Increased Endoplasmic Reticulum Stress in Atherosclerotic Plaques Associated With Acute Coronary Syndrome

Masafumi Myoishi, MD; Hiroyuki Hao, MD, PhD; Tetsuo Minamino, MD, PhD; Kouki Watanabe, MD, PhD; Kensaku Nishihira, MD, PhD; Kinta Hatakeyama, MD, PhD; Yuijro Asada, MD, PhD; Ken-ichiro Okada, MD, PhD; Hatsue Ishibashi-Ueda, MD, PhD; Giulio Gabbiani, MD, PhD; Marie-Luce Bochaton-Piallat, PhD; Naoki Mochizuki, MD, PhD; Masafumi Kitakaze, MD, PhD

Background—The endoplasmic reticulum (ER) responds to various stresses by upregulation of ER chaperones, but prolonged ER stress eventually causes apoptosis. Although apoptosis is considered to be essential for the progression and rupture of atherosclerotic plaques, the influence of ER stress and apoptosis on rupture of unstable coronary plaques remains unclear.

Methods and Results—Coronary artery segments were obtained at autopsy from 71 patients, and atherectomy specimens were obtained from 40 patients. Smooth muscle cells and macrophages in the fibrous caps of thin-cap atheroma and ruptured plaques, but not in the fibrous caps of thick-cap atheroma and fibrous plaques, showed a marked increase of ER chaperone expression and apoptotic cells. ER chaperones also showed higher expression in atherectomy specimens from patients with unstable angina pectoris than in specimens from those with stable angina. Expression of 7-ketocholesterol was increased in the fibrous caps of thin-cap atheroma compared with thick-cap atheroma. Treatment of cultured coronary artery smooth muscle cells or THP-1 cells with 7-ketocholesterol induced upregulation of ER chaperones and apoptosis, whereas these changes were prevented by antioxidants. We also investigated possible signaling pathways for ER-initiated apoptosis and found that the CHOP (a transcription factor induced by ER stress)-dependent pathway was activated in unstable plaques. In addition, knockdown of CHOP expression by small interfering RNA decreased ER stress-dependent death of cultured coronary artery smooth muscle cells and THP-1 cells.

Conclusions—Increased ER stress occurs in unstable plaques. Our findings suggest that ER stress-induced apoptosis of smooth muscle cells and macrophages may contribute to plaque vulnerability.

Key Words: apoptosis ■ plaque ■ myocardial infarction ■ endoplasmic reticulum

Editorial perspective

Most of the acute clinical manifestations of coronary atherosclerosis result from plaque rupture that triggers thrombosis and vessel occlusion, producing the acute coronary syndrome (ACS).1-3 Previous reports have shown that apoptosis affects all of the types of cells residing within atherosclerotic plaques, including smooth muscle cells (SMCs) and macrophages,4,5 with oxidized low-density lipoprotein and several inflammatory factors being known to induce apoptosis.6,7 The number of apoptotic cells depends on the plaque stage and is generally higher in more advanced plaques.6,8 SMCs synthesize most of the interstitial collagen that stabilizes the fibrous cap of a plaque.4,7 Therefore, excessive apoptosis of SMCs in the fibrous cap may compromise plaque integrity and render it vulnerable to proteolytic attack by inflammatory cells, leading to plaque rupture.4,7 Apoptotic macrophages are more frequent at sites of plaque rupture than in areas where the fibrous cap remains intact.9 A decrease in macrophages would reduce the scavenging of apoptotic SMCs and macrophages, allowing the cells to undergo secondary necrosis, thereby increasing thrombogenicity of the plaque.10
including ischemia, hypoxia, heat shock, mutation, increased protein synthesis, and reactive oxygen species, all of which can potentially lead to ER dysfunction. In response to ER stress, there is marked upregulation of various ER chaperones, such as the 94-kDa glucose-regulated protein (GRP94) or GRP78 that stabilizes protein folding. When the ER becomes overloaded with misfolded proteins, the unfolded protein response (UPR) occurs to enhance cell survival. However, prolonged ER stress can trigger apoptotic cell death, which is promoted by transcriptional induction of C/EBP homologous protein (CHOP) and/or by the activation of c-JUN NH2-terminal kinase (JNK) and/or caspase-12–dependent pathways. In support of this concept, our investigation of the effects of prolonged ER stress on hypertrophic and failing hearts revealed that apoptosis of cardiac myocytes was induced via activation of CHOP, an ER-specific proapoptotic factor. An important role of ER-initiated cell death pathways has also been demonstrated in several diseases, including diabetes mellitus, neurodegenerative conditions, and ischemia.

Oxidation of low-density lipoprotein plays a significant pathogenetic role in atherosclerosis. In cultured peritoneal macrophages, excessive accumulation of free cholesterol (induced by acetyl low-density lipoprotein with an acyl-CoA:cholesterol acyltransferase [ACAT] inhibitor) initiates ER stress, increases CHOP expression, and leads to apoptosis. Studies of apoE–/– mice also support the relevance of ER stress to macrophage apoptosis and to enlargement of the necrotic core in advanced atherosclerotic plaques. However, it is still unclear whether ER stress and UPR activation have a role in plaque rupture. Unfortunately, the absence of a suitable animal model has greatly hindered investigation of the molecular mechanisms of plaque rupture and evaluation of the effects of ER stress in vivo.

In the present study, we examined histological sections from atherosclerotic coronary artery lesions obtained at autopsy or after directional coronary atherectomy (DCA) to investigate markers of ER stress/UPR activation and apoptotic cell death. Oxytetrosols such as 7-ketocholesterol (7-KC) have been reported to be partially responsible for the cytotoxicity of oxidized low-density lipoprotein. Exposure of cultured human SMCs to 7-KC induces the UPR and promotes apoptotic cell death, so we investigated 7-KC expression in plaque specimens by immunohistochemistry. We also examined whether 7-KC could activate ER stress using cultured human coronary artery SMCs (CASMCs) and a monocyte cell line (THP-1). Furthermore, we investigated the possible signaling pathways for ER-initiated apoptosis, and we found that the CHOP (a transcription factor induced by ER stress)-dependent pathway was activated in unstable plaques, whereas knockdown of CHOP expression by small interfering RNA (siRNA) decreased ER stress-dependent death of cultured CASMCs and THP-1 cells.

Methods

Coronary Artery Specimens

Two different sets of specimens were obtained under a protocol approved by the Institutional Review Board of the National Cardiovascular Center and Miyazaki University. The first set of specimens was obtained at autopsy, and the second set was obtained by DCA. Classification of the histology of the lesions in autopsy specimens was done morphologically, as described previously (Table 1). Demographic data for the study population are presented in Table I of the Data Supplement.

In brief, 152 coronary artery segments were obtained at autopsy from 71 patients, including 17 consecutive patients who experienced fatal ACS without percutaneous coronary intervention and 54 consecutive patients with noncardiac death. The major coronary arteries and their branches were cut transversely at 5-mm intervals, and 17 ruptured plaques were detected in the 17 ACS patients (ruptured: AHA type VI, n = 17). The remaining 33 patients with noncardiac death and the 17 ACS patients also had advanced unruptured plaques (>75% cross-sectional luminal narrowing), and we assessed each segment at the narrowest point (n = 114). The advanced atherosclerotic unruptured plaques were additionally divided into fibrous plaques (fibrous: fibrocellular tissue was the predominant component, and the lipid core was inconspicuous or absent; AHA type Vc, n = 48), thick-cap atheroma (thick: a lipid core covered by a fibrous cap >65-μm thick; AHA type Va, n = 51), and thin-cap atheroma (thin: a lipid-rich core covered by a fibrous cap <65-μm thick; AHA type Ta, n = 15). Another 21 patients with noncardiac death who had no advanced unruptured plaques and normal coronary arteries that only showed diffuse intimal thickening (normal: AHA type I, n = 21) were used as a control group.

We performed a morphological analysis of multiple lesions (n = 152) obtained at autopsy from 71 patients (Figure 1). The supplementary analyses included pairwise comparison of unruptured and ruptured plaques from each heart of each patient with ACS (Data Supplement Figure I) and investigation of the correlation between traditional cardiovascular risk factors and ER stress (Data Supplement Table II and Figure III). These supplementary analyses were based on representative data from each patient. (Details of the methods used to perform the supplementary analyses are included in the expanded Methods section in the Data Supplement.)

Forty DCA specimens were obtained from 40 patients who were treated for stable angina pectoris (SAP; n = 20) or unstable angina pectoris (UAP; n = 20). One DCA specimen was obtained per patient, and these specimens were classified on the basis of the clinical situation at the time of DCA (Table 2). These specimens were fixed in 4% paraformaldehyde for 6 hours at 4°C and then embedded in paraffin.

Immunohistochemistry

Serial sections were examined by immunohistochemistry, as described previously. In brief, sections were deparaffinized, and endogenous peroxidase activity was blocked by incubation with 0.3% H2O2 in methanol for 30 minutes. For some antibodies, antigen retrieval was performed as specified below. After blocking with 3% normal bovine serum albumin, sections were incubated with the primary antibody overnight at 4°C. KDEL (Lys-Asp-Glu-Leu) antibody, which recognizes both GRP78 and GRP94, was purchased from Stressgen (San Diego, Calif) and was used at a dilution of 1:2000. Anti-CHOP antibody was obtained from Santa Cruz Biotechnology (Santa Cruz, Calif) and was applied at a dilution of 1:600

<table>
<thead>
<tr>
<th>Table 1. Human Coronary Specimens (Autopsy; n=71)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Histological Classification of Lesions</td>
</tr>
<tr>
<td>---------------------------------</td>
</tr>
<tr>
<td>Diffuse intimal thickening (normal)</td>
</tr>
<tr>
<td>Fibrous plaques (fibrous)</td>
</tr>
<tr>
<td>Thick-cap atheroma (thick)</td>
</tr>
<tr>
<td>Thin-cap atheroma (thin)</td>
</tr>
<tr>
<td>Ruptured plaques (ruptured)</td>
</tr>
</tbody>
</table>

AHA Classification indicates American Heart Association histological criteria.
after antigen retrieval by incubation for 10 minutes at room temperature in 5 μg/mL proteinase K. Anti-phospho-c-JUN NH₂-terminal kinase antibody was used to detect c-JUN kinase (JNK), which is involved in the UPR.16 It was obtained from Cell Signaling (Davers, Mass) and was applied at a dilution of 1:100 after heat retrieval for 15 minutes at a sub-boiling temperature in 1 mmol/L EDTA (pH 8.0). Colon carcinoma sections were stained with anti-phospho-JNK antibody as a positive control. Anti-human CD68 antibody (DAKO, Glostrup, Denmark) were used to identify SMCs and macrophages, respectively, and were used

Table 2. Human Coronary Specimens (Atherectomy; n = 40)

<table>
<thead>
<tr>
<th>Origin and Classification of Plaques</th>
<th>No. of Specimens</th>
</tr>
</thead>
<tbody>
<tr>
<td>SAP</td>
<td>20</td>
</tr>
<tr>
<td>UAP</td>
<td>20</td>
</tr>
</tbody>
</table>

**Figure 1.** Induction of ER chaperones and death signals in coronary artery plaques obtained at autopsy. A, Comparison of hematoxylin-eosin (HE) staining, KDEL immunostaining, and CHOP immunostaining of normal arteries (n = 14), fibrous plaques (n = 48), thick-cap atheroma (n = 51), thin-cap atheroma (n = 15), and ruptured plaques (n = 17) obtained at autopsy from 71 patients. Representative HE-stained low-power micrographs from each group (a through e). L indicates the lumen, and the arrow shows the site of plaque rupture. The parts of the intima (a) and fibrous cap (b through e) indicated by boxes are shown at a higher magnification in panels f through j. Panels k through o show KDEL immunostaining. Panels p through t show CHOP immunostaining. Panels u through y show α-smooth muscle actin (α-SMA) immunostaining. Panels z through δ show CD68 immunostaining. B and C, The number of KDEL-positive cells (B) and CHOP-positive (C) cells. The absolute number per square millimeter is shown for the media of a normal artery and for the fibrous caps of fibrous plaques, thick-cap atheroma, thin-cap atheroma, and ruptured plaques (B, C). D, ISH analysis of GRP78 (a, d), CHOP (b, e), and negative control (c, f) mRNA expression in thick- and thin-cap atheromas. E, Comparison of HE staining (a) with double immunostaining (b). Colocalization of CHOP (red) with TUNEL-positive cells (brown) in the cap of a ruptured plaque. The area indicated by asterisks is shown at a higher magnification in the inset. Arrows show CHOP and TUNEL double-positive cells. F and G, Percentage of TUNEL-positive cells (F) and percentage of TUNEL-positive cells among CHOP-positive cells (G) in the fibrous cap. Scale bars represent 1 mm (A, a through e), 50 μm (A, f through δ, D, and E), and 20 μm (E, in inset). *P < 0.05 vs normal plaque.

Terminal dUTP Nick End-Labeling Method and Double Immunohistochemistry

Cells undergoing apoptosis were identified by the terminal dUTP nick end-labeling (TUNEL) method with the ApopTag In Situ Apoptosis Detection Kit (Chemicon, Temecula, Calif), as described previously.9 For simultaneous identification of CHOP and TUNEL immunoreactivity, double immunostaining of specimens was performed. First, the TUNEL method was performed with an ApopTag.
kit, and then CHOP was detected with an alkaline phosphatase–
labeled secondary antibody with Neufuksin (DAKO).

In Situ Hybridization
Digoxigenin-labeled cRNA probes and the negative control
(LNE120) were purchased from Direct Communications Inc (Hiro-
saki, Japan), and the ISH was performed as described previously33 with a Microprobe

**CGAGUCGCCUCUACUUCCCUGGUCAGGCGCUCGAUUUC-
CUUGGUGCAGAUUCACCAUUCGGUCAAUCAGAGCUCGG-
CCA UGUUGAG-3 3; CHOP: 5'- AUGCUCUCCAAUGUUGACG-
CUUGGUGCAGCAGGUCAGUUCGAUGGCAACGCAGAUUCC-
CUGCUAGGGCCUCUUACGUUUCAGCGCCUCUAAUUCUCCU-3'**. In situ hybridization
(ISH) was performed as described previously33 with a Microprobe

mainly staining system (Fisher Scientific, Pittsburgh, Pa). In brief, hybridization
of the probes (1 µg/mL) was performed for 120 minutes
at 50°C, and then anti-digoxigenin-Ap (x250, Roche, Basel,
Switzerland), as the secondary antibody, and NB/BCIP stock
solution (x50, Roche) were added.

7-KC Staining
Snap-frozen samples were obtained from 12 patients, comprising 6
with thick-cap atheroma and 6 with thin-cap atheroma. Frozen
sections were fixed in 10% neutral-buffered formalin for 1 hour at
room temperature. After blocking with 3% normal bovine serum
albumin, the sections were incubated overnight at 4°C with anti-
7-KC antibody (Nikken Seil Corporation, Fukuroi, Japan) at a
dilution of 1:100, followed by incubation with an EnVision kit for 30
minutes.

Statistical Analysis
Data are expressed as mean±SEM. For the autopsy study of multiple
lesions from many patients (Figure 1), statistical analysis was
performed with the Kruskal–Wallis H test and a post hoc Mann-
Whitney U test. For the DCA specimens, statistical analysis was
performed with the Mann-Whitney U test. Experiments with cultured
cells were performed at least 3 times each. Data obtained with
cultured cells were analyzed statistically by the unpaired Student t
test and a post hoc Mann-

Immunohistochemical Detection of 7-KC in the
Fibrous Caps of Atherosclerotic Plaques
To explore the likely molecular mechanism of activation of ER stress and the mechanistic link to apoptosis, we investi-
gated plaque lipids by staining frozen coronary artery sec-
tions with anti-7-KC antibody (Figure 3), and the in vitro
studies were performed (Figure 4). Immunoreactivity for
7-KC was increased in the fibrous caps of thick-cap atheroma,
whereas no immunoreactivity was detected in the fibrous
caps of thin-cap atheroma, whereas no immunoreactivity was detected in the fibrous
caps of thin-cap atheroma, whereas no immunoreactivity was detected in the fibrous
caps of thin-cap atheroma, whereas no immunoreactivity was detected in the fibrous
caps of thin-cap atheroma (Figure 3). In the region around the lipid core, however, 7-KC immunoreactivity was visible
in both types of atheroma (Figure 3).

Upregulation of ER Chaperones, CHOP, and Apoptosis by 7-KC and Effects of CHOP
Knockdown by siRNA in CASMCs or THP-1 Cells
Exposure of cells to 7-KC increased the expression of GRP78 and

Results
Upregulation of ER Chaperones and Apoptosis in the Fibrous Caps of Thin-Cap Atheroma and
Ruptured Plaques
In the fibrous caps of thin-cap atheroma and ruptured plaques, KDEL and CHOP immunostaining showed a marked increase
compared with the level of staining in the fibrous caps of thick-cap atheroma and fibrous plaques (Figure 1A k through
t, Figure 1B, and Figure 1C). KDEL-positive cells were more numerous than CHOP-positive cells in the fibrous caps of
thin-cap atheroma and ruptured plaques. Most of the CHOP-
positive cells also expressed KDEL, as shown by the immunostaining of serial sections. In the same hearts of the ACS patients, there was a significant difference of KDEL- and CHOP-positive cells between the unruptured and ruptured plaques (Data
Supplement Figure I). We also assessed ER chaperone
(GRP78) and CHOP expression at the mRNA level by ISH
(Figure 1D). Furthermore, we confirmed that the KDEL- and
CHOP-positive cells were SMCs or macrophages by immu-
nostaining of serial sections with anti α-smooth muscle actin
and anti-CD68, respectively (Figure 1A, u through δ). In the
fibrous caps of thin-cap atheroma and ruptured plaques, colocalization of CHOP immunoreactivity with TUNEL-
positive cells was observed by double immunostaining (Fig-
ure 1E). The percentage of TUNEL-positive cells (Figure 1F)
and the percentage of TUNEL-positive cells among CHOP-
positive cells (Figure 1G) were increased compared with the
findings in other specimens. Immunostaining for phospho-
JNK, which is a proapoptotic factor involved in ER stress, revealed no immunoreactivity in the fibrous caps of thick,
thin, or ruptured plaques (data not shown). In normal coro-
nary artery specimens with diffuse intimal thickening (Figure
1A, k and p), there was no KDEL or CHOP positivity. In the
region around the necrotic core of advanced plaques, KDEL
positivity was only observed in macrophages. There was no
significant difference in the number of KDEL-positive cells
within the area surrounding the necrotic core of thick-cap atheromas (726±88/mm²), thin-cap atheromas (741±52/
mm²), and ruptured plaques (651±102/mm²).

Upregulation of ER Stress in Atherectomy
Specimens From Patients With UAP
To estimate the activation of ER stress related to the clinical
situation, we examined histological sections obtained at
DCA. Morphometric analysis demonstrated that the number of KDEL- and CHOP-positive cells was significantly higher
in patients with UAP than in patients with SAP (P<0.05; Figures
2A, 2B, and 2C). The KDEL- and CHOP-positive cells were confirmed to be SMCs and macrophages (Figure
2A). When ER chaperone (GRP78) and CHOP mRNA levels were analyzed by quantitative reverse-transcription polymer-
ase chain reaction or ISH, GRP78 expression was increased
in patients with UAP (P=0.14; Figure 2D), whereas CHOP
expression was significantly higher in UAP patients than in
SAP patients (P<0.05; Figure 2E). On the other hand, both
GRP78 and CHOP were significantly increased according to
ISH (Figure 2F), but we could not confirm a significant
increase of GRP78 by reverse-transcription polymerase chain
reaction. This may have been because the number of fresh
DCA specimens was too low.
the antioxidants N-acetylcysteine or glutathione (Figure 4A). We observed intracellular production of reactive oxygen species after exposure to 7-KC, whereas glutathione reduced reactive oxygen species production (Figure 4B). We also examined the effects of 7-KC on apoptosis of CASMCs and THP-1 cells (Figure 4C). Treatment with 7-KC increased FITC-annexin and propidium iodide staining in a dose-dependent (Figure 4C, b and c) and time-dependent (data not shown) manner. Treatment of CASMCs and THP-1 cells with 7-KC for 24 hours also induced apoptosis along with the induction of ER chaperones and CHOP at the protein level (Figure 4D). When CASMCs and THP-1 cells were simultaneously incubated with 7-KC and N-acetylcysteine or glutathione, both antioxidants reduced the induction of ER chaperones (Figure 4D). Quantitative analysis revealed that most of the CHOP-positive cells coexpressed KDEL (88.2% of CHOP-positive CASMCs and 72.7% of CHOP-positive THP-1 cells; P<0.05, Fisher’s exact test), whereas there were few KDEL-negative and CHOP-positive cells, which suggests that CHOP was involved in the mediation of ER-initiated signaling (Figure 4D, c and d). Treatment of THP-1 cells with 7-KC induced CHOP, whereas 2 different siRNAs targeting CHOP caused the knockdown of CHOP expression (Figure 4E, a). Knockdown of CHOP expression by siRNA decreased the number of TUNEL-positive THP-1 cells after exposure to 7-KC (Figure 4E, b and d). Similarly, the knockdown of CHOP expression by siRNA decreased the number of TUNEL-positive CASMCs after exposure to 7-KC (Figure 4E, c).
Figure 4. Upregulation of ER chaperones, CHOP, and apoptosis by exposure to 7-KC and effect of CHOP knockdown by siRNA in cultured CASMCs or THP-1 cells. A, Comparison of GRP78 and CHOP expression normalized for GAPDH by quantitative reverse-transcription polymerase chain reaction. CASMCs or THP-1 cells were incubated with 7-KC (80 mmol/L) in the absence or presence of N-acetylcysteine (NAC) or glutathione (GSH) for 12 hours. B, Measurement of reactive oxygen species (ROS) generation after exposure to 7-KC and propidium iodide staining for apoptosis of CASMCs and THP-1 cells incubated with 7-KC (a). Exposure to 7-KC induced apoptosis of CASMCs (b) and THP-1 cells (c) in a dose-dependent manner. D, KDEL and CHOP staining of CASMCs (a) and THP-1 cells (b) after incubation with 7-KC in the absence or presence of GSH for 24 hours; c and d, quantitative analysis of immunohistochemical staining of CASMCs (c) and THP-1 cells (d). E, Western blotting for CHOP after exposure to 7-KC with or without CHOP siRNA (a). TUNEL staining of THP-1 cells (b) and quantitative analysis of TUNEL-positive CASMCs (c) and THP-1 cells (d). CTL indicates the nonsilenced control. Scale bars represent 50 μm (B, C, and E) and 20 μm (D). *P<0.05 vs control (A, C, and E). #P<0.05 vs treatment with 7-KC (E). Experiments were performed at least 3 times. The data are expressed as mean±SEM. The immunofluorescent staining and Western blotting data are representative of at least 3 independent experiments.

Discussion

The present study revealed a marked increase of ER chaperone expression, CHOP expression, and apoptosis in the fibrous caps of thin-cap atheroma or ruptured plaques, as well as in atherectomy specimens from UAP patients, which suggests that ER stress may play a role in the progression of plaque vulnerability and the occurrence of acute complications of coronary atherosclerosis in humans. Because of the inherent limitations of an autopsy study, we could not exclude the possibility that UPR activation occurred after plaque rupture. Previous reports have shown that ER chaperones, such as GRP78 or GRP94, may have a protective effect against ischemia/reperfusion injury.34 However, the presence of apoptotic changes in the thin-cap atheroma rather than being secondary to plaque rupture or ischemia/reperfusion injury. Only specimens from patients without percutaneous coronary intervention were studied, to exclude the influence of this intervention. We also observed an increase of ER stress-related changes in freshly fixed atherectomy specimens obtained from UAP patients compared with those from SAP patients. This suggests that ER stress activation was related to the clinical situation, and the autopsy specimens were only slightly affected by postmortem protein degradation.

Among the oxyesters, 7-KC is most frequently detected at high levels in atherosclerotic plaques and in the plasma of patients with a high cardiovascular risk.26,35 To the best of our knowledge, however, 7-KC has not previously been detected in human atherosclerotic coronary artery sections by immunohistochemistry. It has been reported that 7-KC induces the...
production of reactive oxygen species, activation of the UPR, and induction of apoptotic death in cultured human SMCs. We demonstrated that the fibrous caps of thin-cap atheroma were immunohistochemically positive for 7-KC, a finding consistent with the increase of ER stress/UPR markers.

Treatment of CASMCs with 7-KC induced ER stress and activation of the UPR, findings that were consistent with the results of a previous study on aortic SMCs, and these changes also occurred in THP-1 cells. This 7-KC–induced cellular damage was prevented by antioxidants (N-acetylcysteine and glutathione), which was also consistent with a previous report. Accordingly, the present findings suggest that an increase of ER stress due to 7-KC induces apoptosis of SMCs and macrophages through the production of reactive oxygen species.

ER stress induces apoptosis via the CHOP-, JNK-, and caspase-12–dependent signaling pathways. CHOP is mainly induced at the translational level by ER stress, after which its overexpression leads to apoptosis. CHOP knockout mice show normal development and normal fertility but exhibit less apoptosis in response to ER stress. Thus, detection of the induction of CHOP indicates an increase of ER-initiated apoptosis. Although the direct transcriptional target of CHOP has not been found, the Bcl-2 pathway may be involved in the downstream connection between CHOP and apoptosis. Caspase-12 is only activated by ER stress. Although caspase-12 has been cloned in mice and rats, it is not yet possible to explore the role of this caspase in humans. JNK is 1 of the stress-activated protein kinases that has been shown to induce apoptosis in response to ER stress. We demonstrated that TUNEL–positive SMCs and macrophages were significantly increased in the fibrous cap, with CHOP (but not JNK) being induced simultaneously. Treatment of CASMCs or THP-1 cells with 7-KC induced CHOP, whereas knockdown of CHOP expression by siRNA led to a decrease of TUNEL–positive cells after exposure to 7-KC. Because CHOP is a transcription factor that specifically mediates ER-initiated apoptosis, the induction of CHOP in ruptured and unstable plaques supports the activation of ER-initiated apoptosis. However, our autopsy study could not exclude the possibility that the cells underwent apoptosis independently of CHOP, whereas the TUNEL assay gave false-positive results in the clinical specimens.

Unfortunately, we could not confirm whether or not the relationship between thinning of the fibrous cap and ER stress was causative because of the lack of a suitable animal model of plaque rupture. On the other hand, together with the present finding that 7-KC induced ER stress, the possibility that ER stress causes plaque vulnerability is also supported by the following reports. In cultured peritoneal macrophages, excessive accumulation of free cholesterol has been found to initiate ER stress, increase CHOP expression, and increase apoptosis. Moreover, in vivo studies with apoE–/– mice have shown that lesional necrosis can be diminished by a decrease in the cholesterol level. In addition, the present study demonstrated that expression of ER chaperones was upregulated to a similar extent in macrophages surrounding the necrotic cores of thick-cap atheroma, thin-cap atheroma, and ruptured plaques, which suggests that ER stress may contribute to the progression of plaque vulnerability by inducing macrophage apoptosis.

In conclusion, the present findings support the possibility that ER stress and/or the UPR induces apoptosis of SMCs and macrophages, thus increasing the vulnerability of coronary artery plaques, which may lead to ACS and a fatal outcome in patients with coronary artery disease.

Acknowledgments

We thank Akiko Ogai, Tomoko Morita, and Kazuyoshi Masuda for their technical assistance and Akiko Kada and Nobuo Shirahashi for their statistical advice.

Sources of Funding

The present study was supported by a grant from the Japan Cardiovascular Research Foundation and a research grant for cardiovascular disease (14C-4) from the Japanese Ministry of Health, Labor and Welfare.

Disclosures

None.
reticulum stress in hypertrophic and failing heart after aortic constriction. 
18. Nakagawa T, Zhu H, Morishima N, Li E, Xu J, Yankner BA, Yuan J. 
Caspase-12 mediates endoplasmic-reticulum-specific apoptosis and cyto-
19. Little E, Tocco G, Baudry M, Lee AS, Schreiber SS. Induction of 
glucose-regulated protein (glucose-regulated protein 78/BIP and glucose-
regulated protein 94) and heat shock protein 70 transcripts in the 
immature rat brain following status epilepticus. Neuroscience. 1996;75: 
340:115–126.
21. Feng B, Yao PM, Li Y, Devlin CM, Zhang D, Harding HP, Sweeney M, 
Kuriakose G, Tabas I. Oxysterols: friends, foes, or just fellow pas-
23. Zhou J, Lhoták S, Hilditch BA, Austin RC. Activation of unfolded protein 
response occurs at all stages of atherosclerotic lesion development in 
Eckardstein A, Exley A, Goddard M, Hofker M, Hurt-Camejo E, Kanters 
E, Kovács P, Lorkowski S, McPheat W, Pentikainen M, Rauterberg J, 
Ritchie A, Staelens B, Weitkamp B, de Winther M; for the MAFAPS 
Consortium. Rupture of the atherosclerotic plaque: does a good animal 
25. Luigens E, Suylen R, Faber BC, Gijbels MJ, Eurlings PM, Bijnen AP, 
Cleutjens KB, Heeneman S, Daemen MJ. Atherosclerotic plaque rupture: 
local or systemic process? Arterioscler Thromb Vasc Biol. 2003;23: 
2123–2130.
28. Pedruzzi E, Guichard C, Ollivier V, Driss F, Fay M, Prunet C, Marie JC, 
Pouzet C, Samadi M, Elbim C, O’Dowd Y, Bens M, Vandewalle A, 
Gougerot-Pocidalo MA, Lizard G, Ogier-Denis E. NAD(P)H oxidase 
Nox-4 mediates 7-ketocholesterol-induced endoplasmic reticulum stress 
2004;24:10703–10717.
29. Naruko T, Ueda M, Haze K, van der Wal AC, van der Loos CM, Itoh A, 
Komatsu R, Ikura Y, Ogami M, Shimagawa Y, Ebara S, Yoshimura M, 
Takeuchi K, Yoshikawa J, Becker AE. Neutrophil infiltration of culprit 
MA, Glombotski CC. Endoplasmic reticulum stress gene induction and 
protection from ischemia/reperfusion injury in the hearts of transgenic 
mice with a tamofoxifen-regulated form of p53. Circ Res. 2006;98: 
1186–1193.
Harding HP, Sweeney M, Bonart A, Sussman MA, Glombotski CC. 
1186–1193.
32. Kolodgie FD, Burke AP, Farb A, Gold HK, Yuan J, Narula J, Finn AV, 
Virmani R. The thin-cap fibroatheroma: a type of vulnerable plaque: the 
major precursor lesion to acute coronary syndromes. Curr Opin Cardiol. 
The relative mRNA expression levels of matrix metalloproteinase to 
E-cadherin in prostate biopsy specimens distinguishes organ-confined 
from advanced prostate cancer at radical prostatectomy. Clin Cancer Res. 
MA, Glombotski CC. Endoplasmic reticulum stress gene induction and 
protection from ischemia/reperfusion injury in the hearts of transgenic 
mice with a tamofoxifen-regulated form of p53. Circ Res. 2006;98: 
1186–1193.
35. Zhou Q, Wasowicz E, Handler B, Fleischer L, Kummerow FA. An excess 
concentration of oxysterols in the plasma is cytotoxic to cultured endo-
G, Lemaire S, Solary E, Gambert G. Glutathione is implied in the control 
of 7-ketocholesterol-induced apoptosis, which is associated with radical 
37. Wang XZ, Lawson B, Brewer DW, Zueger J, Iwata A, Li L, 
Boorstein R, Kreibich G. Hendershot LM, Ron D. Signals from the 
stressed endoplasmic reticulum induce C/EBP-homologous protein ( 
38. Barone MV, Curotta A, Tabae A, Phillipson L, Ron D. CHOP 
(GADD153) and its oncogenic variant, TLS/CHOP, have opposing effects 
Virmani R. The thin-cap fibroatheroma: a type of vulnerable plaque: the 
major precursor lesion to acute coronary syndromes. Curr Opin Cardiol. 
40. Fischer H, Koening U, Eckhart L, Tschachler E, Human caspase 12 has 
aquired deleterious mutations. Biochem Biophys Res Commun. 2002; 
293:722–727.

CLINICAL PERSPECTIVE

Most of the acute clinical manifestations of coronary atherosclerosis result from plaque rupture that produces the acute coronary syndrome, and apoptosis is considered to be essential for plaque rupture. The endoplasmic reticulum (ER) is 1 of the largest cellular organelles and has multiple functions, such as regulating the folding of proteins. Various stimuli cause ER stress, including ischemia, heat shock, mutation, increased protein synthesis, and reactive oxygen species, all of which can potentially lead to ER dysfunction. The ER responds to stresses by upregulation of ER chaperones, but prolonged ER stress eventually causes apoptosis. However, the influence of ER stress and apoptosis on rupture of unstable coronary plaques remains unclear. We examined histological sections from coronary artery segments obtained at autopsy from 71 patients and atherectomy specimens obtained from 40 patients. Smooth muscle cells and macrophages in the fibrous caps of thin-cap atheroma and ruptured plaques showed a marked increase of ER chaperone expression and apoptotic cells. ER chaperones also showed higher expression in atherectomy specimens from patients with unstable angina pectoris than from those with stable angina. We also investigated possible signaling pathways for ER-initiated apoptosis and found that the C/EBP homologous protein (a transcription factor induced by ER stress)–dependent pathway was activated in unstable plaques. In addition, knockdown of C/EBP homologous protein expression by small interfering RNA decreased ER stress-dependent death of cultured coronary artery smooth muscle cells and THP-1 cells. Increased ER stress occurs in unstable plaques. Our findings suggest that ER-stress-induced apoptosis may contribute to plaque vulnerability.
Increased Endoplasmic Reticulum Stress in Atherosclerotic Plaques Associated With Acute Coronary Syndrome
Masafumi Myoishi, Hiroyuki Hao, Tetsuo Minamino, Kouki Watanabe, Kensaku Nishihira, Kinta Hatakeyama, Yujiro Asada, Ken-ichiro Okada, Hatsue Ishibashi-Ueda, Giulio Gabbiani, Marie-Luce Bochaton-Piallat, Naoki Mochizuki and Masafumi Kitakaze

Circulation. published online August 20, 2007;
Circulation is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2007 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circ.ahajournals.org/content/early/2007/08/20/CIRCULATIONAHA.106.682054.citation

Data Supplement (unedited) at:
http://circ.ahajournals.org/content/suppl/2007/08/17/CIRCULATIONAHA.106.682054.DC1

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Circulation can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Circulation is online at:
http://circ.ahajournals.org//subscriptions/