Noninvasive Measurements of Infarct Size After Thrombolysis With a Necrosis-Avid MRI Contrast Agent

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Background—Gadophrin-2 is a new MRI contrast agent with high affinity for necrotic myocardium. The aim of the study was to evaluate whether noninvasive measurements of infarct size after thrombolysis are possible with gadophrin-2–enhanced MRI.

Methods and Results—Coronary artery thrombosis was induced in 3 groups of dogs by the copper-coil technique. Thrombolytic therapy together with aspirin and heparin was initiated after 90 minutes of occlusion. One day (group A), 2 days (group B), or 6 days (group C) after infarction, gadophrin-2 was injected intravenously (50 μmol · kg⁻¹). In vivo T1-weighted segmented turbo-FLASH, in vivo T2-weighted segmented half-Fourier turbo spin echo (HASTE), and T1- and T2-weighted spin-echo MRI of the excised heart were performed 24 hours after gadophrin-2 injection. Regions of strong enhancement were observed on T1-weighted images. Planimetry of short-axis MR images and of corresponding triphenyltetrazolium chloride (TTC)-stained left ventricular (LV) slices showed a close correlation between the enhanced areas and TTC-negative areas for both in vivo (r²=0.98, P<0.0001; mean difference, 0.9±2.0% [SD] of the LV volume [LVV]) and postmortem (r²=0.99, P<0.0001; mean difference, 0.9±1.4% of LVV) measurements. T2-weighted images overestimated the infarct size by 8.1±5.4% of LVV. The mean infarct size was 10.8±11.6% of LVV (group A), 22.4±11.7% (group B), and 5.1±9.3% (group C).

Conclusions—In this animal model, in vivo gadophrin-2–enhanced MRI could precisely determine infarct size after thrombolytic therapy. This technique may be very useful for the noninvasive evaluation of infarct size after reperfusion for AMI. (Circulation. 1999;99:690-696.)

Key Words: magnetic resonance imaging • myocardial infarction • thrombolysis • diagnosis

Noninvasive measurements of infarct size provide valuable information in clinical practice. Several methods have been used: ECG scores, cardiac enzyme measurements, and echocardiographic and scintigraphic imaging techniques. Although good correlations with the histological infarct size were reported, these methods have several limitations, including inability to detect small infarcts, inability to distinguish between infarcted and ischemic myocardium, and relatively poor spatial resolution.

Cardiac MRI emerged as an alternative method for the evaluation of myocardial infarction (MI). Although T2-weighted (T2w) spin-echo sequences are able to detect the presence of necrosis, the areas of increased signal intensity (SI) are probably more related to the presence of myocardial edema, systematically overestimating infarct size. Therefore, contrast enhancement is needed for optimal contrast between viable and necrotic tissue. Gd-DTPA has been used for infarct-size quantification, but the enhancement with this contrast agent is nonspecific, and therefore, infarct-size measurements may be less accurate.

Gadophrin-2 (previously referred as bis-gadolinium-mesoporphyrin) belongs to a type of MRI contrast media originally developed as “tumor-seeking” agents. It was recently shown that gadophrin-2 has affinity only for necrotic tissues. The usefulness of gadophrin-2–enhanced MRI for the identification and quantification of acute MI was recently demonstrated. The aim of the present study was to evaluate whether noninvasive measurements of infarct size after thrombolysis are possible with gadophrin-2–enhanced MRI.

Methods

Experimental Model

Dogs weighing 18 to 30 kg were sedated with xylazine (Rompun, Bayer AG), anesthetized with sodium pentobarbital (Nembutal,
Sanoﬁ), intubated, and artiﬁcially ventilated. Occlusive thrombosis of the left anterior descending coronary artery (LAD) was induced with a copper-coil model. After 90 minutes of occlusion, thrombolysis was initiated with alteplase (0.1 mg/kg bolus and 0.01 mg/kg/h infusion for 30 minutes; Actilyse, Boehringer Ingelheim GmbH, aspirin (5 mg/kg bolus; Aspecig, Synthelabo), and heparin (100 IU/kg bolus and 25 IU/kg/h infusion for 2 hours; Heparine Rorer, Rhône-Poulenc Rorer). LAD ﬂow was monitored with an electromagnetic ﬂowmeter throughout the occlusion and treatment periods. After 2 hours of heparin infusion, the copper coil was withdrawn to prevent reocclusion; the LAD patency was rechecked by angiography 1 hour after withdrawal.

One day (group A), 2 days (group B), or 6 days (group C) after thrombolysis, gadolinium-2 was given intravenously (50 µmol/kg). Twenty-four hours after injection, the dogs were reanesthetized, intubated, and ventilated, and in vivo MRI was performed. After the dogs were killed, the heart was excised and embedded in agar, and postmortem MRI was performed. The agar block was then cut perpendicular to the long axis into 6-mm-thick slices starting from the apex (9 to 11 slices per left ventricle [LV]). The slices were immersed in a buffered triphenyltetrazolium chloride (TTC) solution; on TTC staining, normal myocardium appears brick red and infarcted myocardium pale yellow. With this technique, we were able to obtain TTC-stained LV slices corresponding to every third postmortem MR image and to the in vivo MRI imaging planes (Figure 1). Calibrated photographs of the TTC-stained slices were digitized and stored on photo-CD. All experiments conformed to the “Position of American Heart Association on Research Animal Use” and were conducted with the approval of the Ethics Committee of the University of Leuven.

MRI Study

All measurements were performed with a 1.5-T whole-body MRI system (Siemens, Vision) with a gradient slew rate of 25 mT/m in 300 ms. In vivo MRI was performed with a head coil. We used a triggered segmented turbo-FLASH sequence for T1-weighted (T1w) imaging. The scheme started with an inversion recovery pulse, followed by a train of 33 rapid gradient echoes. The gradient echo acquisitions were characterized by TR/TE/FL of 7.5 ms/3.4 ms/25°; TI 450 to 550 ms. A field of view (FOV) of 240×320 mm was obtained with a 165×256 matrix; the bandwidth was 195 Hz/pixel, and the slice thickness was 6 mm. Images were acquired slice-per-slice in successive breath-hold periods of 5 to 10 heartbeats. This scheme showed tissues with short T1 with hyperintense signal. The SI in the cavity was similar to that of normal myocardium.

From preliminary acquisitions, we observed that a T2w segmented half-Fourier turbo spin echo (HASTE) sequence with “black-blood” pulse provides images with a clearly delineated cavity (see Appendix). With this technique, 2 complementary echo trains of 44 echoes with an acquisition window of 189 ms were acquired over 2 heartbeats during 1 breath-hold period. Images were reconstructed with a 160×256 matrix. The black-blood pulse suppressed the signal in the cavity. Other parameters were bandwidth 650 Hz/pixel, effective echo time 43 ms, interecho spacing 4.3 ms, FOV 225×300 mm, and slice thickness 6 mm. The total acquisition time was 25 seconds for scout images, 120 seconds for T1w short-axis images, and 40 seconds for T2w HASTE acquisitions. The whole in vivo procedure in the magnet took 15 to 25 minutes.

Postmortem MRI was performed with a high-resolution coil. A T1w spin-echo sequence with TR/TE of 450 ms/12 ms, FOV of 75×100 mm, 192×256 matrix, and 2 signal averages was used. T2w images were obtained with a fast spin-echo sequence (echo train length, 5). Other parameters were TR/TE, 3000 ms/45 and 90 ms; FOV, 68×136 mm; matrix, 120×256; and 4 signal averages. For both T1w and T2w images, the slice thickness was 2 mm, and 40 slices were acquired in 2 interleaved scans. The total acquisition times were roughly 3 minutes for T1w imaging and 5 minutes for T2w imaging.

SI was measured with the Adobe Photoshop package and validated by direct comparison with the built-in software on the MR scanner. From this, we were able to calculate the mean SI for the whole enhanced area (EA). Contrast ratio (CR) was calculated by dividing the mean SI of the EA by the mean SI in a normal region of the LV.

Planimetric Measurements

Planimetry was performed with Adobe Photoshop 4.0 software. LV area (LVA) (on MR images and TTC-stained slices), infarct area (IA) (on TTC-stained slices), and in vivo and postmortem EA (on MR images) were measured, and the results were converted from pixels squared to millimeters squared. To evaluate infarct volume (IV), enhanced volume (EV), and left ventricular volume (LVV), we first calculated the slice averages (arithmetic mean of IA, EA, and LVA on the 2 sides of the TTC-stained slices and corresponding MR images). The volumes in 1 slice were obtained by multiplying the corresponding average by the slice thickness. The total volumes (IV, EV, and LVV) were calculated by summation of all slice volumes of 1 heart. The delineation of the LV cavity was not possible on in vivo T1w images; therefore, LVA was calculated only on T2w images for in vivo MRI. The enhanced regions on MRI were compared with the TTC-labeled infarct regions in 2 ways: slice-by-slice (IA and EA were compared; 1 slice gives 1 data point) and globally (IV and EV were compared; 1 heart gives 1 data point).

Statistical Analysis

The agreement between MRI and TTC measurements of infarct size was assessed with the methodology of Bland and Altman. The SI of EAs and of nonenhanced regions was compared by paired Student’s t tests; the CRs in the 3 groups of dogs were compared by unpaired Student’s t tests. A value of P<0.05 was considered signiﬁcant. The normal distribution was tested with the Shapiro-Wilk statistic, and logarithmic transformations were used when appropriate. All calculations were performed with the SAS procedures GLM, UNIVARIATE, and TTEST. Data are presented as mean±SD.

Results

A total of 32 experiments were performed. Twelve dogs died before MRI (during thrombolysis, 6; not recovered from anesthesia, 2; during the postinfarction period, 4) and were excluded. Initially, 18 dogs were divided into 3 groups of 6. The group allocation was based on the availability of the catheterization and MRI laboratories. This nonrandomized design resulted in a selection bias (see below); therefore, 2 more dogs were added to group C. Thrombolysis induced reperfusion in 17 dogs; reocclusion occurred in 6 dogs during the initial 2 hours of therapy. All dogs showed patent LADs 1 hour after removal of the copper coil. T1w and T2w MR images of 189 in vivo slices, of 800 postmortem slices, and
189 pictures of the TTC-stained LV slices were obtained. In 7 dogs, the first apical slice on in vivo MRI was excluded from the analysis because of artifacts (partial volume effects with fatty tissue, poor delineation of the ventricle). The remaining 182 slices were used for the comparison of in vivo MRI and TTC staining. Of the 800 postmortem slices, only the 182 images corresponding to the TTC-stained slices were selected for comparisons (Figure 2).

Anatomic Infarct Size
Planimetry of the TTC-stained slices showed a broad range for infarct size (0% to 98% of LVA, 0% to 36% of LVV). Eight dogs had infarctions of <5% of LVV. Infarcted zones were present on 134 of 182 slices. The infarcts were significantly smaller in group C than in group B (Table 1).

MRI Evaluation of Infarct Size
The results of MRI evaluation of infarct size are summarized in Tables 1 and 2. T1w EAs were observed on 118 slices at in vivo MRI and on 147 slices at postmortem MRI. Both slice-by-slice and global analyses showed an excellent correspondence between T1w images and TTC staining with regard to the quantification of infarct size (correlation coefficients were $r^2=0.98$ between in vivo MRI and TTC and $r^2=0.99$ between postmortem MRI and TTC; $P<0.0001$ for both). The limits of agreement calculated with the methodology of Bland and Altman were narrow; the mean difference between TTC and MRI measurements of infarct size was almost zero (Table 2, Figure 3). No significant differences between groups were observed for the TTC-MRI agreement.

The contrast between infarcted and normal myocardium in black-blood images was poor; these images were used only

<table>
<thead>
<tr>
<th>Group</th>
<th>TTC</th>
<th>In Vivo T1w</th>
<th>Postmortem T1w</th>
<th>Postmortem T2w</th>
</tr>
</thead>
<tbody>
<tr>
<td>A (n=6)</td>
<td>10.8±11.6</td>
<td>12.3±11.5</td>
<td>11.4±11.7</td>
<td>18.7±16.7*</td>
</tr>
<tr>
<td>B (n=6)</td>
<td>22.4±11.7</td>
<td>24.3±12.6</td>
<td>24.9±12.9</td>
<td>33.6±16.4*</td>
</tr>
<tr>
<td>C (n=8)</td>
<td>5.1±9.3†</td>
<td>5.7±9.3</td>
<td>5.0±8.2</td>
<td>10.9±10.8*</td>
</tr>
<tr>
<td>Total (n=20)</td>
<td>12.0±12.3</td>
<td>12.9±13.0</td>
<td>12.9±13.3</td>
<td>20.0±16.8*</td>
</tr>
<tr>
<td>CR, %</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A (n=6)</td>
<td>...</td>
<td>168.8±29.1</td>
<td>139.2±11.3</td>
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</tr>
<tr>
<td>B (n=6)</td>
<td>...</td>
<td>176.7±35.2</td>
<td>137.6±13.2</td>
<td>...</td>
</tr>
<tr>
<td>C (n=8)</td>
<td>...</td>
<td>149.8±20.6†</td>
<td>123.2±12.9</td>
<td>...</td>
</tr>
<tr>
<td>Total (n=20)</td>
<td>...</td>
<td>166.9±31.9</td>
<td>133.3±14.2</td>
<td>...</td>
</tr>
</tbody>
</table>

Data are presented as mean±SD.

*P<0.05 vs TTC measurements; †P<0.05 vs group B; ‡P<0.05 vs groups A and B.
for LVA measurements. Postmortem T2w images showed the presence of myocardial injury. Both hyperintense and hypointense areas were present in most of these images; when these areas were summed, the results systematically overestimated the TTC-measured infarct size by 8.1±5.4% of LVV (range, 1.2% to 18.3%; \( P<0.0001 \)). The interval of agreement with the TTC technique was large for T2w imaging.

**Discussion**

**Evaluation of Infarct Size on Gadophrin-2–Enhanced Images**

This report represents the first study of infarct-size measurements with gadophrin-2–enhanced MRI after thrombolytic therapy. The experimental design mimicked the clinical setting of an acute MI treated with thrombolysis. Furthermore, MRI was performed at different intervals, corresponding to the time points at which patients are generally stabilized and can be discharged from the coronary care unit (2 and 3 days after infarction) or from the hospital (7 days after infarction).

The results of this study support our previous findings that gadophrin-2–enhanced MRI is a reliable, noninvasive method for infarct-size evaluation.\(^{12–16}\) We were able to locate and quantify infarcts as small as 1% or as large as 36% of LVV with similar accuracy. The methodology used allows systematic in vivo quantification of the infarct size: the LV was completely imaged by 9 to 11 parallel slices, 96% of the images being included in the analysis. The mean difference between TTC-labeled infarct size and gadophrin-2 enhanced areas in T1w images was almost zero, with limits of agreement of less than ±5% of the LV for both slice-by-slice and global comparison, indicating a high accuracy of the MRI measurements.

The gadophrin-2–induced enhancement of necrosis was good, with an in vivo CR averaging 166.9%. The CR was similar in dogs monitored 2 or 3 days after acute MI but significantly lower in dogs monitored after 7 days. The lower uptake of gadophrin-2 may be related to the process of healing, during which denatured tissue components responsible for contrast “trapping”\(^{10–13}\) are progressively replaced by scar. This hypothesis is supported by the findings of a lack of enhancement with gadophrin-2 in the presence of strong enhancement with Gd-DTPA 1 month after infarction in pigs,\(^{22}\) indicating that gadophrin-2 is a sensitive marker of necrosis in the early but not in the late stages of MI.

MRI was performed 24 hours after gadophrin-2 administration for a number of reasons. First, early imaging after gadophrin-2 was already reported.\(^{15}\) Second, a protocol with

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**TABLE 2. Agreement Between MRI- and TTC-based Measurements of Infarct Size**

<table>
<thead>
<tr>
<th>Analysis and Group</th>
<th>In Vivo T1w</th>
<th>Postmortem T1w</th>
<th>Postmortem T2w</th>
</tr>
</thead>
<tbody>
<tr>
<td>Slice-by-slice</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean difference MRI-TTC, % of LVA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A (n=55)</td>
<td>-0.9±4.5</td>
<td>0.2±2.3</td>
<td>8.7±10.8</td>
</tr>
<tr>
<td>B (n=54)</td>
<td>-0.9±2.9</td>
<td>0.3±2.3</td>
<td>7.0±6.5</td>
</tr>
<tr>
<td>C (n=73)</td>
<td>0.2±2.1</td>
<td>-0.2±1.0</td>
<td>6.1±7.0</td>
</tr>
<tr>
<td>Total (n=182)</td>
<td>-0.1±2.4</td>
<td>0.0±2.0</td>
<td>7.3±8.1</td>
</tr>
<tr>
<td>Limits of agreement MRI-TTC, % of LVA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A (n=55)</td>
<td>(-9.7, +8.0)</td>
<td>(-4.5, +4.8)</td>
<td>(-12.9, +30.3)</td>
</tr>
<tr>
<td>B (n=54)</td>
<td>(-6.6, +4.8)</td>
<td>(-4.3, +4.8)</td>
<td>(-6.1, +20.0)</td>
</tr>
<tr>
<td>C (n=73)</td>
<td>(-3.9, +4.4)</td>
<td>(-2.3, +1.9)</td>
<td>(-7.9, +20.2)</td>
</tr>
<tr>
<td>Total (n=182)</td>
<td>(-4.7, +4.9)</td>
<td>(-3.9, +4.0)</td>
<td>(-8.9, +23.5)</td>
</tr>
<tr>
<td>Global</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean difference MRI-TTC, % of LVV</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A (n=6)</td>
<td>-0.5±1.2</td>
<td>0.7±0.9</td>
<td>8.0±6.5</td>
</tr>
<tr>
<td>B (n=6)</td>
<td>0.9±3.0</td>
<td>1.6±1.4</td>
<td>8.6±4.8</td>
</tr>
<tr>
<td>C (n=8)</td>
<td>0.6±1.1</td>
<td>-0.1±0.5</td>
<td>5.9±4.4</td>
</tr>
<tr>
<td>Total (n=20)</td>
<td>0.9±2.0</td>
<td>0.9±1.4</td>
<td>8.1±5.4</td>
</tr>
<tr>
<td>Limits of agreement MRI-TTC, % of LVV</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>A (n=6)</td>
<td>(-1.9, +2.9)</td>
<td>(-1.1, +2.5)</td>
<td>(-5.0, +21.0)</td>
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<tr>
<td>B (n=6)</td>
<td>(-5.1, +6.9)</td>
<td>(-1.2, +4.4)</td>
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<td>(-2.9, +14.6)</td>
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<td>Total (n=20)</td>
<td>(-3.0, +4.9)</td>
<td>(-1.9, +3.8)</td>
<td>(-2.8, +18.9)</td>
</tr>
</tbody>
</table>

\( n \) indicates number of slices or hearts. Data are presented as mean±SD and as intervals. The limits of agreement are given within parentheses.
both early and delayed imaging would not allow a period of recovery for the animals, especially in group A. Finally, and most importantly, considering that the highest mortality rates after acute MI are observed during the first 24 hours and that the present technology does not allow monitoring of the patient in the MRI laboratory at standards comparable to those in a coronary care unit, a pragmatic solution is to evaluate infarct size with MRI outside this critical period. Because of its persistence in the necrotic tissue, gadophrin-2 may offer the unique possibility to obtain a “fingerprint” of the infarct size as it was at the time of administration (i.e., early after thrombolysis or direct angioplasty) with an MRI study performed as soon as the patient is stabilized. Furthermore, although an enhancement of the infarcts can be observed as early as 1 to 2 hours after injection, the best images are obtained after the agent is cleared from the blood.

The mechanisms responsible for contrast trapping in the areas of necrosis remain to be elucidated. The observation that gadophrin-2 enhancement of MI persists for a long period despite complete clearance from the blood (in an unpublished experiment, the enhancement was observed during the first 2 weeks but was lost at 4 weeks) suggests a binding or precipitation of gadophrin-2 in the necrotic areas. We have previously shown that the gadophrin-2 content is 10 times higher in infarcted than in normal myocardium. As for safety, no alterations were observed in the ECG, LV or aortic pressure, or dP/dt during intracoronary administration of gadophrin-2 in a previous experiment (unpublished data). Other studies with metalloporphyrins also suggest that these agents are stable compounds. The only known toxic effects are skin discoloration and photosensitization due to uncomplexed porphyrins, but these are reduced by chelation of a metal. The LD₅₀ of gadophrin-2 in rats is comparable with that of Gd-DTPA. However, additional animal studies are needed before the agent can be used in patients.

**Evaluation of Infarct Size on T2w Images**

Delineation of infarcts on T2w HASTE images was in practice not feasible. On postmortem images obtained with a high-resolution coil, T2w imaging did not accurately estimate infarct size. The analysis was complicated by the simultaneous presence of hyperintense and hypointense areas on the same slice, by the smaller CR, and by the difficult delineation. Typically, areas with increased SI at the borders and decreased SI in the center were observed (“hyperintense” and “hypointense” were defined in comparison to the SI of normal myocardium). The hypointensity may be related to the presence of gadophrin-2 and/or hemoglobin in the center of necrosis. Like those of previous reports, our results show that T2w imaging overestimates the infarction in the first week after acute MI.

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**Figure 3. Agreement between MRI and TTC.** Top, Plots of enhanced volumes (as percentage of LV) on MRI versus TTC-labeled infarct volumes (as percentage of LV). Line of identity is presented. Bottom, Plots of differences between methods against their mean; horizontal lines represent limits of agreement between TTC and MRI evaluations of infarct size.
Potential Advantages of Gadophrin-2 Over Gd-DTPA

There are several potential advantages of gadophrin-2– over Gd-DTPA– enhanced MRI in the clinical settings of acute MI. Gadophrin-2 is an agent with high affinity for necrosis\(^{12–15}\) and therefore should be able to distinguish between irreversibly and reversibly injured myocardium. Good correlations were also reported between infarct size and Gd-DTPA imaging\(^{25,26}\), but an analysis of agreement was not performed. Therefore, it is not known how much the quantification of infarct size with TTC staining and Gd-DTPA– enhanced MRI differ. Furthermore, Gd-DTPA allows only a narrow time window for MRI after contrast delivery. It was shown that infarct size is overestimated on MR images obtained early after Gd-DTPA injection\(^{25–27}\). The contrast agent is then rapidly washed out, with a speed depending on many local factors. In contrast, gadophrin-2 uptake into the necrotic area is stable, and our study shows that MRI measurements of infarct size are accurate 24 hours after injection. We have previously reported the same accuracy for early imaging after intracoronary injection of a minimal dose of gadophrin-2\(^{16}\).

Clinical Implications

Our results show that gadophrin-2– enhanced MRI can be used as a reliable noninvasive method to distinguish necrotic from ischemic and normal myocardium. This methodology may allow a better understanding of complex processes, such as stunning, hibernation, and infarct healing. Furthermore, considering that MRI can provide anatomic and functional information within a single imaging session, it may be very useful for the evaluation of the effects of reperfusion and the need for additional revascularization. Furthermore, in vivo infarct-size measurements may be a valuable efficacy end point in clinical trials of new reperfusion strategies.

Study Limitations

Our study has several limitations. First, LVA was calculated only from T2w images for in vivo MRI. Second, the TTC staining was not confirmed by histology. Third, the infarcts were significantly smaller in group C, probably as a result of a selection bias (dogs with severe hemodynamic impairment at the end of thrombolysis were generally not assigned to group C because of lower chances of survival). However, after we added 2 dogs to this group, the infarct size range (0% to 24%) was comparable to those of groups A (0.5% to 31%) and B (0.6% to 36%). It is difficult to assess whether changes in gadophrin-2 uptake over time or changes in the intrinsic properties of the necrotic area were responsible for the lower CR values in group C. Fourth, LAD patency was not systematically evaluated at the time of MRI for practical reasons (availability of the catheterization laboratory). Nevertheless, the homogeneous enhancement pattern observed on T1w images suggests that the LAD was patent at the time of gadophrin-2 injection in all dogs. Furthermore, the LAD was patent 1 hour after removal of the thrombogenic copper coil in all animals. Finally, no dogs with persistent occlusion were studied with this protocol. Thus, no conclusion can be drawn regarding the accuracy of the method for nonreperfused infarcts.

Conclusions

In this canine model, in vivo gadophrin-2– enhanced MRI could precisely evaluate infarct size after thrombolysis. The

Figure 4. A, Black-blood preparation pulse. A nonselective inversion recovery pulse is followed by a selective 180° pulse. Imaging starts after a time delay, chosen at zero crossing of signal in cavity. B, Black-blood single-shot HASTE technique starts with a black-blood preparation pulse and is followed by a long echo train. The k space is partially filled.
contrast-induced enhancement was lower after 1 week. This technique may be very useful in the evaluation of infarct size after reperfusion in patients with an acute MI.

Appendix

The black-blood HASTE technique is an ultrafast MR sequence that combines a preparation pulse to suppress possible artifacts from flow in the cavity with an ultrafast acquisition scheme. The preparation pulse consists of a nonselective 180° inversion recovery pulse followed by a slice-selective 180° pulse (Figure 4A). The succession of these 2 pulses has no effect on the myocardial tissue. The spins of the blood in the cavity are in general not completely realigned with the axis of the main magnetic field due to flow into or out of the slice. A time delay (TI) is kept between the inversion recovery pulses and the start of the acquisition process. Ideally, its timing must be adjusted to the zero crossing of the signal in the cavity. Usually, TI is set to 600 ms, and it may be further adjusted in case of an incomplete suppression of the cavity signal.

The black-blood pulse is followed by a fast spin-echo acquisition with a long echo train. In the single-shot HASTE technique, 1 echo train fills half of the k space plus 8 additional lines (Figure 4B). The segmented HASTE uses 2 preparation pulses followed by echo trains that fill the same k space. The effective echo time is 43 ms. Images have a moderate T2 weighting. A preliminary study in patients has shown that this particular acquisition is very robust.28

Acknowledgments

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References

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