AM on HUVECs. HUVECs were stained with rhodamine-phalloidin to visualize F-actin. The cell in the right panel was stimulated for 30 min with 10^-7 M AM. Scale bars = 50 μm. (r) Effect of AM with and without a Rho kinase or Rac1 inhibitor on mouse LSECs. Mouse LSECs were treated for 180 min with 10^-7 M AM with and
without $10^{-4}$ M NSC23766 (Rac1 inhibitor) or $10^{-5}$ M Y27632 (ROCK inhibitor). Scale bars = 50 μm.

**Figure 7.** Chronic RAMP2-deficiency causes vascular damage and enhanced perivascular inflammation in DI-E-RAMP2-/- mice. (a-c) Histology (H&E staining) of liver (a), kidney (b) and lung (c) collected at the chronic stage. Severe infiltration and accumulation of inflammatory cells around blood vessels was detected in DI-E-RAMP2-/- mice but not in the control mice (Cont). Scale bars =100 μm.

**Figure 8.** Regulation of EC integrity by the AM-RAMP2 system and its disruption in RAMP2 KO mice. The AM-RAMP2 system regulates cortical actin formation in ECs and barrier integrity through upregulation of cAMP and activation of Rac1. RAMP2 deletion disrupts actin polymerization in ECs and causes barrier dysfunction, which in turn leads to enhanced vascular permeability, infiltration of inflammatory cells, vascular damage and accelerated vascular senescence. These accumulated disorders cause chronic organ dysfunction with aging.
Figure 1

(a) Schematic diagram of the RAMP2 targeted locus. The diagram shows the targeted insertion of the LoxP site at two specific locations (1 and 4) and the promoter region (Cdh5) and the reporter gene (VE-cad-Cre) used for induction of RAMP2 expression.

(b) Survival rate graph showing the perinatal period with a peak survival rate at 100%.

(c) Images of control (Cont) and E-RAMP2-/- mice, highlighting the differences in body size.

(d) Bar graph showing body weight (g) comparison between control (Cont) and E-RAMP2-/- mice. The graph indicates a significant difference (**) in body weight.

(e) Comparison of control (Cont) and E-RAMP2-/- tissue sections from the intestine.

(f) Comparison of control (Cont) and E-RAMP2-/- tissue sections from the lung.

(g) Comparison of control (Cont) and E-RAMP2-/- tissue sections from the liver.

(h) Images showing the histological differences between control (Cont) and E-RAMP2-/- mice.

(i) Magnified images of lung tissue sections, highlighting the differences in structure.

(j) Magnified images of liver tissue sections, showing the differences in tissue architecture.

Figure 1
Figure 1, cont’d
Figure 2
Figure 2, cont’d
Figure 3

(a) Cont vs E-RAMP2-/-

(b) Silver stain

(c) Sirius red stain

(d) Cont vs E-RAMP2-/-

(e) Cont vs E-RAMP2-/-

(f) Relative gene expression

- ANP
- BNP
- TGF-β
Figure 3, cont’d
Figure 4
Figure 5
Figure 5, cont’d
Figure 5, cont’d
Figure 6
Figure 6, cont’d
Figure 6, cont’d
Figure 7
Endothelial cell

KO

Rac1\downarrow
Rho\uparrow

Cortical actin ring\downarrow
Disarray of actin polymerization

Barrier dysfunction

Adrenomedullin
-RAMP2 system

cAMP\uparrow

Rac1\uparrow
Rho\downarrow

Cortical actin\uparrow
Stress fibers\downarrow

Barrier integrity

Vascular permeability\uparrow
Edema\uparrow

Vascular damage & accelerated senescence

Aging

Chronic organ dysfunction

Figure 8
Vascular Endothelial Adrenomedullin-RAMP2 System is Essential for Vascular Integrity and Organ Homeostasis

Teruhide Koyama, Laura Ochoa-Callejero, Takayuki Sakurai, Akiko Kamiyoshi, Yuka Ichikawa-Shindo, Nobuyoshi Inuma, Takuma Arai, Takahiro Yoshizawa, Yasuhiro Iesato, Yang Lei, Ryuichi Uetake, Ayano Okimura, Akihiro Yamauchi, Megumu Tanaka, Kyoko Igarashi, Yuichi Toriyama, Hisaka Kawate, Ralf H. Adams, Hayato Kawakami, Naoki Mochizuki, Alfredo Martínez and Takayuki Shindo

_Circulation_. published online January 25, 2013;

_Circulation_ is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2013 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circ.ahajournals.org/content/early/2013/01/25/CIRCULATIONAHA.112.000756

Data Supplement (unedited) at:
http://circ.ahajournals.org/content/suppl/2013/01/25/CIRCULATIONAHA.112.000756.DC1

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in _Circulation_ can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to _Circulation_ is online at:
http://circ.ahajournals.org/subscriptions/
Supplementary methods

Mouse models

RAMP2-/- mice were generated in our group. Vascular endothelial cadherin (VE-cadherin) Cre transgenic mice and mice expressing tamoxifen-inducible Cre-recombinase (Cre-ERT2) under the regulation of VE-cadherin promoter were crossed with floxed RAMP2 mice (RAMP2flox/flox) to create vascular EC-specific RAMP2 conditional KO mice (E-RAMP2-/-) (Fig. 1a) and tamoxifen drug-inducible (DI) vascular EC-specific RAMP2 KO mice (DI-E-RAMP2-/-) (Fig. 6a), respectively. Tamoxifen (Sigma) was dissolved in corn oil (Sigma) to a concentration of 10 mg/ml, after which 1 mg was intraperitoneally injected into male mice (8 weeks old) daily for 5 days. Little toxicity was observed following the injection into wild-type (WT), RAMP2flox/flox, or Cre-ERT2 mice.

Mice with conditional KO of AM in ECs (E-AM-/-) were created by crossing floxed AM mice with transgenic mice expressing Cre recombinase under the control of the VE-cadherin promoter (Stock Number 006137, The Jackson Laboratory). All animal handling procedures were in accordance with a protocol approved by the Ethics Committee of Shinshu University and the CIBIR.

Cell isolation and culture

Primary adult mouse liver sinusoidal endothelial cells (LSECs) were isolated using a two-step collagenase perfusion and centrifugation method. Inhibitors of Rac1 and ROCK used are NSC 23766 (SANTA CRUZ) and Y27632 (Wako).
A RAMP2 overexpressing (RAMP2 O/E) cell line was created using EAhY926 ECs (provided by Dr. Edgell of the University of North Carolina) as described previously. RAMP2 O/E and EAhY926 cells were treated with 200 µmol/L hydrogen peroxide (H$_2$O$_2$) (Sigma) for 24h and with 10$^{-7}$ M AM (provided Dr. Kangawa of the National Cardiovascular Center Research Institute).

Buffy coat (combined leukocyte and platelet fractions) was isolated from blood samples collected from mice as described previously. Peritoneal macrophages were obtained from mice by injecting 2 ml of 5% thioglycolate (DIFCO) intraperitoneally. The elicited macrophages were harvested by peritoneal lavage 4 days after thioglycolate administration.

Human umbilical vein endothelial cells (HUVECs) were purchased from Kurabo (Kurashiki, Japan).

**Histology**

Each organ was fixed overnight in methanol or 4% paraformaldehyde, embedded in paraffin, and cut into 4-µm-thick sections for histological examination. The sections were then deparaffinized for hematoxylin/eosin, Masson’s trichrome, Sirius red and silver staining, and for immunohistochemistry. The antibodies used were rabbit anti-CD3 (A 0452, DAKO, Carpinteria, CA), rat anti-mouse F4/80 (Serotec) and rat anti-mouse 4-hydroxy-2-nonenal (4HNE) (NOF corporation). Whole aorta was stained for senescence-associated β-gal (SA-β-gal) as described previously.
Transmission electron microscopy

Specimens were fixed in 2.5% glutaraldehyde (pH7.2), embedded in epoxy resin (Epok) 812 (Oken shoji Co.), cut into ultrathin sections, double-stained with uranyl acetate and lead citrate, and examined in an electron microscope.

Western blot analysis

Western blot analysis was carried out using protein extracts from aorta. The lysates were subjected to SDS-PAGE, transferred to nitrocellulose membranes and probed with anti-Akt, anti-phospho-Akt (p-Akt) (Cell Signaling Technology), anti-ICAM-1, anti-VCAM-1, anti-p53 and anti-β-actin (Santa Cruz Biotechnology) antibodies. The blots were then developed using a SNAP i.d. system (MILLIPORE, Billerica, MA).

Quantitative real-time RT-PCR analysis

Quantitative real-time RT-PCR was carried out using an Applied Biosystems 7300 real-time PCR System with SYBR green (Toyobo, Japan) or Realtime PCR Master Mix (Toyobo) and TaqMan probes (MBL). The primers and probes used are listed in Supplementary Table. 3. Values were normalized to mouse GAPDH (Pre-Developed TaqMan assay reagents, Applied Biosystems).

Macrophage/leukocyte and LSEC adhesion assay

For macrophage and LSEC adhesion assay, LSECs from WT and RAMP2+/- mice were seeded into 35-mm plates and incubated for 24 h, after which macrophages (10^5
cells) extracted from GFP+, DsRed/RAMP2+- or DsRed/RAMP2+/+ mice were added. For leukocyte and LSEC adhesion assay, Buffy coat (10⁵ cells) extracted from DsRed/RAMP2+/- or DsRed/RAMP2+/+ mice was added. After incubation for an additional 3 h, the cells were washed three times with PBS, and the macrophages were counted in 10 random fields/dish.

**Analysis of phagocytosis of macrophages**

In vitro phagocytic uptake studies were carried out using fluorescent microparticles (Polysciences, Inc.) in elicited macrophages according to the manufacturer’s protocol. In brief, 2 × 10⁵ of thioglycollate-elicited macrophages were mixed with 1 × 10⁶ opsonized fluorescent microparticles overnight and then washed with ice-cold PBS. Thereafter, phagocytosis was examined using a fluorescence microscope and analyzed by flow cytometry.

**Aortic ring assay**

The thoracic aorta was dissected from the posterior mediastinum and placed in serum-free EBM-2 endothelial basal medium (Cambrex). The vessel was then cut into 1-mm-long rings, which were subjected to 8 consecutive washes with serum-free EBM-2. The aortic rings were then embedded in thick collagen gel (Cellmatrix Type I-A; Nitta Gelatin) and cultured for 14 days, with or without recombinant hVEGF (50 ng/ml; R&D Systems).
Rac1 and Rho pull-down assays

The activation of Rac1 was assessed using a Rac1 activation kit (Enzo Life Science) according to the manufacturer’s instructions. The activation of Rho was assessed using a Rho Activation Assay kit (MILLIPORE) according to the manufacturer’s instructions.

Vascular permeability assay

Mice were anesthetized, after which 100 µl of 1% Evans Blue (Wako) were injected intravenously. Thirty minutes after injection, extravasation of the Evans Blue dye was visualized in back skin, ear and hindlimb.

Unilateral hind-limb ischemia model

DI-E-RAMP2-/- mice and their controls were subjected to experimentally induced hind-limb ischemia produced by unilateral occlusion of the femoral artery, as described previously. Laser Doppler perfusion imaging (LDPI) (Omegazone, Omegawave, Inc., Tokyo, Japan) was used to evaluate the blood flow. The calculated perfusion was expressed as a ratio of the left (ischemic) to the right (normal) limb. Capillary density in the limbs at 11 days after surgery was assessed by fluorescence staining with isolectin GS-IB4 (Invitrogen). Interstitial edema area was calculated as the interval spaces within the muscles.

Construction of Plasmid DNA

Expression vector cassette, which contains a 2.53 kb 5′-flanking region of mouse
vascular endothelial (VE)-cadherin (pVE-Cont), was kindly provided by Dr. Huber (Supplementary Figure 1a). To generate RAMP2-gene delivery plasmid (pVE-mRAMP2), mouse RAMP2 ORF (mRAMP2) (0.57 kb) was inserted at the EcoRI site of the pVE-Cont (Supplementary Figure 1b). In this plasmid, the transcript of mRAMP2 cDNA is generated under the control of the VE-cadherin promoter. EGFP expression vector (pVE-EGFP) was also generated to confirm proper gene delivery (Supplementary Figure 1c).

**In vivo electroporation-mediated gene transfer**

Following the hind-limb ischemia operation, the mice were electroporated plasmids into their femoral regions. A volume of 20 µL of plasmid (2 mg/ml) was injected into the femoral muscle with a 29-gauge needle. Immediately after intramuscular injection of the plasmid, the muscle was held by an electrode and *in vivo* electroporation was performed with a pulse generator (BTX T820, BTX, San Diego, CA, USA, CUY 560-5, NEPA GENE, Chiba, Japan). The voltage, pulse length, and number of pulses of the electroporation were 100 V, 50 ms, and 3 pulses followed by 3 inverse pulses, respectively.

**Statistical analysis**

Statistical analysis was performed using Student's t test for data following an expected normal distribution and Mann Whitney U test where data was not normally distributed. Values are expressed as means ± SEM in Student's t test and means in Mann
Whitney U test. Values of *p<0.05 or **p<0.01 were considered significant.
Supplementary figure legends

Supplementary Table. 1
Result of the genotyping of pups from RAMP2^{flox/flox} Cre+-/ intercrosses

Real numbers and estimated pup numbers of each genotype are shown. Estimated pup ratio of RAMP2^{flox/flox} Cre+ to RAMP2^{flox/flox} Cre- = 3:1 for the RAMP2^{flox/flox} Cre+/- intercrosses; therefore, the estimated number of RAMP2^{flox/flox} Cre+ = 207 (3 fold higher than the 69 of RAMP2^{flox/flox} Cre-). The estimated survival rate of RAMP2^{flox/flox} Cre+ pups (EC-specific RAMP2/- (E-RAMP2/-)) is calculated as 7/207 x 100 = 3.4%.

Supplementary Table. 2
Result of the genotyping of pups from AM^{flox/flox} Cre+-/ with AM^{flox/flox} Cre-/- intercrosses

Unlike E-RAMP2/- pups, EC-specific adrenomedullin/- (E-AM/-) pups were all healthy and presented no overtly pathophysiological phenotype. In AM^{flox/flox} Cre+/- and Cre-/- intercrosses, the numbers of Cre+ and Cre- pups were nearly equal, which indicates that the absence of AM in the ECs does not influence birth rate.

Supplementary Table. 3
Primers and probes for quantitative real-time RT-PCR analysis

Supplementary Figure. 1
Plasmid DNA used for the gene delivery experiment

(a) Control plasmid, which contains a 2.53 kb 5’-flanking region of mouse vascular endothelial (VE)-cadherin (pVE-Cont).
(b) RAMP2-gene delivery plasmid (pVE-mRAMP2), in which mouse RAMP2 ORF (mRAMP2) (0.57 kb) was inserted at the EcoRI site of the pVE-Cont.

(c) EGFP expression plasmid (pVE-EGFP), in which EGFP ORF (0.74 kb) was inserted at the EcoRI site of the pVE-Cont.
References


Supplementary Table.1

<table>
<thead>
<tr>
<th></th>
<th>RAMP2^{flox/flow} Cre+</th>
<th>RAMP2^{flox/flow} Cre-</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of pups</td>
<td>7</td>
<td>69</td>
<td>76</td>
</tr>
<tr>
<td>Estimated number of pups</td>
<td>207</td>
<td>69</td>
<td>276</td>
</tr>
<tr>
<td>Estimated survival rates</td>
<td>3.4%</td>
<td>100%</td>
<td></td>
</tr>
</tbody>
</table>
Genotype of adult mouse from AM^flox/flow Cre+/- and Cre-/- male and female mouse intercrosses

<table>
<thead>
<tr>
<th></th>
<th>male</th>
<th>Female</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of pups Cre+</td>
<td>44</td>
<td>36</td>
<td>80</td>
</tr>
<tr>
<td>Number of pups Cre-</td>
<td>50</td>
<td>33</td>
<td>83</td>
</tr>
</tbody>
</table>

Supplementary Table.2
<table>
<thead>
<tr>
<th>Protein</th>
<th>Forward</th>
<th>Reverse</th>
</tr>
</thead>
<tbody>
<tr>
<td>TNFα</td>
<td>ACGGCATGGATCTCAAAGAC</td>
<td>AGATAGCAAATCGGCTGACG</td>
</tr>
<tr>
<td>MCP-1</td>
<td>GCAGTTAACGGCCCACTCA</td>
<td>CCTACTCATGGATCATCTTGT</td>
</tr>
<tr>
<td>F4/80</td>
<td>GATGAATCCCTGTTGTTTG</td>
<td>ATCACGATGTCAGGGAGAGAACA</td>
</tr>
<tr>
<td>ANP</td>
<td>TCCATCACCTGGGCTTCTT</td>
<td>AGGATTGTTCCCAATATGGCC</td>
</tr>
<tr>
<td>BNP</td>
<td>TCCAGAGCAATTCAAGATGCA</td>
<td>GTCTTTTCATGCGCCCTT</td>
</tr>
<tr>
<td>Collagen α1</td>
<td>ATACGCCGCAGGATGACC</td>
<td>TCAGCTGGATAGCGACAT</td>
</tr>
<tr>
<td>AM</td>
<td>CTAACGGCCAGGATGACC</td>
<td>GTAAGCGCCAGGATGACC</td>
</tr>
<tr>
<td>RAMP2</td>
<td>GCAGCCCCACCTCTCTGATC</td>
<td>AACGGATGGAGCGAGATGG</td>
</tr>
<tr>
<td>eNOS</td>
<td>AGGCACGTCTGACGCAG</td>
<td>TTCTCCCAGTGGTTCCCAGG</td>
</tr>
<tr>
<td>ICAM-1</td>
<td>CCTAAAATGGACCTGAGCAG</td>
<td>TTTGACAGACTTCAACCAC</td>
</tr>
<tr>
<td>VCAM-1</td>
<td>CCCTGAAATACAAAACGATTC</td>
<td>CAGCCCGTAGAGTCGGAG</td>
</tr>
<tr>
<td>P22 phox</td>
<td>GCCCATGGCCAGTGGTAC</td>
<td>GTCAATGGGACTCCAG</td>
</tr>
<tr>
<td>P47 phox</td>
<td>ATCCATCTGGAGCCCTTGA</td>
<td>CACCTGGCTGTGGGATCC</td>
</tr>
<tr>
<td>P67 phox</td>
<td>CAGACCCAAAACCACAGAAA</td>
<td>AAAGCCAAACACATACCGG</td>
</tr>
<tr>
<td>IL-6</td>
<td>CCCTACCACTGCTCTCC</td>
<td>TGAAATGGGATGCTTGC</td>
</tr>
<tr>
<td>IL-1β</td>
<td>TCCCTACGGCACTACGAAC</td>
<td>TCTCTTGGCTGTCTCTT</td>
</tr>
<tr>
<td>TGF-β</td>
<td>CCGAAGCAGGACTACTATGC</td>
<td>TAGATGGCGTTGGCGG</td>
</tr>
</tbody>
</table>

**Supplementary Table 3**
Supplementary Figure.1