Analysis of Prostacyclin and Thromboxane Biosynthesis in Cardiovascular Disease

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THE APPRECIATION that two oxygenated metabolites of arachidonic acid, prostacyclin (PGI₂) and thromboxane A₂ (TxA₂), have potent and contrasting effects on vascular tone and platelet function in vitro has prompted attempts to define their importance in human vascular disease. Both compounds are formed from arachidonic acid by the enzyme cyclooxygenase via cyclic endoperoxide intermediates, PGG₂ and PGH₂. TxA₂, the major cyclooxygenase product in the platelet, is a vasoconstrictor and potent stimulus to platelet aggregation in vitro. PGI₂, a vasodilator and inhibitor of platelet aggregation, is the predominant product of the same enzyme in vascular endothelial cells. The development of techniques to study platelet function in vivo and ex vivo preceded the discovery of these compounds, and the application of these tests suggests that increased platelet activation may well accompany vascular occlusive events in man. Thus, it seems reasonable that PGI₂ and TxA₂ might play an important role in this and other syndromes of disordered platelet-vascular homeostasis in man.

Support for this concept relies upon the development of methods to measure PGI₂ and TxA₂ in human biological fluids. However, both PGI₂ and TxA₂ have extremely short half-lives and are rapidly cleared from the bloodstream. Furthermore, arachidonic acid release from biological membranes (and thus synthesis of PGI₂ and TxA₂) is likely to be phasic rather than continuous. This phenomenon also renders measurement of these products highly liable to artifact when performed in media obtained by invasive sampling techniques, such as plasma. Finally, there are the technical difficulties involved in devising assays of requisite sensitivity and specificity.

Three approaches have been used: bioassay, radiomimmoassay (RIA) and stable isotope dilution assays involving gas chromatography–mass spectrometry (GC/MS). Bioassays have the advantage of actually measuring biological activity and have been of critical qualitative importance in the discovery of prostaglandins. Because of the evanescence of both PGI₂ and TxA₂ in the circulation, RIA methods are directed toward metabolites. These methods are more sensitive and specific than bioassay and more applicable to quantitative study of prostaglandin turnover in man. Initially, the various approaches appeared to give similar results. PGI₂, unlike other prostaglandins, is not extensively metabolized by the lung, and experiments using bioassay suggested that production by the lung and perhaps other sources permitted PGI₂ to function as a circulating platelet-inhibitory hormone in rabbits and cats. Apparent confirmation of this observation in man was soon provided by transpulmonary measurements of 6-keto-PGF₁α by GC/MS in patients undergoing cardiac catheterization. The inactive hydration product of PGI₂, 6-keto-PGF₁α, is formed nonenzymatically and is more stable than the parent compound. This study suggested that the pulmonary production rate of PGI₂ was in the order of 5 ng/kg/min in man, higher than the threshold dose required to inhibit platelet aggregation in vivo when synthetic PGI₂ was infused into human volunteers.

Radioimmunoassays for TxB₂, the inactive hydration product of TxA₂, were developed and applied to measurements in human plasma, and many reports characterized a supposed "imbalance of the PGI₂/TxA₂ ratio" in a variety of human diseases. However, in the past 2 years, evidence has appeared that the secretion rate of endogenous PGI₂ into the circulation of human volunteers is extremely low (∼ 0.09 ng/kg/min), that 6-keto-PGF₁α in human plasma obtained under resting conditions is in the low picogram range (< 5 pg), and that pulmonary release of PGI₂ is undetectable in healthy volunteers. Thus, although PGI₂ is probably important in local platelet-vascular interactions, it does not function as a circulating hormone under physiologic conditions in man. Similarly, we have found that immunoreactive TxB₂ is undetectable in human plasma obtained from healthy volunteers when care was taken to avoid artifacts induced by sampling and technically acceptable assay methods with limits of sensitivity in the low picogram range were combined with highly specific antibodies.

The importance of these observations is that they invalidate the majority of studies pertaining to endogenous biosynthesis of PGI₂ and TxA₂ in human disease. Studies of PGI₂ and TxA₂ production in the human coronary circulation illustrate this point. Samples are usually obtained after insertion of catheters, a procedure that is likely to traumatically stimulate vascular PGI₂ synthesis. Other aspects of the procedure, such as the use of heparin and radiocontrast media, may also alter synthesis of either compound. Platelet activation at the catheter tip and ex vivo prostanooid formation represent other sources of potential artifact. Finally, it may not be apparent that levels of these compounds are artifactualy elevated, because the data are often presented as 6-keto-PGF₁α/TxB₂ ratios or coronary sinus/aortic blood ratios of either compound.

It has been proposed that substantial quantitative differences between bioassay estimates of prostacyclin-like activity (2–6 ng/ml) and measurement of 6-keto-PGF₁α in plasma by negative ion chemical ioniza-
tion GC/MS (0.5–2 pg/ml) may result from predominant conversion of PGI to 6-keto-PGE, a biologically active compound that is a putative metabolite of PGI in vivo. However, this seems highly unlikely because studies of the metabolite disposition of PGI in man have failed to detect metabolites with the 6-keto-PGE structure and because infusion of synthetic PGI in volunteers does not result in accumulation of 6-keto-PGE in plasma.

One method of circumventing problems of sampling-induced artifact and ex vivo prostaglandin formation is by measurement of metabolites that have an extended half-life. Although the 13,14-dihydro-6,15-diketo-PGF, metabolite of PGI, has a longer half-life than 6-keto-PGF, and has been identified in human plasma after PGI infusions, experience with other prostaglandins has shown that the metabolites initially formed from the hydration products are rapidly replaced by more polar products as the major compounds in blood. Measurement of tetranor metabolites has been recently utilized to detect phasic release of PGF, in patients with primary dysmenorrhea who suffer episodic pain. Quantitative assays for stable oxidative metabolites of PGI and TXA2 in plasma are currently being developed.

An alternative approach has been the measurement of prostaglandin breakdown products in urine. When this is performed by GC/MS, it represents a highly specific, noninvasive index of endogenous prostaglandin biosynthesis. Measurement of urinary metabolites by RIA is even more liable to problems of sensitivity and specificity than in plasma. Performance of RIAs on unextracted urine and the use of antibodies with high cross-reactivity to structurally related compounds have been common sources of error. However, the combination of rigorous sample purification with highly specific antibodies has permitted the measurement of 6-keto-PGF in urine by RIA. Although 6-keto-PGF and TXB are urinary metabolites of infused PGI and TXB, in man, these indexes are likely to predominantly reflect renal PGI and TXA2 generation under physiologic conditions. Should extrarenal production of either compound be increased (e.g., during a vascular catastrophe), one would anticipate an increase in the urinary levels of the relevant hydration product. For example, urinary 6-keto-PGF excretion is increased in preterm infants with persistent ductus arteriosus and respiratory distress syndrome in whom an increase in systemic prostaglandin biosynthesis is likely. The dinor metabolites of 6-keto-PGF and TXB, are the most abundant products of systemically administered, radiolabeled PGI and TXB, respectively, in man (table 1). Thus, urinary levels of 2,3-dinor-6-keto-PGF (PGI-M) and 2,3-dinor-TXB (TX-M) have been used to quantify the effects of aspirin and similar drugs upon systemic PGI2 and TXA2 synthesis in man and to accurately predict plasma levels of endogenous PGI in the low picogram range.

Although these measurements represent the only noninvasive method of quantifying endogenous generation of PGI and TXA2, they are not specific to the tissue or origin of these compounds. More specific indexes have been studied ex vivo, such as TXB formation in serum or in stimulated platelet-rich plasma or 6-keto-PGF formation by biopsies of vascular

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<th>Prostacyclin:</th>
<th>Biological fluid</th>
<th>Assay method</th>
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<tr>
<td>1. 6-keto-PGF&lt;sub&gt;1α&lt;/sub&gt;</td>
<td>Plasma</td>
<td>GC/MS and RIA</td>
<td>Levels ≤ 2 pg/ml under physiologic conditions.</td>
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<tr>
<td>2. 6,15-diketo-13,14-dihydro-PGF&lt;sub&gt;1α&lt;/sub&gt;</td>
<td>Plasma</td>
<td>RIA</td>
<td>Longer half-life than 6-keto-PGF&lt;sub&gt;1α&lt;/sub&gt; in plasma.</td>
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<tr>
<td>3. 6-keto-PGE&lt;sub&gt;1&lt;/sub&gt;</td>
<td>Plasma</td>
<td>GC/MS</td>
<td>Maximal estimates in the low picogram range. No conversion to this compound from infused PGI&lt;sub&gt;2&lt;/sub&gt;.</td>
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<tr>
<td>4. 6-keto-PGF&lt;sub&gt;1α&lt;/sub&gt;</td>
<td>Urine</td>
<td>GC/MS and RIA</td>
<td>Many RIAs unreliable due to problems of urine purification and/or antibody specificity. May largely reflect renal production under physiologic conditions.</td>
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<tr>
<td>5. 2,3-dinor-6-keto-PGF&lt;sub&gt;1α&lt;/sub&gt;</td>
<td>Urine</td>
<td>GC/MS</td>
<td>Reflects mainly systemic (extrarenal) PGI2 biosynthesis.</td>
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<tr>
<td>6. 6,15-diketo-13,14-dihydro-2,3-dinor-PGF&lt;sub&gt;1α&lt;/sub&gt;</td>
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<th>Thromboxane A&lt;sub&gt;2&lt;/sub&gt;:</th>
<th>Biological fluid</th>
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<tr>
<td>1. Thromboxane B&lt;sub&gt;2&lt;/sub&gt;</td>
<td>Plasma</td>
<td>RIA</td>
<td>Levels &lt; 15 pg/ml under physiologic conditions.</td>
</tr>
<tr>
<td>2. Thromboxane B&lt;sub&gt;2&lt;/sub&gt;</td>
<td>Urine</td>
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<td>3. 2,3-dinor-thromboxane B&lt;sub&gt;2&lt;/sub&gt;</td>
<td>Urine</td>
<td>GC/MS</td>
<td>Reflects mainly systemic (extrarenal) thromboxane synthesis.</td>
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Abbreviations: GC/MS = gas chromatography–mass spectrometry; RIA = radioimmunoassay.
tissue. However, these measurements are related to capacity rather than to actual production rates in vivo. For example, although biopsies of atherosclerotic vessels have a lower capacity to release 6-keto-PGF1α than do those of healthy vessels ex vivo, endogenous PGI2 biosynthesis is actually increased in patients with diffuse atherosclerosis and platelet activation, perhaps resulting from traumatic platelet-vascular interactions in vivo.

In conclusion, measurement of urinary metabolites represents the only noninvasive approach to quantitation of endogenous PGI2 and TxA2 biosynthesis. Methodologic difficulties and sampling artifacts have cast serious doubt on much of the published data, which are dependent on plasma measurements of TxB2 and 6-keto-PGF1α. The relationship of tissue-specific, capacity-related indexes to endogenous production rates of PGI2 and TxA2 under conditions of stimulation and depression is largely unknown. Clarification of the importance of PGI2 and TxA2 in human cardiovascular disease awaits the more widespread application of reliable analytical techniques in this area.

References


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