Adipose Tissue ATP Binding Cassette Transporter A1 Contributes to High-Density Lipoprotein Biogenesis In Vivo

Soonkyu Chung, PhD; Janet K. Sawyer, MS; Abraham K. Gebre, MS; Nobuyo Maeda, PhD; John S. Parks, PhD

Background—Adipose tissue (AT) is the body’s largest free cholesterol reservoir and abundantly expresses ATP binding cassette transporter A1 (ABCA1), a key cholesterol transporter for high-density lipoprotein (HDL) biogenesis. However, the extent to which AT ABCA1 expression contributes to HDL biogenesis in vivo is unknown.

Methods and Results—Adipocyte-specific ABCA1 knockout mice (ABCA1

Conclusions—We provide in vivo evidence that AT ABCA1-dependent cholesterol efflux and nascent HDL particle formation contribute to systemic HDL biogenesis and that AT ABCA1 expression plays an important role in adipocyte cholesterol homeostasis. (Circulation. 2011;124:00-00.)

Key Words: adipose tissue • adipocytes • apolipoproteins • cholesterol • lipids • lipoproteins

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vivo to plasma HDL formation. For example, macrophage ABCA1 expression is abundant, but it has no significant impact on plasma HDL levels. The other hand, hepatic and intestinal ABCA1 expression is estimated to contribute 70% to 80% and 15% to 20% of the plasma HDL pool, respectively. Intriguingly, deletion of both hepatic and intestinal ABCA1 resulted in 90% reduction in plasma HDL, whereas global ABCA1 knockout (KO) mice have <5% of normal levels of HDL cholesterol, similar to subjects with Tangier disease. These results suggest that ABCA1 expression in cells other than hepatocytes and intestinal epithelial cells would minimally affect plasma HDL formation. However, formal testing of the contribution of a cell type to the plasma HDL pool requires specific deletion of ABCA1 in that cell.

AT is a major storage pool of FC that expands during obesity and abundantly expresses ABCA1, but the role of adipocyte ABCA1 expression in FC efflux and HDL formation in vivo is unknown. AT has been proposed as being important in plasma HDL production on the basis of indirect evidence such as an inverse association between adipose cholesterol content and plasma cholesterol concentrations. A recent study provided in vivo evidence that adipocytes support the transfer of FC to plasma HDL. In that study, mouse embryonic fibroblasts (from wild-type [WT], ABCA1 KO, and scavenger receptor class B type I [SR-BI] KO mice) were differentiated into adipocytes, radiolabeled with 3H-cholesterol, and injected into the peritoneal cavity of mice. Reverse cholesterol transport of 3H-cholesterol from adipocytes to the feces depended on adipocyte expression of ABCA1 and SR-BI. However, direct in vivo evidence for the role of adipose ABCA1 in plasma HDL formation remains lacking, and the extent to which adipose ABCA1 expression contributes to plasma HDL, particularly in the obese state, has not been investigated.

The purpose of this study was to directly test the role of adipocyte ABCA1 in the formation of nascent HDL particles and its contribution to the plasma HDL pool in vivo. To accomplish this, we generated adipocyte-specific ABCA1 KO (ABCA1<sup>−/−</sup>) mice and investigated their plasma lipid and lipoprotein phenotypes during chow and high-fat (HF) diet feeding. We also determined the role of ABCA1 in adipocyte cholesterol homeostasis and nascent HDL particle formation using AT explants from these mice. Our results suggest a significant role for AT ABCA1 expression in nascent HDL particle formation and AT cholesterol homeostasis.

**Methods**

**Animals**

Adipocyte-specific ABCA1 KO mice (ABCA1<sup>−/−</sup>) were generated by crossing ABCA1<sup>floxed</sup> mice<sup>15</sup> (backcrossed into the >99% C57BL/6 background) with fatty acid binding protein 4 (apoA2) transgenic mice (Jackson Laboratories). For all experiments, 10- to 16-week-old male mice were used. To expand AT mass, mice were switched from a commercial chow diet at 8 weeks of age to a commercial HF diet (42% calories from fat, 0.2% cholesterol; TD 88137, Harlan) for an additional 8 to 16 weeks, depending on the experiment. The mice were housed in the Wake Forest University Health Sciences animal care facilities with a 12-hour light/12-hour dark cycle. All protocols and procedures were approved by the Wake Forest University Health Sciences Animal Care and Use Committee.

**Plasma Cholesterol Determination and Fast Protein Liquid Chromatography**

Plasma was collected by tail bleeding of mice fasted for 4 hours. Plasma from 4 mice was pooled and size fractionated by fast protein liquid chromatography with a Superose 6 TM column (Amersham Biosciences). Plasma cholesterol was determined with an enzymatic colorimetric assay kit (Roche Applied Science).

**Quantitative Polymerase Chain Reaction and Western Blot Analysis**

Total RNA was extracted by Trizol (Invitrogen) according to the manufacturer’s protocol. cDNA preparation and quantitative polymerase chain reaction were conducted as described previously. To prepare tissue lysate, ~0.1 g snap-frozen tissue was homogenized in radioimmunoprecipitation assay buffer. For AT, the lysate was incubated on ice for 10 minutes to remove the solidified fat cake on top. Immunoblotting of ABCA1 was conducted as previously described with 4% to 16% SDS-PAGE. Monoclonal antibodies directed against PPARγ and GAPDH were purchased from Sigma. Polyclonal antibodies against apoE, apoA-I (Biodesign), and anti–SR-B-I (Novartis) were also used for Western blotting.

**Adipose Cholesterol Determination**

AT cholesterol was determined by gas-liquid chromatography after extraction of total lipid from 0.1 g AT with chloroform:methanol (2:1). Data were normalized to tissue wet weight or protein, measured by the Lowry assay.

**Preparation of Ex Vivo Cultures of AT**

To establish the ex vivo cultures of AT, freshly isolated epididymal fat (0.1~0.2 g) was minced into ~2-mm pieces, moved into 10-cm cell culture dishes, and incubated overnight in 1.5 mL serum-free Dulbecco Modified Eagles Medium F/12 Ham (Santa Cruz). The next day, AT cultures were washed thoroughly with caution to remove floating cells without aspirating the minced tissue explants.

**Cholesterol Efflux From AT**

Measurement of cholesterol efflux was conducted according to standard methods, except ex vivo AT cultures were used. Briefly, ex vivo cultures of AT (pooled from n = 4 mice per genotype; n = 4 replicates) were incubated with 5 μCi of [3H]-cholesterol (complexed to bovine serum albumin) for 24 hours. AT explants were then washed carefully, and 1 mL fresh serum-free medium was added. Cholesterol efflux was initiated by adding either 20 μg protein/mL of human apoA-I (ABCA1-dependent efflux) or 50 μg protein/mL mouse HDL (ABCA1-independent efflux) as acceptors. After 4 hours of additional incubation, cholesterol efflux was quantified by measuring [3H]-cholesterol in 500 μL medium and in the cells after isopropanol extraction. Percentage efflux was calculated as follows: dpm [3H]-cholesterol in medium/dpm [3H]-cholesterol (medium + cells)×100%.

**Nascent HDL Formation**

All the procedures for nascent HDL formation were similar to our previous study. Briefly, 10 μg/mL lipid-free [125I]-apoA-I (105 cpm/μg) was incubated with AT explants for 24 hours. Conditioned medium (1 mL) was then fractionated by high-resolution fast protein liquid chromatography size–exclusion chromatography with 3 Superdex 200 HR columns in series. 125I radioactivity in each fraction was quantified with a gamma counter (Beckman). Electrophoretic mobility of adipose-derived nascent HDLs was analyzed with a Paragon lipoprotein agarose gel electrophoresis system (Beckman) according to the manufacturer’s instructions. Electrophoretic mobilities (α, β, pre-β) were determined after lipid staining of plasma from low-density lipoprotein receptor KO mice. The size distribution of nascent HDL particles in conditioned medium was also deter-
mained on a 4% to 30% nondenaturing gradient gel as described previously.20

HDL Catabolism and Tissue-Specific Uptake

$^{125}$I-radiolabeled tyramine cellubiose (TC)-HDL ($^{125}$I-TCHDL) was prepared as previously described, except HDL was isolated from WT mouse plasma.15,21 Recipient mice (n = 4 per genotype) were anesthetized with isoflurane, and $^{125}$I-TCHDL (500 000 cpm/animal) was injected via a jugular vein catheter. Blood samples (30 μL) were drawn at 5 minutes, 30 minutes, and 1, 3, 6, and 24 hours after tracer injection by retro-orbital bleeding, and 15 μL plasma was used for quantification of $^{125}$I radioactivity. Data were expressed as a percentage of the 5-minute plasma radioactivity remaining in plasma after injection of the tracer. The plasma die-away curves were generated with a biexponential curve-fitting program (GraphPad Prism 5; San Diego, CA). For the determination of tissue-specific uptake of HDL, mice were euthanized 24 hours after $^{125}$I-TCHDL injection, and liver, kidney, and epididymal fat pads were removed to quantify $^{125}$I radioactivity.

Statistics

Results are presented as mean±SEM. Data were analyzed statistically by the use of multiple tests. A general linear regression model was used to assess relationships between FC and ABCA1 within and across fat depots while controlling for intra-animal correlations. A Student t test was used to test for differences in AT ABCA1 expression and radiolabeled HDL tracer tissue uptake between WT and ABCA1$^{-/-}$ mice. For comparison of variables (ie, total plasma cholesterol, epididymal fat cholesterol content) among all 3 fat depots, with 5 distinct subfractions ranging in size from 7 to 17 nm in diameter.

Results

ABCA1 Expression Varies Among Fat Depots

AT exhibits regional heterogeneity with regard to cellular composition, response to hormonal signals, and metabolic function.22 To gain insight into the role of fat depot-specific cholesterol regulation by ABCA1, C57BL/6 mice (n = 3) were fed an HF (HF/high-cholesterol) diet (42% fat and 0.2% cholesterol) to increase fat mass. After the mice had consumed the diet for 8 weeks, we measured cholesterol content by gas-liquid chromatography and relative ABCA1 expression by Western blot in different fat depots. There was a 6-fold variation in FC content (no detectable cholesteryl ester) among fat depots (epididymal > mesenteric > pericardial > inguinal (subcutaneous) > brown fat). ABCA1 expression was equally variable among depots, with a strongly positive correlation ($r^2=0.34, P=0.01$) between fat depot FC content and ABCA1 expression, but not SR-BI or apoE expression (Figure 1A and 1B).

Next, we examined AT ABCA1 expression using epididymal fat depots from C57BL/6 mice. There are conflicting results in the literature on the extent to which ST3-L1 fibroblasts express ABCA1 before they are differentiated into adipocytes.17,19 To determine whether ABCA1 is expressed in nonadipocyte cells residing in AT, stromal vascular cells were fractionated from floating mature adipocytes with the use of previously described methods.23 ABCA1 expression was almost exclusively confined to adipocytes, with little expression in stromal vascular cells (online-only Data Supplemen). To establish a primary adipocyte culture system from mouse AT, isolated stromal vascular cells were incubated with a differentiation cocktail.23 After 10 days, $\approx$60% of cells were converted to lipid-laden adipocytes by Oil Red O staining (Figure IA in the online-only Data Supplement). ABCA1 expression levels were also increased during adipogenesis (Figure 1D). To determine the cellular localization of ABCA1, adipocyte cultures were immunostained with ABCA1 monoclonal antibody. ABCA1 staining was observed in the plasma membrane and cytosol of adipocytes, suggesting that ABCA1 may traffic between the plasma membrane and cytosol (Figure IB in the online-only Data Supplement).

AT Generates Nascent HDL Particles of Multiple Sizes

ABCA1-dependent FC efflux has been reported for ST3-L1 adipocytes19,24 and mouse embryonic fibroblast–derived adipocytes.17 However, ABCA1-dependent nascent HDL particle formation by unilocular adipocytes in AT has not been investigated. Thus, we established ex vivo cultures of mouse epididymal AT and incubated the freshly isolated AT explants with $^{125}$I–apoA-I for 24 hours, after which nascent HDL subpopulations in the conditioned medium were size fractionated by high-resolution fast protein liquid chromatography.10,11 Consistent with our previous studies using HEK293 cells stably expressing human ABCA1,10,11 AT from C57BL/6 mice generated heterogeneously sized nascent HDL particles that migrated in the pre-$\beta$ position on agarose gels (Figure 1E). More important, the size distribution of nascent HDL species from AT followed a pattern similar to that from HEK293-ABCA1–expressing cells (Figure 1F) and mouse peritoneal macrophages and McArdle 7777 hepatoma cells,11 with 5 distinct subfractions ranging in size from 7 to 17 nm in diameter.

Deletion of Adipocyte ABCA1 Decreases Plasma HDL Levels

To investigate the role of adipocyte ABCA1 in AT cholesterol mobilization and its impact on HDL plasma level in vivo, we generated adipocyte-specific ABCA1 KO mice (ABCA1$^{-/-}$). Adipocyte-specific ABCA1 inactivation was achieved by crossing C57BL/6 ABCA1$^{floxed}$ mice with ApoA1$^{-/-}$ mice with aP2 (fatty acid binding protein 4)-Cre transgenic mice backcrossed in the C57BL/6 background (Jackson Laboratories). ABCA1$^{-/-}$ mice were fertile, and pups were born at the expected mendelian frequency. Recombination of the floxed ABCA1 allele resulted in a $>90\%$ decrease in ABCA1 protein expression in ABCA1$^{-/-}$ mouse epididymal fat and 87% reduction of ABCA1 mRNA in epididymal fat (Figure 2A). Inactivation of adipose ABCA1 had little effect on ABCA1 expression in liver and intestine, which are the main organs known to be important for HDL biogenesis (Figure 2B). However, ABCA1 expression was noticeably reduced in resident peritoneal macrophages (Figure 2C), thioglycolate-elicited macrophages (Figure 2D), and, to a lesser extent, in bone marrow–derived macrophages (Figure 2C). ABCA1 expression in WT epididymal and mesenteric AT was abundant and comparable to that in liver and

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macrophages, but its expression was markedly blunted in ABCA1−/−A mouse epididymal, brown, and mesenteric fat (Figure 2D). Thus, deletion of ABCA1 expression was highly efficient in AT and somewhat variable in macrophages. However, macrophage ABCA1 expression has minimal impact on plasma HDL levels,14,25–27 so we proceeded with studies on the role of adipocyte ABCA1 expression on HDL metabolism and adipocyte cholesterol balance.

Next, we evaluated the contribution of adipose ABCA1 expression on plasma cholesterol concentrations and distribution using WT and ABCA1−/−A mice consuming chow or an HF diet. There was a stepwise reduction in total plasma cholesterol with successive deletion of ABCA1 alleles (WT, 83±3 mg/dL; heterozygotes, 70±6 mg/dL; homozygotes, 62±4 mg/dL; Figure 3A) that reached statistical significance for male WT versus ABCA1−/−A mice (P<0.0084 by ANOVA with the Tukey post hoc test). In addition, Western blot analysis of plasma suggested a reduction in apoA-I (KO/WT ratio 0.72) and apoE (KO/WT ratio 0.86) concentration but not lecithin-cholesterol acyltransferase (KO/WT ratio 1.03) (Figure 3B). The lower plasma cholesterol levels for ABCA1−/−A mice persisted throughout the 16 weeks of HF diet (2-way ANOVA with repeated mea-
sures; time effect, \( P < 0.0001 \); genotype effect, \( P < 0.001 ; n = 4 \) each), suggesting that deletion of adipose ABCA1 results in decreased plasma cholesterol pool size despite expanding AT mass (Figure 3C). Fast protein liquid chromatography fractionation of plasma revealed that the decreased plasma cholesterol in ABCA1\(^{-/-}\) mice was mainly in the HDL region of the column (Figure 3D). These data provide evidence that AT ABCA1 expression contributes a small but significant fraction of the plasma HDL pool.

To exclude the possibility that decreased plasma HDL concentration in ABCA1\(^{-/-}\) mice was due to increased hepatic uptake and degradation of HDL, we measured plasma clearance and tissue uptake of HDL apoA-I. Because HDL from WT and ABCA1\(^{-/-}\) mice was similar in chemical composition (data not shown), HDL was isolated from plasma of WT mice and radiolabeled with TC, a tissue residuallizing compound that allows quantification of tissue uptake of radiolabeled tracer during turnover studies.\(^{15,21}\) The plasma removal of \([^{125}\text{I}]\)-TC-HDL tracer in ABCA1\(^{-/-}\) mice was indistinguishable from that of WT mice over the 24-hour turnover study (Figure 4A). There also was no difference in tissue uptake of HDL tracer by liver, kidney, or AT (Figure 4B), indicating that selective inactivation of adipocyte ABCA1 did not affect HDL catabolism in vivo. These results suggested that the decreased plasma HDL levels in ABCA1\(^{-/-}\) mice were due to decreased HDL production by AT.

Deletion of Adipocyte ABCA1 Blunted ABCA1-Dependent Cholesterol Efflux and Large Nascent HDL Formation in AT

To determine whether deletion of ABCA1 in adipocytes resulted in increased FC content, similar to what we observed in macrophages from macrophage-specific ABCA1 KO mice,\(^{14}\) we measured TC and FC in epididymal and brown fat from 3 to 4 mice after 16 weeks of HF/high-cholesterol diet feeding. There was a gene dose-related increase in FC content (\( P < 0.001 \)) in epididymal fat that reached statistical significance in ABCA1\(^{-/-}\) mice (Figure 5A). Cholesteryl ester was low to undetectable in the samples (data not shown). These results suggested that FC efflux from adipose explants was severely compromised in the absence of functional ABCA1. To address this question, we incubated AT explants from chow-fed mice with \(^3\text{H-cholesterol} for 24\) hours to radiolabel the cells, washed the cells, and incubated them for 4 hours with apoA-I (20 \( \mu \text{g/mL} \)) to measure ABCA1-dependent efflux or with HDL (50 \( \mu \text{g/mL} \)) to measure ABCA1-independent efflux. ABCA1-dependent \(^3\text{H-FC efflux} to apoA-I was completely eliminated in ABCA1\(^{-/-}\) epididymal fat compared with WT. However, HDL-mediated FC efflux (ABCA1 independent) was unaffected by deletion of adipose ABCA1 expression (Figure 5B). To determine whether the ablation of ABCA1-dependent efflux to apoA-I in ABCA1\(^{-/-}\) epididymal fat was accompanied by loss of...
nascent HDL formation, we incubated $^{125}$I–apoA-I with ex vivo cultures of WT or ABCA1$^{-/-}$ epididymal fat and monitored nascent HDL formation by non-denaturing gel electrophoresis and high-resolution fast protein liquid chromatography (Figure 5C). WT epididymal fat generated multiple, heterogeneous-sized HDL subfractions that ranged in size from 7 to 12 nm in diameter, which we have designated pre-β 1 to 5 in order of increasing particle size. We have also observed similar nascent HDL particle size heterogeneity in cultures of HEK293 cells stably expressing ABCA1, mouse peritoneal macrophages, and rat McArdle 7777 hepatoma cells. In contrast, epididymal fat from ABCA1$^{-/-}$ mice generated only particles <10 nm in diameter (ie, pre-β 2).

**Discussion**

This study was designed to elucidate the role of adipocyte ABCA1 expression in nascent HDL particle formation and its contribution to the plasma HDL pool in vivo. To address this issue, we generated ABCA1$^{-/-}$ mice and studied them while they were consuming a chow or HF diet. Several novel observations were made in this study. First, AT ABCA1 expression varied over a 5-fold range among different ATs, suggesting an important role for ABCA1 expression in AT sterol homeostasis. Second, ABCA1$^{-/-}$ mice had <10% expression of ABCA1 protein in AT compared with WT mice, with a concomitant 2-fold increase in AT cholesterol content. Although FC efflux from AT explants to HDL was similar between WT and ABCA1$^{-/-}$ mice, FC efflux to apoA-I was eliminated in ABCA1$^{-/-}$ AT, suggesting an important and specific role for ABCA1 in AT FC homeostasis. Third, WT AT explants were capable of generating multiple, discrete-sized nascent HDL subfractions from apoA-I, ranging in diameter from 7 to 12 nm, similar to other cell types. However, ABCA1$^{-/-}$ AT generated nascent HDL particles <10 nm, suggesting a considerable loss of function in the ability to generate nascent HDL. The decreased ability to generate nascent HDL was reflected in vivo as a 15% reduction in plasma HDL concentration in ABCA1$^{-/-}$ mice versus WT mice that was attributed to HDL production because HDL catabolism was similar between the 2 genotypes.

**Figure 3.** Deletion of adipocyte ATP binding cassette transporter A1 (ABCA1) reduces plasma cholesterol and high-density lipoprotein (HDL) concentration. A, Total plasma cholesterol (TC) levels of chow-fed wild-type (WT; +/+), heterozygous (−/−/+), and homozygous (−/−−) adipocyte-specific ABCA1 knockout mice. B, Plasma levels of apolipoprotein (apo) A-I, apoE, and lecithin-cholesterol acyltransferase determined by Western blot analysis of 1 μL plasma from 4 chow-fed WT and ABCA1$^{-/-}$ mice. C, Total plasma cholesterol concentration of WT (+/+; ○) and ABCA1$^{-/-}$ (−/−−; ●) mice over 16 weeks of a high-fat (HF) diet (2-way ANOVA with repeated measures; time effect, P<0.0001; genotype effect, P<0.0001; n=4 of each genotype). D, Fast protein liquid chromatography (FPLC) separation of plasma lipoproteins from WT (+/+; ○) and ABCA1$^{-/-}$ (−/−−; ●) mice before (chow) and after 2 and 16 weeks of HF diet feeding. Each profile represents a separation of a plasma pool from 4 mice of each genotype. Lipoprotein elution was monitored by enzymatic assay of cholesterol in individual fractions (Fr); elution positions of very-low-density lipoprotein (VLDL), low-density lipoprotein (LDL), and HDL are indicated on the left.
Adipocytes were thought to have low ABCA1 expression, but subsequent studies using 3T3L1 adipocytes and AT demonstrated that ABCA1 is abundantly expressed. Furthermore, we observed that ABCA1 expression varies over a 5-fold range among AT depots and that this variation is highly correlated with AT FC content. These data suggest that as metabolically active visceral fat stores enlarge during TG accretion, FC also increases; this in turn upregulates expression of ABCA1 to rebalance cellular cholesterol homeostasis. ABCA1 gene transcription is highly regulated by liver X receptor expression in some tissues, but whether this pathway is responsible for variable ABCA1 protein expression among AT depot sites is unknown. Nevertheless, these data establish a critical association between AT ABCA1 expression and FC content, which may be important in the modulation of adipocyte metabolic activity.

To determine the role of AT ABCA1 expression in plasma HDL concentrations, we generated ABCA1−/− mice by crossing ABCA1+/− mice with aP2 Cre recombinase transgenic mice. ABCA1 protein expression in epididymal fat decreased in proportion to the number of inactivated ABCA1 alleles (Figure 1A). Recombination of the ABCA1 gene was efficient in all AT depots examined, with >90% elimination of ABCA1 protein expression in ABCA1−/− mice. Expression of SR-BI, another protein important in adipocyte FC efflux, was unaffected (Figure 1D). ABCA1−/− mice had lower plasma total and HDL cholesterol concentrations compared with WT mice, whether they were fed chow or an HF diet (Figure 3C and 3D). To determine whether this phenotype was due directly to depletion of AT ABCA1 or indirectly to changes in ABCA1 expression in other tissues, the specificity of the recombination was investigated. ABCA1 expression in liver and intestine, 2 tissues previously shown to be important in determining plasma HDL concentrations, was similar in WT and ABCA1−/− mice and could not account for the differences in plasma HDL concentrations. aP2 Cre recombinase has been reported to be active in macrophages, so we also examined ABCA1 expression in resident, elicited peritoneal, and bone marrow–derived macrophages. ABCA1 expression was variably reduced in macrophages from ABCA1−/− mice, confirming previous results that aP2 Cre recombinase was active in macrophages. However, evidence from our laboratory and others using transplantation of total ABCA1 KO bone marrow has demonstrated that macrophage ABCA1 expression has no significant impact on plasma HDL concentrations. Taken together, these data suggest that AT ABCA1 expression plays a significant role in determining plasma HDL concentrations.

Although our data suggested that HDL production by AT was reduced in the absence of adipocyte ABCA1 expression, deletion of adipocyte ABCA1 may have indirectly increased HDL catabolism, resulting in reduced plasma HDL levels. Previous studies in humans suggest that HDL catabolism plays an important role in determining plasma HDL concentrations. To formally examine this possibility, we performed HDL turnover studies using radiolabeled WT HDL because HDL composition was similar between WT and ABCA1−/− mice (data not shown). Plasma decay of the HDL tracer and uptake of radiolabel by the liver and kidney, the 2 most important tissues in HDL protein uptake and catabolism, were similar for both genotypes of mice (Figure 4). From these data, we conclude that decreased plasma HDL concentrations in ABCA1−/− mice were due to decreased HDL production by AT relative to WT mice and not to differences in HDL catabolism.

Although our study and others illuminate AT in HDL production, direct evidence for the assembly of nascent HDL particles by AT has not been demonstrated. In this study, we show that epididymal fat from ABCA1−/− mice had a doubling of FC content, was defective in FC efflux to apoA-I, and was defective in the production of nascent HDL particles >10 nm in diameter (Figure 5). Zhang et al have shown that both ABCA1 and SR-BI, but not ABCG1, are important for FC efflux from adipocytes, differentiated from mouse embryonic fibroblasts. Our study confirms and extends their results by showing that ABCA1 is critical for maintaining FC content and efflux to ABCA1 in primary AT explants. Furthermore, the 2-fold increase in AT FC content suggests that SR-BI cannot compensate for the loss of ABCA1 expression. This was confirmed by the similar expression of SR-BI among AT depots in WT and ABCA1−/− mice (Figure 2D). Previous studies have shown that ectopic expression of ABCA1 in cell lines is necessary and sufficient to
generate several discrete-sized nascent HDL particles. We have designated these particles pre-HDL1 to 5 on the basis of increasing particle size on nondenaturing gradient gels and fast protein liquid chromatography columns and mobility on agarose or 2-dimensional gels. These nascent pre-HDL particles appear to be formed simultaneously in cell culture systems, with no evidence for a precursor-product dependency (ie, small particles → larger particles). Similar discrete, nascent HDL particle sizes have previously been observed for McArdle 7777 hepatoma cells, mouse peritoneal macrophages, and primary hepatocytes (unpublished data), in addition to transfected cell lines, suggesting that ABCA1 expression is the necessary and sufficient condition for nascent HDL size heterogeneity among these varied cell types. Epididymal fat explants from WT mice also generated nascent HDL particles separated by nondenaturing gradient gel electrophoresis (4% to 30%). Migration positions of the radiolabeled nascent HDL particles were visualized with a Phosphorimager.

Figure 5. Adipocyte ATP binding cassette transporter A1 (ABCA1) generates heterogeneous-sized nascent high-density lipoprotein (HDL) particles. A, Free cholesterol accumulation in epididymal adipose tissue (AT) with progressive inactivation of adipocyte ABCA1 alleles. Wild-type (WT; +/+), heterozygote (−/−), and homozygote (−/−) of adipocyte-specific ABCA1 knockout mice. B, Cholesterol efflux to apolipoprotein (apo) A-I (ABCA1 dependent) and HDL (ABCA1 independent). Ex vivo cultures of epididymal fat from +/+ and −/− mice were radiolabeled with [3H]-cholesterol for 24 hours, washed, and then incubated with no addition (no Trt), apoA-I (hA-I; 20 μg/mL), or mouse HDL (mHDL; 50 μg/mL) for 4 hours to measure [3H]-FC efflux into the medium. For A and B, values are mean ± SEM; n = 4. *P < 0.05, **P < 0.01, ***P < 0.001; not significant (ns) at P > 0.05. C, Nascent HDL formation in the presence of [125I]-apoA-I in ex vivo cultures of epididymal AT from +/+ and −/− mice. After a 24-hour incubation, conditioned medium was fractionated by high-resolution fast protein liquid chromatography, and radiolabeled apoA-I in each fraction was quantified with a gamma counter. Elution regions for different-sized nascent pre-β HDL particles are denoted by the vertical dashed lines. Inset shows the size of AT–derived nascent HDLs separated by nondenaturing gradient gel electrophoresis (4% to 30%). Migration positions of the radiolabeled nascent HDL particles were visualized with a Phosphorimager.
adipocyte plasma membrane, resulting in increased ABCA1-mediated HDL particle formation and increased plasma HDL concentrations. Thus, adipocyte ABCA1 may be a critical negative regulator of obesity and metabolic syndrome, preventing adipocyte dysfunction and abnormal plasma membrane FC accumulation by facilitating FC efflux and nascent HDL formation.

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Disclosures

None.

References

CLINICAL PERSPECTIVE

Adipose tissue is a major pool of whole-body free cholesterol (FC) and abundantly expresses ATP binding cassette transporter A1 (ABCA1), a membrane protein necessary for high-density lipoprotein (HDL) particle formation. Although adipocytes abundantly express ABCA1, their contribution to HDL production in vivo is unknown. Using adipocyte-specific ABCA1 knockout mice, we demonstrate for the first time that adipocytes make nascent, discrete-sized HDL particles that represent a significant (~15%) source of plasma HDL. Furthermore, deletion of ABCA1 in adipocytes results in a 2-fold increase in FC content, suggesting that other FC export mechanisms cannot compensate for loss of ABCA1. Adipose tissue ABCA1 expression varies over a 5-fold range among different adipose depots in wild-type mice and is significantly correlated with adipose tissue FC content, suggesting an important role for ABCA1 in adipose tissue FC homeostasis. Because adipocyte FC content increases with obesity and adipocyte hypertrophy, we speculate that ABCA1 is upregulated in adipocytes to maintain optimal FC balance between the lipid droplet and plasma membrane and adipocyte function. Accretion of FC during adipocyte expansion may sequester FC on lipid droplets, shunting it away from ABCA1-mediated HDL particle formation, resulting in decreased plasma HDL concentrations with development of obesity. In contrast, rapid weight loss may result in FC mobilization to the adipocyte plasma membrane, resulting in increased ABCA1-mediated HDL particle formation and increased plasma HDL concentrations. Thus, adipocyte ABCA1 quantitatively contributes to plasma HDL levels and may be critical as a negative regulator of obesity and metabolic syndrome by preventing adipocytes from abnormal FC accumulation and dysfunction.
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Supplement Figure 1. ABCA1 is exclusively expressed in adipocytes in primary cultures of adipocytes. A. Oil red O staining of primary stromal vascular cells and differentiated adipocytes (at d10). B. Localization of adipocyte ABCA1, immunostained with monoclonal ABCA1 antibody (red) in differentiated cultures of adipocytes. Nuclei were counter-stained with DAPI (blue).
Supplement Figure 1

A

Stromal Vascular  Differentiated Adipocytes

B