Background—Hypertrophic cardiomyopathy (HCM) often causes sudden, unexpected death in adolescents and young adults. Alterations in myocardial metabolism are considered to be causes for contractile dysfunction. We examined the question of whether metabolic abnormalities antedate the manifestation of symptoms in patients with HCM.

Methods and Results—Proton-decoupled $^{31}$P NMR spectroscopy of the anterior left ventricular wall of the heart of 14 young, asymptomatic patients with HCM was performed with a 1.5-T whole-body imager. Spectra of the phosphate metabolites were compared with those of normal control subjects. The patients exhibited a significantly reduced ($P<0.02$) ratio of phosphocreatine (PCr) to ATP of 1.98±0.37 (mean±SD), compared with 2.46±0.53 obtained in 11 normal control subjects. In addition, the group of patients with severe hypertrophy of the interventricular septum (n=8) showed a significantly increased ($P<0.05$) P$_i$-to-PCr ratio, with a P$_i$×100/PCr of 20.0±8.3 versus 9.7±7.2 in control subjects. Both abnormalities are similar to those found in ischemic myocardium. This view is also supported by a significantly increased ($P<0.01$) phosphomonoester (PME)-to-PCr ratio, with a PME×100/PCr of 20.7±11.2 compared with 8.4±6.7 in control subjects, indicating altered glucose metabolism.

Conclusions—$^{31}$P NMR spectroscopy detects alterations of myocardial metabolism in asymptomatic patients with HCM. These alterations may contribute to the understanding of the pathophysiology and natural history of the disease. (Circulation. 1998;97:2536-2542.)

Key Words: cardiomyopathy ■ ischemia ■ metabolism

Hypertrophic cardiomyopathy is a primary myocardial disorder characterized by localized hypertrophy of the IVS and the left ventricle that occurs in the absence of aortic stenosis, systemic hypertension, or other obvious causes. Rarely, symmetrical hypertrophy occurs in HCM, involving the right ventricle as well. The myocardium shows zones of disarrayed myocytes and myofibrils with interstitial fibrosis in both hypertrophied and nonhypertrophied regions.

In ≈50% of the patients, a genetically heterogeneous disease is responsible for the defects. Irrespective of the cause, the following abnormalities have been described: abnormal calcium fluxes, abnormal sympathetic stimulation due to increased responsiveness to catecholamines, thickened intramural coronary arteries (occurring in >80% of the patients), abnormal microcirculation leading to increased diastolic stiffness and subendocardial ischemia, and structural abnormalities manifested as cell hypertrophy and disarray. HCM patients, especially adolescents and young adults, have a high risk of sudden death. This risk cannot always be related to the existence of a left ventricular outflow tract obstruction. Myocardial ischemia and altered myocardial metabolism cause contractile dysfunction and ventricular arrhythmias. Research has focused on symptomatic patients with HCM. Myocardial perfusion abnormalities were detected by positron emission tomography or thallium scintigraphy. A more recent study described heterogeneity in regional myocardial glucose uptake and function, but no impairment in blood flow. Because we know little about how myocardial phosphate metabolites are affected in asymptomatic patients with HCM, we performed the following studies using $^{31}$P in vivo NMR spectroscopy. This technique has been used previously to study various heart diseases in humans by determining the myocardial PCr/ATP ratio and, in a few cases, the P$_i$/PCr ratio also. We used these parameters and the PME/PCr ratio as indicators of altered myocardial metabolism. Our results suggest a shift from fatty acid to glucose metabolism, which may be a consequence of metabolic adaptation to hypertrophy and/or chronic ischemia.

Methods

Patients and Control Subjects
We examined 14 young, asymptomatic HCM patients and 11 young, normal control subjects (see Table 1). The protocol was approved by the Ethical Committee of the University of Tübingen. The study complied with the current regulations of the internal review boards and guidelines of the Declaration of Helsinki. All patients gave their written informed consent.

The 14 young asymptomatic patients with HCM were evaluated in the outpatient cardiology clinic. All underwent an electrocardiogram, echocardiography, and a nuclear cardiology study (SPECT). On the basis of echocardiography and SPECT, both the severity of hypertrophy and the presence of ischemic abnormalities were evaluated.

Patients with HCM were selected if they had a left ventricular wall thickness of ≥15 mm, no or atypical obstruction of the left ventricular outflow tract, no obstruction of the aortic valve, a normal left ventricular ejection fraction, and a calculated pulmonary artery pressure ≤40 mm Hg. They were also required to have a normal clinical examination, electrocardiogram, and laboratory findings.

The control group included 11 young normal subjects, in whom echocardiography, electrocardiography, and blood pressure measurement were performed.

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Selected Abbreviations and Acronyms
2,3-DPG = 2,3-diphosphoglycerate
FFA = free fatty acid
HCN = hypertrophic cardiomyopathy
HCN I = group of patients with HCN and MEIST≤250%
HCN II = group of patients with HCN and MEIST>250%
IVS = interventricular septum
MEIST = maximum end-diastolic IVS thickness
NOE = nuclear Overhauser effect
P = phosphocreatine
PDE = phosphodiester
PME = phosphomonoester

TABLE 1. Patients and Controls

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>HCN I</th>
<th>HCN II</th>
<th>HCM all</th>
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<tr>
<td>n</td>
<td>11</td>
<td>6</td>
<td>8</td>
<td>14</td>
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<tr>
<td>Age, y</td>
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<td>16±5</td>
<td>17±9</td>
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<td>0/6</td>
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<td>5/9</td>
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<td>MEIST, mm</td>
<td>10±1</td>
<td>14±4*</td>
<td>33±7†</td>
<td>25±11§</td>
</tr>
<tr>
<td>MEIST, %</td>
<td>111±9</td>
<td>159±49†</td>
<td>387±79‡</td>
<td>289±135§</td>
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<td>5</td>
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<td>LVOTO, n/mm Hg</td>
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<td>2/8.32</td>
<td>3/8.23/32</td>
<td></td>
</tr>
<tr>
<td>Verapamil, n</td>
<td>. .</td>
<td>1</td>
<td>4</td>
<td>5</td>
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</tbody>
</table>

LVOTO indicates left ventricular outflow tract obstruction. Mean values ±SD are given. See text for details.

Unpaired two-tailed Student’s t test resulted in *P<0.02 vs Control; †P<5×10−5 vs HCM II; ‡P<10−7 vs Control; and §P<5×10−4 vs Control.

the institutional Committee for the Protection of Human Subjects, and all subjects gave informed consent.

Five of the patients came to our attention because of their positive family history, 12 because of heart murmurs, and 1 because of signs of left ventricular hypertrophy on the ECG. The diagnosis was confirmed by echocardiography in all patients. Cardiac catheterization was performed in 6. A left ventricular outflow tract obstruction was found in 3 of the 14 patients (see Table 1). All patients were asymptomatic. Holter monitoring for 24 hours revealed no evidence of significant arrhythmias. The blood chemistry was normal in all patients. Five patients were treated with verapamil. Because the characteristic changes of HCM can be found in most cases in the IVS and because in all of our patients the IVS was hypertrophied, the MEIST was used as a criterion for the extent of hypertrophy. MEIST was determined from ECG-gated MRI from images in the long-axis view. The IVS thickness was determined from the end-diastolic image in the thickest region of the IVS. We divided the patients into 2 groups. The group called HCM I (n=6) had a MEIST of <250% and the group called HCM II (n=8) of ≥250% compared with matched healthy control subjects. Mean MEIST values were 159% for the HCM I group and 387% for the HCM II group, resulting in average end-diastolic septum thicknesses of 14 and 33 mm, respectively.

In control subjects, MEIST was determined from the NMR images in the anteroseptal region, leading to a mean value for MEIST of 111% and a mean septum thickness of 10 mm. The control subjects had normal echocardiograms, and none of them had a known medical disorder.

In Vivo NMR Spectroscopy
Examinations were carried out on a Magnetom SP 63 Helicen whole-body imager (Siemens) operating at 1.5 T with 31P and 1H Larmor frequencies of 25.74 and 63.60 MHz, respectively. The whole-body imager was equipped with a second radiofrequency channel for proton decoupling. Transmission and reception were performed with a 100-mm double-resonant single-turn surface coil with the decoupling frequency set to the Larmor frequency of the water protons.

Patients were examined under resting conditions in a prone position, and the surface coil was placed below the anteroseptal region. Flow-rephased gradient-echo proton images were then acquired to control the position. Subsequent ECG-gated nonlocalized shimming was also performed on protons leading to water linewidths between 13 and 25 Hz within ±5 minutes.

A 2-dimensional phosphorous chemical-shift imaging sequence in combination with axial slice-selective excitation, coronal slice-selective chest muscle saturation, and proton decoupling was used to perform a complete 3-dimensional localization as previously described. The pulse angle in the coil center was set to 180°. The application of this sequence with 2048 ECG-gated acquisitions resulted in spectra of 5×2.5×3 cm or 38-mL volume elements within 25 to 35 minutes. Two typical spectra are shown in Figure 1.

In addition to the signals from the myocardium, the spectra also include signals from blood in the left ventricular chamber (see Figure 1): the 2,3-DPG signals and a part of the PDE signals. Furthermore, a small amount of P, and a somewhat larger amount of ATP also originate from blood.

The blood contamination in the spectra is a result of the size and shape of the volume element that was necessary to create a satisfying signal-to-noise ratio, but the blood contributions to ATP and P could be corrected by using its 2,3-DPG signals. In 2 examinations of HCM II patients, the volume element could be completely positioned within the myocardium, leading to undetectable 2,3-DPG signals in the spectra (see Figures 1 and 2).

Data Analysis
We used only 1 of the acquired array of 8×16 volume elements from each chemical-shift imaging sequence measurement. This volume element was positioned such that it covered the anterior part of the insertion of the septum into the anterior wall as well as that part of the anterior left ventricular wall that was located close to it. In addition, the volume element was positioned as close as possible to the surface coil, but special care was taken to exclude surrounding tissue, especially chest muscle.

For the evaluation of the signals from the chosen volume element, the time-domain fitting routine VARPRO (VARiable PROjection) was used with gaussian model functions (see Appendix). The metabolite integral ratios of PCr/ATP and P/Pc were then corrected for blood contamination with a blood 2,3-DPG/ATP ratio of 3 and a blood 2,3-DPG/P, ratio of 15. The correction for saturation
was performed with the average reported longitudinal relaxation times (T1) as given by Bottomley and Ouwerkerk with PCr 4.37 seconds, P_i 4.30 seconds, and g-, α-, and β-ATP 2.52, 2.26, and 2.28 seconds. In addition, the enhancement due to the NOE had to be taken into account. The average enhancements found for our examination protocol were PCr, 61%; P_i, 50%; and g-, α-, and β-ATP, 39%, 34%, and 40%. In addition, pH_i was calculated according to Petroff et al from the chemical shift of P_i (δPi).

Because of the lack of a suitable model function, the PME signal could not be analyzed with the fitting routine. Therefore, in this case we performed additional data processing by subtracting the fitted time-domain signals of PCr, ATP, and 2,3-DPG from the measured signal (see Figure 2). From the resulting spectra, the peak area of PME was determined with a peak integration routine (see Appendix). Average spectra obtained for each group by adding up all individual spectra scaled to equal PCr were analyzed in the same way. These average spectra (see Figure 3) reflect the mean value of each group for PME, P_i, and PDE. The PME integrals are given in Table 2. T1 and NOE corrections for the PME/PCr ratios were not possible because neither T1 nor the NOE enhancement of PME in human heart is known. Correction for blood contamination was not necessary for PME/PCr (see “Results”).

Statistical Analysis

Statistical evaluation was carried out with the unpaired 2-tailed Student’s t test and the Wilcoxon rank order test (U test). Correlations were analyzed by linear regression. Error probabilities of P<0.05 were considered significant. For the significant correlations, an additional check was carried out by discarding the data point with the maximum values. It was required that the remaining points still resulted in a significant correlation. All data are presented as mean±SD, unless otherwise indicated.

Results

PCr/ATP Ratio, P_i/PCr Ratio, and pH_i

Figure 1 shows spectra of a 25-year-old normal man (control, left) and a 23-year-old woman with HCM II (patient, right). The septum thickness was 43 mm, and MEIST was 478% in the patient. The spectrum on the right shows almost no 2,3-DPG signal of blood and clear signals of P_i and PME. In the control spectrum, the P_i signal was smaller, and a possible PME signal was hidden by the pronounced 2,3-DPG signal. Collective data obtained from fit results of the spectra are given in Table 2 with and without correction for blood contamination, saturation, and NOE enhancement. Compared with control subjects, the spectra of HCM II patients showed a significant (100%) increase in the P_i/PCr ratio and a significant (20%) decrease in the PCr/ATP ratio. The group of all HCM patients also revealed a significant (20%) decrease in the PCr/ATP ratio. No significant difference in the PCr/ATP ratio was obtained between HCM I and either control or HCM II. However, the P_i/PCr ratio in HCM I was significantly smaller than in HCM II. The pH_i was identical in all groups.

PME/PCr Ratio

Figure 2 shows sections of the individual spectra of all 8 HCM II patients scaled for equal PCr integral. The left column represents the measured spectra; the middle column the Fourier transform of the time-domain fit results for 2,3-DPG, PCr, and ATP; and the right column the difference between the left and the middle columns. Only the part of the spectrum with positive ppm values was plotted to focus on the PME and P_i signals. The P_i and PME signals were visible in all spectra. The blood contamination decreases from top to bottom and in the 2 bottom spectra was too small for a successful fit of the 2,3-DPG signals. The PME signal observed in all spectra showed no visible dependence on the amount of blood contamination. The 2 bottom spectra showed a decrease from top to bottom and in the 2 bottom spectra was too small for a successful fit of the 2,3-DPG signals. The PME signal observed in all spectra showed no visible dependence on the amount of blood contamination. The 2 bottom spectra showed a decrease from top to bottom and in the 2 bottom spectra was too small for a successful fit of the 2,3-DPG signals. The PME signal observed in all spectra showed no visible dependence on the amount of blood contamination. The 2 bottom spectra showed a decrease from top to bottom and in the 2 bottom spectra was too small for a successful fit of the 2,3-DPG signals.

Figure 3. Average 31P NMR spectra obtained by adding up the spectra within the groups HCM II, HCM I, and Control. 2,3-DPG, PCr, and ATP are eliminated by the subtraction introduced in Figure 2 and the text. In addition to these average spectra, the difference between HCM II and Control is provided in bottom row. These average spectra visualize the mean value within each group. See text for details.
HCM II patients showed significant increases of 150% and 100% compared with control subjects and HCM I patients, respectively (Table 2). Even if the 2 HCM II spectra in which the 2,3-DPG signals were too small for a successful fit were discarded, significant differences were still obtained.

Figure 3 shows sections of the average spectra of the 3 groups HCM II, HCM I, and Control scaled for equal PCr integral and with eliminated 2,3-DPG, PCr, and ATP signals. The PME signal decreased progressively from HCM II to HCM I and to Control. The largest P signal was visible in the HCM II average spectrum, whereas HCM I and Control exhibited similar P signals, which were both smaller. The right signal of the 2 PDE signals observed was predominantly due to blood contamination and was largest in the control group. Thus, although it exhibited the largest amount of blood contamination, the Control group also showed the smallest PME signal, again suggesting that the PME signal must originate from the myocardium. The differences between the HCM II and control groups could be more easily derived from the difference spectrum in the bottom line of Figure 3, which demonstrated the existence of greater PME and P signals and a smaller PDE signal from blood in the HCM II group.

The area under the curve of each average spectrum is given in Table 2. All areas of the average spectra agreed very well with the mean values of the areas of the individual spectra. The PME/PCr ratio was threefold greater in the HCM II group than in the control group.

The difference spectrum of the 2 average spectra HCM II and Control $\Delta$ (HCM II $-$ Control) showed a $\sqrt{2}$ times greater noise level but a superior baseline compared with the 3 group spectra because baseline distortions were subtracted. The flat baseline allowed a quantitative evaluation of the P signal in the difference spectrum and revealed a clearly greater P in HCM II, thus confirming the fit results of the individual spectra.

### Extent of Hypertrophy

Figure 4 shows the correlations of the corrected P/PCr ratio with the corrected PCr/ATP ratio (top left) and with the MEIST (top right) and the correlation between the uncorrected PME/PCr and MEIST (bottom). The correlations with

![Figure 4](https://example.com/figure4.png)

**Figure 4.** Correlations of the P/PCr ratio corrected for blood contamination, saturation, and NOE enhancement with the corrected PCr/ATP ratio (top left) and with the MEIST (top right). In addition, the correlation between the uncorrected PME/PCr ratio and MEIST is given (bottom). See text for details.
MEIST showed that $P/\text{PCr}$ increased and $\text{PCr}/\text{ATP}$ decreased with the extent of hypertrophy.

**Discussion**

The results can be summarized as follows. First, in vivo $^{31}$P NMR spectroscopy yielded well-resolved spectra with a good signal-to-noise ratio from small volume elements. Second, we found a significantly decreased $\text{PCr}/\text{ATP}$ ratio and significantly increased $P/\text{PCr}$ and $\text{PME}/\text{PCr}$ ratios in the myocardium of asymptomatic patients with advanced HCM (HCM II, see Table 2).

Our results on $\text{PCT}/\text{ATP}$ and $P/\text{PCr}$ agree well with findings from de Roos et al.\(^{20}\) who also found increased $P/\text{PCr}$ and decreased $\text{PCT}/\text{ATP}$ in HCM patients. However, the authors’ patient group consisted of both symptomatic and asymptomatic patients. Only 3 of their patients had a markedly low $\text{PCr}/\text{ATP}$ ratio, 2 of whom exhibited symptoms of heart failure.

Our findings provide the first evidence that alterations in the myocardial phosphate metabolism are also present in the myocardium of asymptomatic HCM patients. According to Hochachka et al.\(^{28}\) the 3 most likely causes for such metabolic abnormalities are (1) accelerated work of the heart under resting whole-body conditions, (2) oxygen limitation to cell metabolism severe enough to invoke significant anaerobic contribution to ATP turnover rates, and (3) altered carbon and energy sources fueling the cardiac “engine.” These possible causes will now be discussed in more detail.

1. The large muscle mass itself and the contractile state with increased myocardial shortening and diastolic dysfunction have to be regarded as major determinants of the observed metabolic changes.\(^{30}\) This will be aggravated even further by the hyperactive action that arises from increased sensitivity to catecholamines.\(^{6}\) In addition to these changes of the contractile status, the metabolic effects corresponding to the accelerated hormone drive also have to be considered, especially increased rates of glycogenolysis and lipolysis.\(^{31}\) In this context, it is interesting to note that we found a significant increase in the $\text{PME}/\text{PCr}$ ratio whereby PME is known to contain glycolytic intermediates such as glucose-6-phosphate and $\alpha$-glycerophosphate, and to a lesser degree, AMP.\(^{32}\)

2. Energy demand and supply pathways are tightly coupled, and even large changes in the energy expenditure will not lead to gross alterations of the concentrations of $\text{PCr}$ and $\text{ATP}$.\(^{33}\) In normal human heart, the myocardial $\text{PCr}/\text{ATP}$ ratio does not change with exercise.\(^{34}\) Thus, in addition to an increased energy demand of the HCM heart, a limited supply of oxygen and substrates has to be considered. In fact, the same changes in $\text{PCr}/\text{ATP}$ and $P/\text{PCr}$ can also be induced by a mild reduction in blood flow, as shown by Schaef er et al.\(^{15}\) in animal experiments. Such a putative myocardial energy imbalance in HCM patients may also be due to reduced blood flow caused by the abnormal intramural coronary arteries found in $>80\%$ of the patients that have thickened vessel walls and a reduced lumen size.\(^{12,13}\) These vessels also show abnormal dilatation and, together with the inadequate capillary density and systolic compression of the arteries, the vasodilatory reserve is markedly reduced.\(^{12,13}\) The fact that no changes of myocardial pH, were observed in the present study on patients examined under resting conditions seems to contradict the possible presence of myocardial ischemia. However, Arai et al.\(^{36}\) demonstrated a gradual adaptation of the myocardium to ischemia, showing lactate production returning to lactate consumption within 45 minutes. The occurrence of a limited supply of oxygen and substrates in asymptomatic HCM has in fact been suggested by Camici et al.\(^{12}\) who used pharmacological techniques to estimate the coronary flow reserve. This assumption is supported by O’Gara et al.\(^{11}\) who used thallium scintigraphy and postulated that silent ischemia occurs transiently in asymptomatic HCM patients after workload.

3. A considerable increase of the utilization of glucose in preference to FFAs is known to occur in myocardial hypertrophy.\(^{37}\) When the extent of hypertrophy exceeds a critical heart weight, a decrease in the myocardial $\text{PCr}/\text{ATP}$ ratio can be found that is proportional to the degree of hypertrophy.\(^{38}\) This decrease is thought to be due to subendocardial ischemia.\(^{37}\)

Another plausible explanation is the hypothesis advanced by Holden et al.\(^{39}\) who studied the hearts of Sherpas, who live under hypobaria hypoxia in the Himalaya Mountains. Although their hearts are not hypertrophied, it was suggested that they prefer oxidizing glucose in preference to FFAs. This seems to be indispensable under their circumstances, because the oxidative yield of ATP per mole of oxygen is higher when glucose is oxidized instead of FFAs (P/O ratios, 3.0 for glucose, 2.8 for FFAs\(^ {40}\)). Most interestingly, a reduced myocardial $\text{PCr}/\text{ATP}$ ratio was also reported by the same group.\(^ {29}\) Having no evidence for ischemia or increased workload, the authors explained these changes through accelerated aerobic glycolysis. Provided that the creatine kinase reaction functions close to equilibrium, a decrease in $\text{PCr}/\text{ATP}$ leads to an increase of the ADP concentration, which, in turn, activates the phosphoglycerate kinase and pyruvate kinase enzymes involved in aerobic glycolysis.

Indeed, increased rates of glucose retention were also found in symptomatic patients with HCM by use of $^{18}$F-FDG and positron emission tomography.\(^{13,14}\) Keeping in mind that no changes in myocardial pH were found in the asymptomatic HCM patients, the hypothesis of an increase in glucose oxidation offers a possible explanation for the metabolic alterations observed. The accumulation of glycolytic intermediates represented by the increased $\text{PME}/\text{PCr}$ ratio have to be regarded under these aspects to arise from increased oxidative glucose metabolism as well as from glycogenolysis and lipolysis of triglycerides. This interpretation is strengthened by the correlations of $P/\text{PCr}$ with both $\text{PCr}/\text{ATP}$ and MEIST. The former connects increased $P/\text{PCr}$ ratios with increased ADP concentrations (which are calculated from $P/\text{PCr}$\(^ {41}\)). The latter correlation of $P/\text{PCr}$ with MEIST indicates that increased ADP concentrations occur predominantly in hearts with a greater extent of hypertrophy. These hearts also show higher $\text{PME}/\text{PCr}$ ratios, as demonstrated by the correlation of $\text{PME}/\text{PCr}$ with MEIST (Figure 4, bottom).

These interpretations, however, are correct only if no changes in total creatine occur, because a loss in total creatine could also explain the suggested changes in ADP. The determination of total creatine was not possible in our in vivo
NMR investigations, and this is a limitation of the present method.

The metabolic abnormalities show not only a significant but also an obviously metabolically consistent increase with the extent of hypertrophy. This hypothesis of a true biochemical adaptation to myocardial hypertrophy would support the assumption that the myocardium of patients with advanced HCM (HCM II) suffers from reduced oxygen supply, which is compensated for by an increased amount of glucose oxidation as a mechanism of hypoxia-defense adaptation.

However, the presence of such a mechanism in the HCM heart is questioned by the preliminary results of Zhang et al., who found that changes in the high-energy phosphates in the hypertrophied heart are not a result of impaired oxygen diffusion into the cells but rather reflect alterations in the control of energy metabolism.

Irrespective of their interpretation, the metabolic abnormalities found raise the question of their clinical relevance. Because none of the treatment modalities we have today for HCM, pharmacological or surgical, have thus far been able to correct the basic disorder or to have a favorable effect on the natural history of HCM, Braunwald recently suggested a multicenter clinical trial to evaluate the efficacy of current treatment concepts. However, a great difficulty is that for ethical reasons, symptomatic patients cannot be used as control subjects treated with placebo only. Thus, the findings in asymptomatic patients presented here not only provide tantalizing new insight into the pathophysiology of the disease but also may perhaps help to solve these ethical concerns. We propose that in vivo NMR spectroscopy may be an important tool for the diagnosis and treatment response of asymptomatic patients with HCM.

Appendix

Details on Quantification

After application of the time-domain fitting routine VARPRO (VARiable PROjection)28 using gaussian model functions, standardization of the PCr signal integral was carried out and PCr was set to 0.00 ppm. The main signals of all spectra could easily be fitted with Cramer-Rao SDs, which exceeded 0.07 ppm for the chemical shift interval 6.2 to 7.6 ppm. The region 7.6 to 8.6 ppm was used for quantification of PME in the region 6.2 to 7.6 ppm was possible for all average spectra, the evaluation of P in the region 4.3 to 5.8 ppm was possible only for the difference spectrum between the Control and HCM II average spectra, because otherwise, the computer program was not successful in finding a common baseline.

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2542 NMR Spectroscopy in Hypertrophic Cardiomyopathy


31P NMR Spectroscopy Detects Metabolic Abnormalities in Asymptomatic Patients With Hypertrophic Cardiomyopathy
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