Part III: Molecular Basis of Cardiac Electrophysiology and Arrhythmias

In Parts I and II of this article, we discussed monogenic arrhythmic disorders. These are determined or favored by an inborn alteration and for the most part are characterized by a single genetic alteration. This has allowed the use of “paradigms”; namely, diseases, such as the long-QT syndrome (LQTS), in which it has been possible to trace specific mutations on ion channel genes to their electrophysiological consequences in the patient. Unfortunately for the practicing cardiologist, these “simple” diseases constitute only a small part of the clinical conditions associated with cardiac arrhythmias. The majority of cases affect patients in whom the arrhythmogenic substrate is complex. Indeed, the expression of the molecular systems responsible for normal and abnormal electrical activity vary significantly, depending on a variety of factors, including age, regional factors (type of cells, myocardial perfusion), and such underlying chronic diseases as cardiac hypertrophy, myocardial infarction, and heart failure.

The study of this complex system of interacting molecular functions requires an approach somewhat different from that required to consider monogenic disease. Accordingly, in this section we discuss broader themes that are essential to understand the integration of gene expression, ion channel function, and cell coupling in multicellular networks as a first step toward the comprehension of more frequent and more complex arrhythmogenic conditions.

Diversity of Gene Expression in the Heart

Understanding cell-to-cell variability in the cardiac action potential shape and the mechanisms underlying impulse propagation is the key to understanding normal and abnormal cardiac electrophysiology. Much of this variability can be attributed to variability in the characteristics of individual ion currents whose integrated behavior determines the shape and duration of action potentials in individual cardiac cells, as well as to variability in cell-to-cell communications. Ion currents are now recognized to flow through specific pore-forming membrane proteins called ion channels. The first gene encoding an ion channel protein was cloned in 1984, and the succeeding decade and a half has seen the cloning of genes encoding most ion channels expressed in heart and in many other tissues. Many of the proteins these genes encode share common structures and can be viewed as members of the same superfamily. For example, Figure 3 shows the tremendous diversity of mammalian genes that make up the family of potassium channel genes. Because potassium channels are made up of 4 ion channel α-subunits, which are not necessarily identical, the potential for diversity in potassium currents is even greater than shown. This is further compounded by the identification of ancillary subunits (the products of different genes) that can assemble with potassium channel tetramers to modulate their function. Figure 4 illustrates the major ion currents in heart and the genes whose protein products are thought to form...
their structural basis. The dramatic increase in molecular genetic information underlying cardiac function is not confined to ion channels but rather has extended to multiple other genes, including those controlling cell-to-cell communication (the connexins, Cx), the contractile apparatus, and cardiac development, to name a few. With this cloning effort have come important advances not only in understanding mechanisms of normal cardiac function but also in new insights into the mechanisms underlying common cardiac diseases and their therapy. A common method of studying individual cardiac ion channel gene function is to express the gene of interest in noncardiac (heterologous) study systems, such as mammalian cell lines or the eggs of the African clawed toad, *Xenopus laevis*. In some cases, expression of a single gene in such heterologous systems is sufficient to reproduce the physiological and pharmacological characteristics of a specific cardiac ion current; HERG expression to recapitulate $I_{Kr}$ is an example, although coexpression of the minK subunit may increase $I_{Kr}$ amplitude. The systems have been especially valuable in delineating the functional consequences of ion channel gene mutations, although it should be recognized that mechanisms other than a simple dominant negative effect on channel gating (eg, altered trafficking) may also play a role.

In other cases, faithful recapitulation of a specific cardiac ion current requires coexpression of >1 gene. Heterotetramers of *Kv4.2* and *Kv4.3* may determine the $I_{Kr}$ in some

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**Figure 3.** Family tree of pore-forming K⁺ channel subunits in mammals. Dendrogram represents degree of similarity between each gene, as determined by ClustalW alignment program. Sequences were extracted from Genbank database and correspond exclusively to rodent or human channels. K⁺ channel subunits are divided into 3 major groups according to their structural and functional properties as shown in blue, green, and red boxes. Major genes expressed in heart are indicated in gray boxes, together with probable current they underlie.
species. Other examples include coexpression of a structural gene and an ancillary subunit; one good example is the finding that coexpression of \textit{KvLQT1}, a member of the potassium channel family shown in Figure 3, with the \textit{minK} gene is required to recapitulate \textit{I}_kr. Another example is the \textit{\alpha-\beta} interaction during the development of an adult sodium current as described below. In yet other cases, a gene product (eg, \textit{Kv2.1}) can be detected in heart without a recognized counterpart among the known ion currents, and ion currents (\textit{I}_L) remain for which no corresponding gene has yet been identified. One very important observation is that ion channel genes are virtually never expressed exclusively in the heart. Thus, mutations or blocking drugs may affect not only cardiac function but also function in other organs. The best-described example to date is the deafness displayed by patients with the recessive (Jervell and Lange-Nielsen) form of LQTS.4 This arises because the 2 genes involved in the disease, \textit{KvLQT1} and \textit{minK}, are expressed not only in the heart but also in the inner ear, where together they control endolymph homeostasis. It is not yet known whether parent carriers, who have mild, usually but not always asymptomatic mutations in \textit{KvLQT1} or \textit{minK}, display subtle defects in hearing. This is but one example of the potential for a molecular genetic explanation for a diversity of symptoms through common mutations affecting function in multiple organs.

**Development**

One well-recognized form of variability in cardiac function is the stereotypical changes that are observed during development. Much of the information has been gathered in small rodents and may not be directly applicable to humans, but it may be important because a common form of cardiac response to injury is regression to a fetal phenotype. Whether, for example, the electrophysiological changes associated with hypertrophy (eg, in patients with hypertension or heart failure) represent such a patterned response is an important consideration. Understanding the mechanisms underlying such a change in phenotype may be an important step in the prevention of arrhythmias in these common acquired disorders of cardiac function.

The earliest stage at which ion currents have been recorded from heart tissue is embryonic (postcoital, pc) day 11 in mouse (normal gestation period, 20.5 days). At this stage, the predominant inward current is \textit{I}_Ca(L), and the predominant outward current is the rapidly activating component of the delayed rectifier, \textit{I}_kr. Sodium current \textit{I}_Na appears later and increases markedly just before birth. There are important differences between sodium current recorded in neonatal animals and those recorded in adult animals: \textit{I}_Na in neonates is smaller; it activates, inactivates, and recovers from inactivation more slowly than that in adults; and it has a more positive voltage dependence of inactivation than that in adults. Some data suggest that this difference between neonatal and adult sodium current may reflect expression of a \textit{\beta}-subunit and/or \textit{\alpha-\beta} assembly to produce the “mature” phenotype and that this change may reflect the sympathetic innervation of the heart that occurs around the time of birth. This may be but one example of a more general influence of sympathetic innervation as a modulator of cardiac electrophysiology. It is likely that multiple mechanisms will be identified. The possibility that regional cardiac denervation may play a role in acquired diseases (eg, myocardial infarction) is an obvious one that requires further study. Another intriguing observation during development is the consistent embryotoxicity of specific \textit{I}_Kr blockers, such as dofetilide or almokalant, in the rat. Because \textit{I}_Kr is the predominant (if not the sole) repolarizing current at this stage, it has been postulated that embryotoxicity is due to failure of cardiac repolarization, with death due to arrhythmias secondary to triggered activity (which have been demonstrated under these experimental conditions) or simply membrane depolarization. Whether similar considerations apply to humans has not been determined.

The pattern of expression of the connexins, which form the gap junction channels and belong to one gene family, also varies during development. \textit{Cx43} mRNA is detectable in mouse heart from day 9.5 pc. Initially, it is expressed only in ventricle, but later it spreads throughout the whole heart. Two weeks after birth, the message starts to diminish again to a steady level that is maintained during adulthood. \textit{Cx40} is detected from 9.5 days pc. The message is initially confined to atrium and left ventricle, but during development it spreads throughout the whole heart. After 14 days pc, \textit{Cx40} message starts to diminish in the ventricles from epicardial to endocardial until in the adult, \textit{Cx40} is restricted to both atria and the proximal conduction system. \textit{Cx45} mRNA is expressed from day 11 pc at a constant level throughout the whole heart until week 3 postpartum, when it starts to decrease until in adulthood, only low-level expression in the proximal conduction system is detected.

**Regional Diversity in Cardiac Electrophysiology**

Although heterogeneity of cardiac electrophysiology is increasingly recognized as a contributor to cardiac arrhythmias, it should also be recognized that there is substantial heterogeneity in the electrophysiological properties of individual cells even under physiological conditions. A trivial example is the differences among the electrophysiological properties of sinoatrial node, atrium, AV node, conducting system, and ventricular myocardium. These differences presumably reflect variability in expression and/or function of the repertoire of ion channels, whose integrated activity determines the distinctive action potentials in each of these regions. More recently, it is increasingly recognized that there is considerable potential for cell-to-cell variability in action potentials and gene expression within such specified regions. For example, a survey of atrial myocytes revealed a consistent \textit{I}_Na only in ~60% of cells, a consistent \textit{I}_K in ~15% of cells, and both currents in 30% of cells. Studies of mRNA expression have also demonstrated striking cell-to-cell variability in expression of individual ion channel genes. Two LQTS genes, \textit{HERG} and \textit{KvLQT1}, were identified in a majority of cells in most regions. In contrast, \textit{minK} was most abundant in sinoatrial node (in ~33% of cells) but was much less abundant in ventricular muscle cells (10% to 29%). This is consistent with a more recent report that, at least in the mouse, \textit{minK} expression appears to be restricted largely to the
Electrophysiological studies have identified Kv4.2 and/or 4.3 (depending on the species) as the ion channel gene whose expression in heterologous systems results in a current most closely resembling human \( I_{\text{kr}} \). One of the important features of human \( I_{\text{kr}} \) is its usually rapid recovery from inactivation. It was this observation that first raised the suggestion that \( Kv1.4 \), an initial leading candidate for \( I_{\text{kr}} \), might not, in fact, encode this current, because \( Kv1.4 \) recovers very slowly from inactivation. Interestingly, the human endocardium also displays an \( I_{\text{kr}} \), but one that, unlike that recorded in epicardium, recovers from inactivation very slowly and is therefore not regularly observed in endocardial action potentials. It is this observation that first raised the suggestion that \( Kv1.4 \), an initial leading candidate for \( I_{\text{kr}} \), might not, in fact, encode this current, because \( Kv1.4 \) recovers very slowly from inactivation. It was this observation that first raised the suggestion that \( Kv1.4 \), an initial leading candidate for \( I_{\text{kr}} \), might not, in fact, encode this current, because \( Kv1.4 \) recovers very slowly from inactivation.

The connexins are also expressed in a chamber-specific and tissue-specific fashion. In heart, Cx40, Cx43, and Cx45 have been detected at the protein level. The phosphoprotein Cx43 is the most abundant cardiac connexin. It forms gap junctional channels with a main conductance of 45 pS between cardiac myocytes in all parts of the heart, with the possible exception of sinoatrial and AV nodes. In ventricle, Cx43 is more abundant in the intercalated disk than in the lateral cell borders, which partially explains anisotropic impulse conduction. In atrium (with the exception of the crista terminalis), the difference between end-to-end and side-to-side connections is much less pronounced. Although Cx43 has been reported to be present in sinoatrial node, atrial cells in the presence of sinoatrial and AV nodes. In ventricle, Cx43 is more abundant in the intercalated disk than in the lateral cell borders, which partially explains anisotropic impulse conduction. In atrium (with the exception of the crista terminalis), the difference between end-to-end and side-to-side connections is much less pronounced. Although Cx43 has been reported to be present in sinoatrial node, atrial cells in the presence of sinoatrial and AV nodes. In ventricle, Cx43 is more abundant in the intercalated disk than in the lateral cell borders, which partially explains anisotropic impulse conduction. In atrium (with the exception of the crista terminalis), the difference between end-to-end and side-to-side connections is much less pronounced. Although Cx43 has been reported to be present in sinoatrial node, atrial cells in the presence of sinoatrial and AV nodes.


diagram.png

Figure 5. Schematic of a computer model used to simulate electrical activity of a cardiac ventricular cell. Cellular environment is highly interactive and modulates behavior of membrane ion channels. Integrated behavior of cell generates action potential. \( I_{\text{Na,b}} \) indicates fast sodium current; \( I_{\text{Ca,b}} \), calcium current through L-type calcium channels; \( I_{\text{Ca,L}} \), calcium current through T-type calcium channels; \( I_{\text{Kp}} \), fast component of delayed rectifier potassium current; \( I_{\text{Kr}} \), slow component of delayed rectifier potassium current; \( I_{\text{K1}} \), inward rectifier potassium current; \( I_{\text{Ks}} \), plateau potassium current; \( I_{\text{KATP}} \), ATP-sensitive potassium current; \( I_{\text{K1ak}} \), sodium-potassium pump current; \( I_{\text{Caex}} \), sodium-calcium exchange current; \( I_{\text{Ca}} \), calcium pump in sarcolemma; \( I_{\text{Na,b}} \), sodium background current; \( I_{\text{Ca,b}} \), calcium background current; \( I_{\text{Ca,Ca}} \), nonspecific calcium-activated current; \( I_{\text{Ca}} \), calcium uptake from myoplasm to network sarcoplasmic reticulum (NSR); \( I_{\text{CaL}} \), calcium release from junctional sarcoplasmic reticulum (JSR); \( I_{\text{Ca,L}} \), calcium leakage from NSR to myoplasm; and \( I_{\text{Ca}} \), calcium translocation from NSR to JSR. Calmodulin, troponin, and calsequestrin are calcium buffers.

Integration of Ion Channel Function Into the Cellular Environment

Although ionic channels and connexins participate in the generation of the cardiac action potential and in cell-to-cell communication, it is important to recognize that experimentally, single-channel data are obtained primarily in preparations that are removed from the cardiac cell environment (eg, membrane patches, cloned channels in \textit{Xenopus} oocytes). Figure 5 is a schematic of a cardiac ventricular myocyte, demonstrating the complex physiological environment in which ion channels function to generate an action potential. The cellular environment is highly interactive and modulates the behavior of the single channel through interactions with other channels or with the ionic milieu of the cell. An example is illustrated in the diagram in Figure 6. In this scheme, the \( I_{\text{Ca,L}} \) induces \( \text{Ca}^{2+} \) release from the sarcoplasmic reticulum through the \( \text{Ca}^{2+} \)-induced \( \text{Ca}^{2+} \) release process. The released \( \text{Ca}^{2+} \) in the myoplasm, in turn, modulates several ionic currents (including \( I_{\text{Ca,L}} \), itself). In Figure 6, the myoplasmic \( \text{Ca}^{2+} \) is shown to increase the
The conductance of $I_{Na}$ to drive $I_{NaCa}$ for the purpose of Ca$^{2+}$ extrusion, and to participate in the inactivation of $I_{CaL}$ results in decreased action potential duration (APD), as does reduced $I_{Ks}$, $I_{CaL}$, $I_{NaCa}$ is an inward current (when operating to extrude Ca$^{2+}$ from the cell), and its augmenting acts to prolong APD. The net effect depends on the quantitative balance of these processes. This balance can depend on the basal expression of these channels or of the proteins (such as kinases) that regulate their activity, diseases or other processes that modulate the expression, and other important factors, such as rate or adrenergic activity. Importantly, many of these latter processes may also be modulated by changes in intracellular calcium.

The effect on the action potential is complicated; increased $I_{Ks}$ acts to shorten the action potential duration (APD), as does reduced $I_{CaL}$, $I_{NaCa}$. It is conceivable that such conditions are created by some mutations but not by others. It has been successfully applied to describe global effects of inhibitors of Na$^{+}$ channels, 145 and of hypoxia/anoxia 146 on conduction. However, in many instances, more complicated models are needed to describe the conduction process appropriately.

### Ion Channel Function in Multicellular Networks

The simplest model used to explain cardiac electrical propagation was originally derived from conduction models developed for the nervous system. In this model, cardiac cells are merged into a syncytium-like conducting structure composed of a cell membrane carrying the ion channels that generate the action potential and a single continuous intracellular space (in this space, the electrical conductances of the cytoplasm and the gap junctional connexons are merged together). Many important insights into the electrical propagation process have been derived from this so-called “continuous” model. 143

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### Interaction Between Cell-to-Cell Coupling and Ion Channel Function.

A first step in describing the relation between electrical propagation and cardiac structure more closely involved simulation of single-cell chains with interconnections representing the gap junctions. 147 Although the properties of such models are not markedly different from the “continuous” model when electrical cell-to-cell coupling is normal, a distinctly different behavior is unmasked once the electrical coupling of the cells diminishes. Interestingly, this behavior includes feedback interaction between cell-to-cell coupling and ion channel function. Partial closure of connexins (ischemia, hypoxia 145,146) or a decrease in expression (heart failure 148) of connexins might affect conduction in several ways. First, conduction velocity decreases to a much greater extent than with inhibition of ion channels 149 and can reach a few centimeters per second or be even slower. Accordingly, reentrant arrhythmias that occur in partially uncoupled tissue can form circulating excitation of very small dimensions (microreentry). By contrast, simulations suggest that depressed $I_{Na}$ alone cannot account for this phenomenon. Second, a change in cell-to-cell coupling feeds back on the way...
ionic channels in the membrane are activated and on the role of ionic channels in conduction (and most probably, repolarization). This is illustrated in Figure 7, which depicts a row of simulated cells in a state of advanced cell-to-cell uncoupling. Because of the high degree of discontinuity that is introduced by the uncoupled cells, there is a long conduction delay between the cells. Because the action potential in the driver cell has to furnish local electrical circuit current to the driven cell, this current has to flow as long as the driven cell has not reached its threshold for depolarization, ie, for activation of the Na\(^+\) inward current. In the normal case of propagation, the conduction delay between 2 cells is very short. During this time, the Na\(^+\) channels in the membrane are activated and on the role of ionic channels in conduction and of Ca\(^2+\) inward current–dependent, merely by a change in the degree of cell-to-cell coupling.\(^{149,150}\) Feedback interactions between electrical coupling and ionic current flow are also expected to occur during the action potential plateau and during repolarization. Because of the small ion current density and the high membrane resistance during the action potential plateau,\(^{151}\) the electrical interactions between adjacent tissue regions are stronger and extend over longer distances during this phase than during rest or during the action potential upstroke. As a consequence, differences in intrinsic APDs among adjacent tissue regions will be relatively small during normal cell-to-cell coupling and will be unmasked during uncoupling. In this way, the reduction of electrotonic interaction by cell-to-cell uncoupling is predicted to unmask intrinsic heterogeneities in repolarization, with both events serving to promote reentrant arrhythmias.

**Interaction Between Tissue Structure and Ion Channel Function**

As with the discrete pattern of gap junctions at the microscopic level, the macroscopic cardiac structure introduces obstacles and discontinuities for propagating electrical waves. Typical examples of discontinuities are branching fibers (Purkinje system, atrial trabeculae) and/or connective tissue layers (remodeled discontinuities are branching fibers (Purkinje system, atrial trabeculae) and/or connective tissue layers (remodeled, ventricular trabeculae in aging myocardium\(^{153}\)). Such discontinuities have been shown to affect membrane channel function. In any situation in which electrical waves propagate through anatomically discontinuous tissue and emerge from an isthmus,\(^{154}\) turn around the end of an obstacle,\(^{155}\) or emerge from a small fiber into a large tissue mass,\(^{156,157}\) the propagating wave becomes curved. In a curved wave front, the mismatch between the excitatory local current produced by the excited cells upstream and the electrical load of the nonexcited cells ahead of the wave front downstream affects both conduction velocity and the activation of ion currents. At a convex wave front, this mismatch will lead to a local conduction delay and to a localized increase of the amount of depolarizing inward current.\(^{156}\) Conversely, at a concave wave front (or, similarly, at a site of collision), conduction velocity will locally increase and the inward current will be locally reduced.\(^{158}\) This dependence of ion current activation on wave-front curvature is likely to be responsible for the larger effect of inhibitors that bind to open Na\(^+\) channels at sites at which the propagation wave is convex than at sites at which it is linear.\(^{159}\) The curvature of the wave front will also play an important role in determining the intrinsic heterogeneities involved in impulse propagation. Thus, at divergence points, if the wave front is markedly curved, large local conduction delays result. As with the case of advanced cell-to-cell uncoupling (see Figure 7), the Ca\(^{2+}\) inward current then becomes essential for propagation, and application of Ca\(^{2+}\) entry blockers produces localized conduction block.\(^{150}\) Interestingly, the interaction between the macroscopic tissue architecture and excitation, leading to local divergence or convergence of wave fronts and changes in ion channel function, is further modulated by the degree of local gap junctional coupling.\(^{161}\)

**Prospective**

In the near future, most genes responsible for inherited arrhythmogenic conditions will be identified and the genomic structure of disease-related genes will also be defined. This will produce results that are needed for successful management of patients and families. One major result will be the definition of the “molecular epidemiology” of inherited arrhythmogenic conditions. Qualitative statements such as

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**Figure 7.** Involvement of \(I_{\text{Ca,L}}\) and \(I_{\text{Na}}\) in propagation. Middle panel illustrates schematically a column of 4 cells connected by gap junctions. Propagation during normal cell-to-cell coupling is depicted by action potential upstrokes in left row. Note that normal propagation is relatively continuous, ie, major delays between excitation of subsequent cells are absent. Propagation during advanced cell-to-cell uncoupling at a propagation velocity that amounts to \(1/10\) of normal velocity is shown in left column. In this case, propagation is highly discontinuous, ie, membrane sites in a single cell are activated almost simultaneously, and a long conduction delay exists between subsequent cells. Because of this delay, action potential upstroke of driven cell occurs at a time when driver cell is at its early plateau phase (red). Therefore, \(I_{\text{Ca,L}}\) is necessary to support propagation.
“rare” or “common” should be replaced by actual numbers defining the prevalence of each condition in the general population and the relative prevalence of each variant of a disease.

The availability of screening methods with sensitivity and specificity close to 100%, combined with complete clinical information prospectively collected, will define the penetrance of each disease and, within a disease, of each genetic variant. This will lead to guidelines for the management of asymptomatic gene carriers based on the predicted risk of becoming symptomatic.

Given the very large number of mutations associated with arrhythmogenic disorders, it may be more realistic to study carefully the larger group of individuals with defects in the same gene rather than attempting to define genotype/phenotype correlations for each mutation. If distinctive features are identified that segregate patients with specific genetic variants of the same disease, gene-specific therapy may result.

The identification of the genetic or environmental factors that modulate the expression of these diseases and understanding the mechanisms whereby relatives with the identical mutation can have radically different clinical histories will also be useful for patient management.

An intriguing aspect of investigating the molecular bases of inherited arrhythmogenic disorders lies in the hope that information provided by studies of relatively rare inherited conditions may help elucidate mechanisms for the acquired variants. The importance of the discovery of the gene for familial atrial fibrillation, for example, will be enhanced if it will lead to an understanding of the causes of the much more prevalent “ lone” (and even disease-associated) atrial fibrillation. Similarly, recent data suggest that the acquired LQTS may develop in individuals carrying otherwise apparently “mild” mutations in LQTS-related genes. Polymorphisms of these genes could also predispose to other acquired arrhythmias.

Finally, the step from defective gene function to the clinically manifest arrhythmias involves further complexities related to the environment in which the abnormal proteins exert their function. Further study of the multiple interactions among ion channels, pumps, and exchangers on one hand and the structure and connectivity of the cellular network on the other should improve our understanding of the mechanisms that determine the occurrence of the electrical disturbances that lead to lethal arrhythmias.

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Appendix

This article summarizes the outcome of a workshop held at the European Heart House C, Sophia Antipolis, France, October 2–5, 1997. The need for the workshop was proposed by Silvia G. Priori. It was organized by the Study Group on Molecular Basis of Arrhythmias of the Working Group on Arrhythmias of the European Society of Cardiology, and its funding was administered by the Working Group itself. The workshop was cochaired by Silvia G. Priori and André G. Kléber. The final preparation and organization of the manuscript were the responsibility of André G. Kléber, Silvia G. Priori, Dan M. Roden, and Peter J. Schwartz.

References

Note, References 1 through 94 appear in Parts I and II of this article, published in Circulation. 1999;99:518–528 (February 2 issue).


a tentative mechanism for the embryotoxic effect of the class III anti-arrhythmic agent almoklanol. 


143. Le Grice SJ, Small BH, Chau LZ, Edgar SG, Gavin JB, Hunter PJ. Laminar structure of the heart: ventricular myocyte arrangement and connective tissue architecture in the dog. *Am J Physiol.* 1995;338:H571–H582.


145. Priori et al. February 9, 1999


152. Le Grice SJ, Small BH, Chau LZ, Edgar SG, Gavin JB, Hunter PJ. Laminar structure of the heart: ventricular myocyte arrangement and connective tissue architecture in the dog. *Am J Physiol.* 1995;338:H571–H582.


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Part III
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