Is There a Need to Add Another Dimension (Time) to the Evaluation of the Arrhythmogenic Potential of New Drug Candidates In Vitro?

Claire Townsend, PhD

Most drug therapy regimens expose the human body to a foreign chemical for several hours to days and even years. Hence, before a new drug is approved by regulatory agencies, extensive safety studies are conducted to ensure that exposure to the drug will not cause undesirable effects in patients. A major cause of adverse events and drug attrition is cardiovascular toxicity. Drug developers have attempted to identify these issues earlier in medicine development to reduce risks to human volunteers in clinical trials and costs of pursuing the development of unsafe drugs. In 2005, the International Conference on Harmonization of Technical Requirements for Registration of Pharmaceuticals for Human Use issued a guideline for the examination of new drug candidates in a series of in vitro and in vivo tests to assess their arrhythmogenic potential. Inhibition of the delayed rectifier potassium current (\(I_{Kr}\)) in the heart has been linked to the majority of drug-induced arrhythmias. As a result, in vitro safety testing has been focused on acute drug effects on \(I_{Kr}\) or hERG, the potassium channel that underlies \(I_{Kr}\). The development of high-throughput automated methods to measure hERG currents in heterologous expression systems has fueled the emphasis of in vitro testing on \(I_{Kr}\) early in drug discovery and perhaps prevented the development of new medicines by discarding compounds prematurely. This testing paradigm has been challenged over the past several years with the realization that verapamil and a number of other drugs that inhibit \(I_{Kr}\) at therapeutic concentrations do not cause arrhythmias in patients. Verapamil has compensatory effects on other cardiac ion channels such that action potential duration is not affected by this drug and it is safe to administer to patients. In addition, arsenic trioxide, pentamidine, and other drugs associated with a prolonged QT interval and cardiac arrhythmias do not block \(I_{Kr}\) acutely but instead inhibit its trafficking to the cell surface when applied for prolonged (overnight or longer) periods of time. More recently, Lu and colleagues showed that prolonged dofetilide exposure increased late sodium currents and caused early afterdepolarizations in canine ventricular myocytes. Intracellular infusion of phosphatidylinositol 3, 4, 5-trisphosphate (PIP3) reversed these effects. Lu and colleagues were also able to demonstrate a link between prolonged action potential duration and increases in late sodium currents, \(I_{NaL}\), in those cells. Most surprising perhaps was their finding that terfenadine, a well-known antihistamine associated with \(I_{Kr}\) block and cardiac arrhythmias, exerted the same effects on \(I_{NaL}\), in a PIP3-sensitive fashion. The report in this issue of Circulation by Yang and colleagues that several proarrhythmic drugs thought to be “selective” \(I_{Kr}\) inhibitors could also increase late sodium currents after prolonged (≥5 hours) exposure further highlights the limitations of examining acute \(I_{Kr}\) effects as a sole in vitro predictor of arrhythmias.

**Article see p 224**

Yang and colleagues describe the effects of acute versus “chronic” (≥5 hours) exposure to dofetilide on action potentials in mouse cardiomyocytes and human induced pluripotent stem cell–derived cardiomyocytes. In both mouse and human cells, prolonged exposure to dofetilide increased action potential duration and caused both early and delayed afterdepolarizations. Although these effects could be attributed to \(I_{Kr}\) block in human cells, adult mouse cells do not express \(I_{Kr}\). In the latter, dofetilide must therefore be affecting other important determinant(s) of cardiac excitability. The authors then showed that prolonged dofetilide exposure increased late sodium currents in mouse cardiomyocytes, human induced pluripotent stem cell–derived cardiomyocytes, and Chinese hamster ovary cells transfected with the cardiac sodium channel Nav1.5. Peak sodium currents were also increased, but sodium channel protein levels were unchanged. These findings prompted Yang and colleagues to examine the gating properties of sodium channels after a 5-hour exposure to dofetilide. Channel inactivation was shifted toward more positive potentials by ≈20 mV, yielding window currents between −60 and −40 mV. Channel recovery from inactivation was faster, and the rate constants for fast and slow inactivation of macroscopic currents were increased. Taken together, these changes in channel gating are consistent with increased peak and late sodium currents. The mechanism by which the P13K pathway regulates sodium channel gating remains to be determined. However, the results from Yang and colleagues show that it is conserved in mouse cardiomyocytes, human stem cell–derived cardiomyocytes, and hamster ovary cells. The protein kinase Akt, a downstream effector of P13K, may be involved; the authors observed reduced Akt phosphorylation after prolonged exposure to dofetilide. In addition, the P13K inhibitor LY294002 decreased Akt phosphorylation and increased late sodium currents in cells expressing Nav1.5.

The opinions expressed in this article are not necessarily those of the editors or of the American Heart Association. From GlaxoSmithKline, Research Triangle Park, NC. Correspondence to Claire Townsend, PhD, GlaxoSmithKline, 5 Moore Dr, Research Triangle Park, NC 27709-3398. E-mail claire.y.townsend@gsk.com (Circulation. 2014;130:219-220.) © 2014 American Heart Association, Inc. Circulation is available at http://circ.ahajournals.org DOI: 10.1161/CIRCULATIONAHA.114.010819

219
Yang and colleagues also found that effects on late sodium currents were not limited to dofetilide or terfenadine. They reported that other $I_{kr}$ blockers also increased $I_{Na,L}$ in Chinese hamster ovary cells transfected with Nav1.5. D-sotalol and E-4031, both methane sulfonamides like dofetilide, increased $I_{Na,L}$ after prolonged exposure. They also observed increased late sodium currents, although to a lower extent, with 3 unrelated drugs, the antipsychotics haloperidol and thioridazine and the antibiotic erythromycin. Finally, moxifloxacin and verapamil did not alter the amplitude of $I_{Na,L}$. This range of effects on late sodium currents could significantly contribute to the variety of proarrhythmic activities of these $I_{kr}$ blockers.

Increased late sodium currents are a known cause of cardiac arrhythmias. Several mutations in SCN5A, the gene encoding the cardiac sodium channel Nav1.5, underlie the long-QT syndrome type 3. These mutations lead to various levels of late sodium currents, all very small relative to peak current amplitudes (≤3%) but large enough to disrupt action potentials and to cause arrhythmias.

This article may show us the tip of the iceberg with regard to the effects of prolonged drug exposure on cardiac action potentials and their underlying ionic currents. Additional studies are warranted to determine the prevalence of drug effects on late sodium currents. Most early safety pharmacology studies on cardiac ion channels are performed on automated electrophysiology instruments. These instruments, however, generally do not have the sensitivity to measure late sodium currents, all very small relative to peak current amplitudes (≤3%) but large enough to disrupt action potentials and to cause arrhythmias.

The need to examine the effects of drug candidates on multiple cardiac ion channels and to review the current testing paradigm is the focus of a recently initiated public-private project called Comprehensive In Vitro Proarrhythmia Assay. This project proposes, in part, the examination of drug effects on various cardiac channels and their integration in silico by computer models of human cardiac electrophysiology, with the aim of proposing a new paradigm for the assessment of the proarrhythmic potential of new drugs. However, most cardiac ion channel assays and modeling efforts examine short-term exposure to drugs and do not consider prolonged or long-term effects. For example, they do not incorporate effects on channel trafficking, which have been well documented for several drugs and the hERG channel. However, recent studies point out the need to include binding kinetics to refine in silico models. The findings of Yang and colleagues are therefore timely and should stimulate discussions in the field of cardiac safety on the necessity to look at prolonged effects of drug candidates on the various determinants of cardiac excitability.

**Disclosures**

None.

**References**


**Key Words:** Editorials • arrhythmias, cardiac • phosphatidylinositol 3-kinase • safety • sodium channels
Is There a Need to Add Another Dimension (Time) to the Evaluation of the Arrhythmogenic Potential of New Drug Candidates In Vitro?

Claire Townsend

*Circulation* 2014;130:219-220; originally published online June 3, 2014; doi: 10.1161/CIRCULATIONAHA.114.010819

*Circulation* is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2014 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circ.ahajournals.org/content/130/3/219

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in *Circulation* can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to *Circulation* is online at:
http://circ.ahajournals.org//subscriptions/