Role for DNA Damage Signaling in Pulmonary Arterial Hypertension

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Background—Pulmonary arterial hypertension (PAH) is associated with sustained inflammation known to promote DNA damage. Despite these unfavorable environmental conditions, PAH pulmonary arterial smooth muscle cells (PASMCs) exhibit, in contrast to healthy PASMCs, a pro-proliferative and anti-apoptotic phenotype, sustained in time by the activation of miR-204, nuclear factor of activated T cells, and hypoxia-inducible factor 1-α. We hypothesized that PAH-PASMCs have increased the activation of poly(ADP-ribose) polymerase-1 (PARP-1), a critical enzyme implicated in DNA repair, allowing proliferation despite the presence of DNA-damaging insults, eventually leading to PAH.

Methods and Results—Human PAH distal pulmonary arteries and cultured PAH-PASMCs exhibit increased DNA damage markers (53BP1 and γ-H2AX) and an overexpression of PARP-1 (immunoblot and activity assay), in comparison with healthy tissues/cells. Healthy PASMCs treated with a clinically relevant dose of tumor necrosis factor-α harbored a similar phenotype, suggesting that inflammation induces DNA damage and PARP-1 activation in PAH. We also showed that PARP-1 activation accounts for miR-204 downregulation (quantitative reverse transcription polymerase chain reaction) and the subsequent activation of the transcription factors nuclear factor of activated T cells and hypoxia-inducible factor 1-α in PAH-PASMCs, previously shown to be critical for PAH in several models. These effects resulted in PASMC proliferation (Ki67, proliferating cell nuclear antigen, and WST1 assays) and resistance to apoptosis (terminal deoxynucleotidyl transferase dUTP nick end labeling and Annexin V assays). In vivo, the clinically available PARP inhibitor ABT-888 reversed PAH in 2 experimental rat models (Sugen/hypoxia and monocrotaline).

Conclusions—These results show for the first time that the DNA damage/PARP-1 signaling pathway is important for PAH development and provide a new therapeutic target for this deadly disease with high translational potential. (Circulation. 2014;129:786-797.)

Key Words: DNA damage ■ microRNAs ■ pulmonary arterial hypertension ■ PARP1 protein, human

Pulmonary arterial hypertension (PAH) is a disease with no known cure,1 characterized by obstructive vascular lesions of the distal pulmonary arteries (PAs) due in part to excessive proliferation2,3 and resistance to apoptosis4,5 of the pulmonary artery smooth muscle cells (PASMCs). This phenotype is often associated with a chronic inflammatory response characterized by sustained activation of circulating proinflammatory molecules such as tumor necrosis factor-α (TNF-α) and interleukin 6 (IL-6).6 Although an environment of strong inflammation is known to be toxic for the cells and is known to induce DNA damage,7,8 PAH-PASMCs apparently adapt to these unfavorable environmental conditions and are able to survive and proliferate. Indeed, PAH-PASMCs share several features with cancer cells (which also proliferate under unfavorable conditions of accumulated genetic damage or hypoxia), including the expression of prosurvival proteins such as Pim-110 and survivin11 and a metabolic switch toward glycolysis even under normoxia (known as the Warburg effect).12 We have recently shown that all of these changes can be sustained in time by the downregulation of the microRNA miR-20413 and the subsequent activation of the transcription factors nuclear factor of activated T cells (NFAT) and hypoxia-inducible factor 1-α (HIF-1α).14

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 Nonetheless, a question remains unanswered: How can PAH-PASMCs adapt to the DNA damage caused by sustained inflammation? We hypothesized that PAH is associated with a sustained activation of the DNA repair machinery that, in addition to DNA repair, may also directly promote PASMC proliferation.

Poly(ADP-ribose) polymerase-1 (PARP-1) is the best characterized and most abundant member of the PARP family, which currently comprises 6 members.13 On activation, by binding to DNA strand breaks, PARP-1 uses nicotinamide adenine dinucleotide to generate large amounts of poly(ADP-ribose) along the break site and therewith contributes actively to DNA repair.16–18 PARP-1 can trigger DNA repair and thus allow or promote cell survival (when limited stress leads to a light to moderate PARP-1 activation) or promote cell death when the stress and the subsequent PARP-1 activation and hence its nicotinamide adenine dinucleotide consumption are excessive. Thus, PARP-1 can function as a cellular stress sensor.19 In addition to DNA repair, PARP-1 may be implicated in many other features seen in PAH. For example, PARP-1 is implicated in the transcriptional regulation of a number of genes involved in PAH including IL-6.20 Cells with activated PARP-1 produce high levels of IL-6, which can amplify inflammation and, by activating STAT3,21 can contribute to NFAT and HIF-1α activation that promotes the apoptosis resistance that characterizes PAH-PASMCs. Using lung tissues, distal PAs, and primary cultures of PASMCs, we demonstrate for the first time that PAH is associated with sustained DNA damage leading to PARP-1 activation. PARP-1 activation promotes cell survival and proliferation by triggering a miR-204–dependent activation of NFATc2 and HIF-1α. Most importantly, we show that the clinically available PARP inhibitor ABT-888 reverses the PAH in vivo in 2 PAH rat models.

**Methods**

All experiments were performed with the approval of Laval University and the IUCPO Biosafety and Ethics Committees. The online-only Data Supplement provides more details on all methods, including the supplies and chemicals used.

**Human Tissue Samples**

Tissues were obtained from patients who had previously given signed consent. PAH and control small PAs (<1000 μm diameter) were freshly isolated from explanted lungs. Non-PAH lung tissues (controls) were obtained during lung resection for tumors from the noncancerous segments. PAH and control tissues (lungs, brain, kidneys) were obtained from Respiratory Health Network tissue bank (Table).

**Cell Culture**

PAH-PASMCs were isolated from <1000-μm-diameter small PAs from 4 PAH patients, all confirmed by right catheterization (3 idiopathic PAH and 1 associated PAH [scleroderma]). PASMC phenotype was confirmed by using α-smooth muscle actin staining. Control PASMCs (n=5 cell lines) were purchased from Cell Application (San Diego, CA).

**PARP Activity Assay**

PARP activity was assayed by using the Trevigen Universal chemiluminescent PARP assay kit with modifications.

**Table. Clinical Characteristics of PAH Patients and Controls**

<table>
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<th>Control (n=18)</th>
<th>IPAH (n=7)</th>
<th>HPAH (n=1)</th>
<th>APAH* (n=9)</th>
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<td>86</td>
<td>100</td>
<td>78</td>
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<td>45±10</td>
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<tr>
<td>III</td>
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<td>1 (100)</td>
<td>4 (44)</td>
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<td>IV</td>
<td>3 (43)</td>
<td>0</td>
<td>5 (56)</td>
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<td>2.2±0.6</td>
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<td>403</td>
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<td>0</td>
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</tbody>
</table>

Values are mean±standard error of the mean. 6MWD indicates 6-minute walk distance; APAH, associated pulmonary arterial hypertension; CI, cardiac index; CO, cardiac output; HPAH, heritable pulmonary arterial hypertension; IPAH, idiopathic pulmonary arterial hypertension; mPAP, mean pulmonary arterial pressure; PAH, pulmonary arterial hypertension; PDE5, phosphodiesterase-5; RAP, right atrial pressure; PVR, pulmonary vascular resistance; and SvO₂, venous oxygen saturation. *APAH includes patients with connective tissue disease and pulmonary veno-occlusive disease–associated PAH. Please note than some patients take >1 type of medication.

**Proliferation and Apoptosis Measurements**

PASMC proliferation was measured by using Ki67, proliferating cell nuclear antigen, and WST1 assay, and terminal deoxynucleotidyl transferase dUTP nick end labeling and Annexin V assessed apoptosis.

**DNA Damage**

Immunofluorescence for γ-H2AX and immunohistochemistry for 53BP1 were performed to assess DNA damage.

**Animal Models**

The Sugen-hypoxia (Sugen) and the monocrotaline (MCT) PAH rat models were used in the present study. Once PAH established (after 14 days for MCT and week 5 for Sugen), the PARP inhibitor (ABT-888, 6 mg/kg, Enzo Life Sciences) or vehicle (drinking water) was administered per os every day for 2 weeks.

**Statistical Analysis**

Values are expressed as fold change or mean±standard error of the mean, as they follow a normal distribution. Unpaired Student t tests were used for comparisons between 2 groups, and 1-way analysis of variance followed by a Tukey-Kramer post test was used for >2 groups. When same cells were used under different conditions, paired analyses were performed among treatments. Furthermore, if variance depended on the mean of the data, statistical analyses were performed on log transformation of the data. Probability
values <.001 (***) , .01 (**), and .05 (*) were considered as statistically significant.

Results
Increased DNA Damage and PARP-1 Expression in Human PAH

We measured DNA damage in human distal PAs from 6 PAH and 6 control patients (5 distal PAs/patient), using the expression of the p53-binding protein 1 (53BP1) measured by immunohistochemistry, as marker of DNA damage.22 As shown in Figure 1A, DNA damage is significantly increased in PAH in comparison with normal tissue controls. Cells with positive nuclear staining were considered positive and this was divided by the total amount of cells (total nuclei) to calculate the percentage of positive cells. Immunohistochemistry negative control is provided in Figure IA in the online-only Data Supplement. We then measured PARP-1 protein expression in human distal PAs from 5 control and 5 PAH patients by immunoblot (Figure 1B) and found that PARP-1 expression is significantly increased in PAH in comparison with control distal PAs. Thus, we studied whether these changes were also present in primary cultured human PAH-PASMCs. PAH-PASMCs were freshly isolated from small PAs of 4 PAH patients, and 5 control PASMC cell lines were purchased. DNA damage and PARP-1 expression and activation were measured. Using γ-H2AX as DNA damage marker, we showed that, as in distal PAs, DNA damage is sustained in cultured PAH-PASMCs in comparison with healthy PASMCs. The number of γ-H2AX foci in PAH-PASMCs are further increased by the addition of the PARP-1 inhibitor ABT-888 or AG14361 and with PARP-small interfering RNA as well (in comparison with scramble siRNA treated cells; Figure 1C), suggesting that PARP-1 activation is important for the repair of these damages, but not enough to eliminate them completely, because a certain level of DNA damage persists in PAH-PASMCs. This suggests that PARP-1 activation in PAH is mild, far from the level of activation that can be reached after γ-radiation for example.19 By Western blot, we then demonstrated that PAH-PASMCs have increased levels of PARP-1, and the

![Figure 1. DNA damage, PARP-1 expression, and activation are increased in PAH distal PAs and PAH-PASMCs. A, DNA damage (53BP1) was quantified in distal PAs in 6 control vs 6 PAH patients (5 PAs/lung). B, PARP-1 expression was quantified in freshly isolated distal PAs from 5 control and 5 PAH patients. PARP-1 is increased in PAH distal PAs. C, PAH-PASMCs are also associated with increased DNA damage measured by γH2AX nuclear staining. As shown, PAH-PASMCs have increased DNA damage, which is significantly increased by PARP-1 inhibition (by ABT-888, AG14361, or siPARP-1; n=4). D, Consequently, PAH-PASMCs have increased PARP-1 expression, and this expression is further increased with ABT-888 treatment as demonstrated by Western blot (n=4). E, Although PARP-1 expression is increased after ABT-888 treatment, we confirmed that it does block PARP-1 activity (measured by chemiluminescence assay; n=3). *P<0.05; **P<0.01; ***P<0.001. DAPI indicates 4′,6-diamidino-2-phenylindole; PAH, pulmonary arterial hypertension; PARP-1, poly(ADP-ribose) polymerase-1; PASMC, pulmonary arterial smooth muscle cells; and si, small interfering.
further DNA damage increased (by blocking the repair by ABT-888), the further PARP-1 expression is increased (Figure 1D). The same effect is also seen with AG14361 treatments (Figure IB in the online-only Data Supplement). Moreover, as expected, although PARP-1 small interfering RNA increases DNA damage, it did not increase PARP-1 expression demonstrating small interfering RNA efficiency (Figure IB in the online-only Data Supplement). Note that, although ABT-888 increases PARP-1 expression, it inhibits its activity23 assayed by using the chemiluminescent PARP assay kit (Figure 1E). We also confirmed that AG14361 blocks PARP-1 activity by using the same assay (Figure 1E).

To evaluate DNA repair, we measured fold changes in DNA damage after DNA-damaging agent etoposide24 treatment and a 2-hour recovery. Fold changes were analyzed because PAH-PASMCs have higher basal levels of DNA damage (Figure 1C). In brief, cells were treated with 1 μmol/L (micro M) etoposide for 1 hour and after withdrawal of etoposide, cells were incubated in fresh medium for 2 hours to allow DNA repair, as previously described.25 The remaining DNA damage was then assessed (γ-H2AX). For ABT-888 treatments, cells were incubated with 10 μmol/L (micro M) ABT-888 for 48 hours before etoposide treatment, and ABT-888 was kept in medium for the recovery period.

As shown in Figure 2A, control PASMCs treated with etoposide have increased DNA damage (3.5-fold increase). On the other hand, in PAH-PASMCs, etoposide does not significantly increase DNA damage, suggesting that increased PARP-1 expression in these cells provides a more efficient DNA repair in the 2-hour recovery period. Indeed, if PARP activity is inhibited by ABT-888, etoposide treatment then shows increased DNA damage (2-fold increased in comparison with nontreated PAH-PASMCs; Figure 2A).

To determine whether the increased expression and activation of PARP-1 is restricted to the lungs, we analyzed other tissues from PAH patients and demonstrated that PARP-1 was not overexpressed (brain, kidney; Figure IIA in the online-only Data Supplement). We also confirmed that PARP-1 expression is increased in both MCT- and Sugen-induced PAH lungs (Figure IIB in the online-only Data Supplement). The efficiency of treatment in vivo was measured, as shown in Figure IIB in the online-only Data Supplement. As in humans, PARP-1 is upregulated within distal PAs, but not other tissues such as left ventricle and liver (Figure IIC in the online-only Data Supplement).

The mechanism accounting for increased DNA damage and PARP-1 protein expression is likely to be multifactorial, but the presence of sustained inflammatory signaling in PAH may

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**Figure 2.** Inflammation triggers DNA damage and PARP-1 expression and activation. **A**, We measured fold changes in DNA damage after DNA-damaging agent etoposide treatment (1 μmol/L (micro M) for 1 hour) and a 2-hour recovery, allowing DNA repair. The remaining DNA damage was then assessed (γ-H2AX; n=3). **B**, Control cells (n=4) stimulated with TNF-α (100 ng/mL for 48 hours) have increased DNA damage as shown by γ-H2AX nuclear staining. This was further increased with ABT-888 treatment (n=3). **C**, This increase in DNA damage correlated with increased PARP-1 expression (measured by Western blot; n=3). *P*<0.05; **P**<0.01. DAPI indicates 4′,6-diamidino-2-phenylindole; PAH, pulmonary arterial hypertension; PARP-1, poly(ADP-ribose) polymerase-1; PASMC, pulmonary arterial smooth muscle cells; and TNF-α, tumor necrosis factor α.
play a major role. Hence, we measured whether TNF-α, an inflammatory cytokine increased and implicated in PAH pathogenesis, can trigger DNA damage and subsequent PARP-1 activation in PASMCs. As shown in Figure 2B, control PASMCs treated with TNF-α had an increased number of DNA damage foci (γ-H2AX) in comparison with vehicle-treated PASMCs. Similar results were also found when PASMCs were treated with IL-6 or platelet-derived growth factor (Figure IC in the online-only Data Supplement). The increase in DNA damage was associated with a significant upregulation of PARP-1 protein expression measured by immunoblot and immunofluorescence (Figure 2C). As expected, PASMCs treated with both TNF-α and ABT-888 showed more DNA damage leading to a greater PARP-1 expression (Figure 2C).

PARP Inhibition Decreases Proliferation and Promotes Apoptosis in PAH-PASMCs

We next studied the implication of PARP-1 in PAH-PASMC survival and proliferation. We showed that PAH-PASMCs have significantly greater proliferation rate (proliferating cell nuclear antigen, Ki67, and WST1 assays) and a significant apoptosis-resistant phenotype (as indicated by Annexin V and terminal deoxynucleotidyl transferase dUTP nick end labeling assays), in comparison with control-PASMCs placed in the same conditions (Figure 3 and Figure III in the online-only Data Supplement). PARP-1 inhibition by ABT-888 in PAH-PASMCs restores a normal phenotype, thus decreasing proliferation and restoring apoptosis to levels similar to those found in control cells. We further confirmed PARP-1 implication by the use of PARP-1 small interfering RNA, showing similar findings to ABT-888 (Figure 3).

PARP-1 Inhibition Reverses miR-204–Dependent Upregulation of NFAT and HIF-1α in PAH-PASMCs

We have previously described a critical role of the microRNA miR-204 in the etiology of PAH. We had demonstrated that,
through a STAT3-dependent mechanism, the downregulation of miR-204 accounts for the upregulation of the transcription factors NFAT and HIF-1α in PAH-PASMCs, which is contributing to their proliferation and resistance to apoptosis.5,27 Cells with activated PARP-1 produce high levels of IL-6, 28 which, by activating STAT3,21 contributes to miR-204 downregulation. We previously published that PAH-PASMCs produce higher levels of IL-6 and have activated STAT3.14 Thus, by regulating IL-6 production, PARP-1 will downregulate miR-204.14 To determine whether PARP-1 can affect miR-204 expression, we performed quantitative reverse transcription polymerase chain reaction in PAH-PASMCs in the presence and absence of PARP-1 inhibitor ABT-888. We found that miR-204 downregulation in PAH-PASMCs is reversed by ABT-888 (Figure 4A). As predicted, PARP-1–dependent downregulation of miR-204 triggers NFAT and HIF-1α activation measured by nuclear translocation (Figure 4B and 4C).

In PAH-PASMCs, NFAT- and HIF-1α–mediated proliferation has been linked to the downregulation of K+ channels,4,29 resulting in membrane depolarization,29,30 opening of the voltage-dependent calcium channels, thereby increasing intracellular calcium concentration ([Ca2+]i).5,30,31 Using Fluo-3 AM, we measured the effect of PARP-1 inhibition on [Ca2+]i. The inhibition of PARP-1 (ABT-888) in PAH-PASMCs decreased [Ca2+]i to the levels seen in control-PASMCs (Figure 4D). Furthermore, resistance to apoptosis observed in PAH-PASMCs has been linked to mitochondrial membrane potential (ΔΨm) hyperpolarization, blocking the release of proapoptotic mediators such as cytochrome c.5,27,32 Using tetramethylrhodamine methyl ester, we observed that ABT-888 treatment in PAH-PASMCs decreases ΔΨm to similar levels of control-PASMCs (Figure 4D). To further investigate the implication of PARP-1, miR-204, NFAT, and HIF-1α, we performed cotreatments of ABT-888 with miR-204 mimic, VIVIT (NFAT inhibitor), or small interfering HIF-1α and measured the effect on calcium concentration (FLUO-3 AM) and mitochondrial membrane potential (tetramethylrhodamine methyl ester). Note that the VEET peptide was used as control for VIVIT and had no effects (not shown) as previously published by our group.7 As previously demonstrated,
NFAT or HIF-1α inhibition \(^5,3^3\) and miR-204 restoration \(^1^3\) in PAH-PASMCs decrease intracellular \([Ca]_{2+}\) and reduce \(\Delta\Psi_m\), to a level similar to that seen in control-PASMC \((n=3, P<0.001)\). We also observed that the ABT-888 effect is similar to the effects with the other treatments. Moreover, adding ABT-888 to other treatments does not change \([Ca]_{2+}\), or \(\Delta\Psi_m\) (Figure 4D), demonstrating that there are no synergic effects and showing that these targets act in the same molecular pathway.

**In Vivo, ABT-888 Treatment Reverses Both Monocrotaline- and Sugen-Induced PAH**

As in human distal PAs, distal PAs of both Sugen-PAH and MCT-PAH rats had increased DNA damage, measured by γ-H2AX lung expression and 53BP1 immunohistochemistry (Figure 5A and Figure IVA in the online-only Data Supplement) and increased PARP-1 expression (Figure IIB and IIC in the online-only Data Supplement). As suggested by our in vitro studies, inflammation could account for the increase in DNA damage; inflammatory cytokine levels are increased (IL-6) in both Sugen-PAH and MCT-PAH lungs (Figure IVB in the online-only Data Supplement). Furthermore, NFkB lung protein expression and hematoxylin and eosin staining show increased inflammation in PAH models, and this inflammation seems to be decreased on ABT-888 treatment (Figure IVB in the online-only Data Supplement). To test whether PARP-1 inhibition can reverse PAH in our established rat models, ABT-888 was given orally (6 mg/kg per day) once PAH was established (5 weeks after Sugen injection or 14 days after MCT injection). We confirmed that ABT-888 administration per os decreases PARP-1 activity in the lungs by using a modified PARP-1 activity assay (Figure IIB in the online-only Data Supplement). We performed a 2-week longitudinal study to assess the efficacy of our treatment by using noninvasive hemodynamic assessment with Doppler echocardiography. We observed that in both Sugen-PAH and MCT-PAH rats, PARP inhibition reduces PA pressure assessed by PA acceleration time, a Doppler parameter linked to PA pressure (PA acceleration time being inversely correlated to PA pressure), and also decreased right ventricular hypertrophy (Figure VB in the online-only Data Supplement);

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**Figure 5. Clinically available PARP inhibitor ABT-888 reverses MCT- and Sugen-induced PAH.**

A, DNA damage was assessed in lungs of PAH models with or without ABT-888 treatment (6 mg/kg per day administered per os for 2 weeks once PAH is established). DNA damage, measured by γ-H2AX (immunoblot, n=3), is increased in PAH rat models' lungs and slightly increased with ABT-888 treatment. B, PARP-1 inhibition decreases PA pressure (measured right heart catheterization), RV hypertrophy (measured by Fulton index), and PA wall thickness (hematoxylin and eosin staining; n=6–10 rats per group). Because of the variance of these results with mean value (unequal variances), statistical analyses were performed on log transformation of the data. *P<0.05; **P<0.01; ***P<0.001. H&E indicates hematoxylin and eosin; LV, left ventricle; MCT, the monocrotaline model; mPAP, mean pulmonary artery pressure; PA, pulmonary artery; PAH, pulmonary arterial hypertension; PARP-1, poly(ADP-ribose) polymerase-1; RV, right ventricle; and S, septum.
n=8 rats per group, \( P<0.05 \); Figure VA in the online-only Data Supplement). These findings were invasively confirmed by right heart catheterization (direct PA pressure) and right ventricle/left ventricle with septum weight ratio (Figure 5B). Also, ABT-888 significantly improved exercise capacity in both models, using standard treadmill exercise protocol, compatible with the improvement of the hemodynamic measurements (Figure VA in the online-only Data Supplement). Furthermore, no kidney or liver toxicity was seen after ABT-888 treatment in both models, assessed by plasma creatinine and aspartate aminotransferase levels (Figure VB in the online-only Data Supplement).

To determine whether ABT-888 reduced PA remodeling in Sugen-PAH and MCT-PAH animals, we measured medial wall thickness by using hematoxylin and eosin staining. We observed that animals treated with ABT-888 displayed a significant reduction in medial thickness of distal PAs (≤ 100μm; Figure 5C). As in vitro, these findings were associated with a significant decrease in PASMC proliferation (as assessed by Ki67; Figure 6A) and resistance to apoptosis (terminal deoxynucleotidyl transferase dUTP nick end labeling; Figure 6B).

Finally, ABT-888 provides a therapeutic effect similar to the combination of both standard PAH therapies (bosentan+tadalafil) in MCT-PAH rats, whereas ABT-888 given in addition to bosentan and tadalafil provided greater therapeutic effects than in MCT-PAH rats treated with bosentan+tadalafil only. These findings suggest that ABT-888 could be used in humans in combination with current therapies and would provide a more efficient treatment (Figure VI in the online-only Data Supplement).

### Discussion

Using PAH lungs from both humans and rats, we showed for the first time that DNA damage may be etiologically implicated in PAH. Clinically, DNA damage has been suspected to be responsible for the occurrence of pulmonary hypertension in patients taking etoposide, an anticancer drug promoting DNA damage and PARP-1 activation.\(^{34}\) We showed that sustained DNA damage correlated with PARP-1 expression (Figure 1 and Figure I in the online-only Data Supplement), suggesting that it could modulate PARP-1 directly or indirectly. In addition to the DNA repair that may allow survival, PARP-1 appears to promote the activation of a pro-proliferative and anti-apoptotic program, characterized by miR-204 downregulation and the subsequent NFAT and HIF-1 upregulation, both of which have been shown to promote PAH-PASMC proliferation and resistance to apoptosis (Figures 3 and 4 and Figure III in the online-only Data Supplement). PARP-1 inhibition by using the clinically available inhibitor ABT-888 reverses all these abnormalities in vitro (Figures 3 and 4) and reverses PAH in vivo in 2 widely used models of PAH (Figure 5 and Figures V and VI in the online-only Data Supplement). A schematic representation of our signaling pathway is presented in Figure 7.

Although not previously reported, the presence of increased DNA damage in distal PAs in PAH in comparison with control patients is not surprising. PAH is characterized

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**Figure 6.** ABT-888 decreases proliferation and apoptosis resistance in PAH-PASMCs of distal PAs. 

A, ABT-888 treatment decreases smooth muscle cell proliferation (Ki67) in distal PAs of both animal models (n=5 rats per group). B, Apoptosis was assessed by using the TUNEL assay, and we showed that ABT-888 treatment increases apoptosis levels in distal PAs of both animal models. Only smooth muscle actin–positive cells were considered in the calculation of positive cells. These experiments were performed in 5 distal PAs per rat in 5 rats per group. \(*P<0.05; **P<0.01; ***P<0.001.\) DAPI indicates 4′,6-diamidino-2-phenylindole; MCT, the monocrotaline model; PA, pulmonary artery; PAH, pulmonary arterial hypertension; PARP-1, poly(ADP-ribose) polymerase-1; PASMC, pulmonary arterial smooth muscle cell; and TUNEL, terminal deoxynucleotidyl transferase dUTP nick end labeling.
PARP-1 may act as a sensor of intracellular stress. In the presence of extensive DNA damage (as seen after treatment with high doses with γ-radiation, for example), PARP-1 is highly activated and promotes cell death, whereas, in the presence of low to moderate DNA damage, as perhaps seen in PAH, PARP-1 triggers DNA repair by recruiting important repair factors to the DNA damage sites, allowing PASMCs to survive and even proliferate. Interestingly, recent studies have demonstrated that a mild to moderate PARP-1 activation can induce pro-proliferative responses by promoting the activation of NFAT, a transcription factor critical for PAH-PASMC proliferation. We previously reported that NFAT activation in PAH occurs downstream of the downregulation of miR-204 and the subsequent activation of STAT3 and Pim-1. In the present study, we report that PARP-1 inhibition is associated with a complete reversal of miR-204 downregulation, limiting NFAT activation, which decreases PAH-PASMC proliferation. In line with our findings, studies in hematopoietic cells and neurons have shown a significant increase in Pim-1 in response to DNA damage contributing to cell survival. In addition to miR-204 downregulation, upregulation of Pim-1 in PAH could contribute to the PARP-1–dependent cell survival in PAH-PASMCs. In addition, several recent studies have attributed numerous posttranscriptional functions to PARP proteins, including the regulation of the microRNA binding protein Argonaute 2. These could contribute to the activation of other pathways implicated in PAH. For example, Argonaute 2 is implicated in stem cell proliferation through the modulation of voltage-gated K+ channels including Kv1.5, which is implicated in PAH. In addition to proliferation, resistance to apoptosis is another important feature of PAH-PASMCs. Several reports have demonstrated that abnormal metabolic and mitochondrial dysfunction play a key role in this phenomenon, notably by promoting a normoxic activation of HIF-1α. In fact, studies have presented PARP-1 as a transcriptional coactivator of HIF-1α. On hypoxic induction of cells, PARP-1 was shown to interact with HIF-1α and to regulate the transcriptional activity of HIF-1α–dependent genes. In the present study, we showed that PARP-1 inhibition significantly decreases HIF-1α activity resulting in a significant induction of apoptosis. We recently showed that the normoxic upregulation of HIF-1α in PAH-PASMCs is attributed to endoplasmic reticulum stress, disrupting endoplasmic reticulum mitochondria units, impairing mitochondrial reactive oxygen species production resulting in HIF-1α activation. Recently in neurons, PARP-1 activation has been shown to promote endoplasmic reticulum stress. This could explain why PARP-1 inhibition in PAH-PASMCs decreases HIF-1α (Figure 4C) activation and restores mitochondria membrane potential (Figure 4D).
Further experiments are required to understand the relative importance of the several mechanisms accounting for PARP-1 activation (DNA damage, inflammation, oxidative stress, etc.) in PAH and to elucidate the pathways implicated in the beneficial effect of ABT-888 treatment in experimental PAH. Although our data support a role for DNA damage in PARP-1 activation, previous studies reported that proinflammatory factors including TNF-α might also contribute to its activation. Regarding the beneficial effects of PARP-1 inhibitors in experimental PAH, we provide evidence that PARP-1 inhibition upregulates miR-204 and subsequently reduces NFAT and HIF-1α activation in PAH-PASMCs. Nonetheless, the implication of other pathways cannot be ruled out. In fact, the potential role of PARP-1 as a stress sensor in PASMCs directly implies the involvement of multiple pathways, because many signals may converge to PARP-1 and produce several downstream signals, which may vary according to experimental conditions or disease states. For example, PARP-1 activation can promote cytokine production, suggesting that its inhibition may reduce the inflammatory response seen in PAH (as suggested in Figure IVB in the online-only Data Supplement) and thus improve vascular remodeling. This possibility will be addressed in future studies. Given the diversity of signaling mechanisms and protein expression between proximal and distal PAs, a limitation in the study is the use of human PASMCs isolated from some-what larger distal PAs (1000 μm in diameter), as opposed to the <300- to 500-μm vessels that are considered to be the typical resistance vessels. However, we managed to confirm our findings by using immunofluorescence and immunohistology measurements made in distal PAs (≤100 μm in diameter). In addition, there is emerging evidence that the proximal PAs may also be involved in PAH pathology, exhibiting remodeling that results in vascular stiffness, in contrast to earlier views suggesting that the primary signal for the disease is restricted within the resistance PAs. Also, the inflammatory environment in PAH, which is increasingly appreciated, implies that the more proximal PAs are equally exposed to circulating cytokines in comparison with the more distal arteries.

We focused our study on PASMCs and not endothelial cells, which are also implicated in PAH. Although most investigators agree that PA endothelial cell apoptosis is one of the earliest events in the pathogenesis of PAH in which DNA damage might be implicated, less is known about endothelial cell biology as the disease progresses; many suspect that there is a switch toward a proliferative and apoptosis-resistant phenotype. Although we did not study the possible PARP-1 effects in the biology of PAH-PA endothelial cells, it is possible that, as in PASMCs, increased PARP-1 marks a proliferative endothelial cell with downregulated miR-204 and activated STAT3, NFAT, and HIF-1α. Indeed, it has recently been shown that downregulated miR-204 and increased HIF-1α are found in plexiform lesions.

Although PARP-1 inhibition appears to be effective in terms of reversing established PAH in vivo, potential induction of apoptosis in other organs (following PARP-1 inhibition and thus accumulation of DNA damage) might be catastrophic, and this needs to be monitored before clinical translation. On the other hand, it is expected that PARP-1 inhibitors (several of which are actively used clinically, including ABT-888, AZD 2281, and MK4827) will have effects preferentially in tissues where PARP-1 is activated. All of these compounds are already in phase II or III clinical trials in cancer.

In conclusion, this is the first study to provide mechanistic evidence for the implication of DNA damage and the resulting induction of PARP-1 in the etiology of human PAH. In addition to its implication in DNA repair, PARP-1 may account for many PAH features, including impaired microRNA regulation, cell proliferation (through STAT3 and NFAT), and resistance to apoptosis (through HIF-1α; Figure 7). In that sense, PARP-1 may be one of the earliest factors contributing to the PAH phenotype. Finally, our findings support the translational potential of existing PARP-1 inhibitors as new therapies for PAH.

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Disclosures

None.

References


**CLINICAL PERSPECTIVE**

Pulmonary arterial hypertension (PAH) is characterized by progressive pulmonary vascular remodeling. Despite newly developed therapies, most patients eventually develop right ventricular failure and ultimately die. Advances in our understanding of the PAH pathophysiology is thus urgently needed to develop new therapeutic strategies and improve long-term outcomes in this devastating disease. Over the past few years, research in this field has focused on understanding the mechanisms of the enhanced pulmonary artery smooth muscle cell proliferation and suppressed apoptosis accounting for pulmonary vascular remodeling. Many abnormalities contribute to this phenotype, including increased activation of the transcription factors signal transducer and activator of transcription-3, nuclear factor of activated T cells, and hypoxia-inducible factor 1-α, and miR-204 downregulation, as well. In the present study, we demonstrated using a translational approach, that PAH is characterized by significant increase in DNA damage resulting in sustained activation of poly(ADP-ribose) polymerase-1 (PARP-1), a key component of the DNA repair machinery. PARP-1 activation promoted cell survival and proliferation by triggering a miR-204–dependent activation of nuclear factor of activated T cells and hypoxia-inducible factor 1-α, which was reversed by using PARP-1 inhibitors. PARP-1 activation may thus represent a common origin to many abnormalities promoting pulmonary vascular remodeling in PAH. PARP-1 inhibition was not only effective in vivo, but provided even greater efficacy in reversing PAH when combined with currently available therapies. Interestingly, PARP-1 is minimally expressed in the majority of healthy tissues, and phase 2/3 clinical trials investigating PARP-1 inhibitors in cancer are currently ongoing. Consequently, PARP-1 inhibition represents a novel and attractive therapeutic target for PAH.
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SUPPLEMENTAL MATERIAL

Supplemental Methods

Cell culture and treatments

PASMC were grown in high-glucose DMEM supplemented with 10% FBS (Gibco, Invitrogen, Burlington, ON, Canada) and 1% antibiotic/antimyotic (Gibco, Invitrogen, Burlington, ON, Canada). PARP was inhibited by a clinically relevant dose of ABT-888 (10 µM for 48h, Enzo life sciences). We also used a more specific PARP-1 inhibitor, AG14361 (5µM for 48h, Selleck Chemicals), to confirm the major implication of PARP-1. PASMC were transfected by CaPO₄ precipitation with 20nM small interfering RNA (siRNA) oligonucleotides (siPARP-1 (Ambion), siHIF-1α (Thermo scientific) or their negative control siSCRM (Ambion)). The Ca phosphate transfection method was also used to transfect miRIDAN miR-204 mimics (200nM for 48 h). NFATc2 was inhibited by a specific competitive peptide VIVIT 4 µM for 48 h (Calbiochem). Control PASMC were exposed to 100ng/mL TNF-α, 100µM IL-6 or 30ng/mL PDGF (all from EMB Canada) for 48 hours.

Quantitative RT-PCR and immunoblots

To measure PARP-1 mRNA expression, total mRNA was extracted from PASMC using a trizol protocol. Quantitative RT-PCR (qRT-PCR) was calculated with 18S as housekeeping gene (Taqman Gene expression Assay, Applied Biosystem, Foster, CA, USA) as previously described. All the experiments were performed in triplicate. To measure miR-204 expression, the mirVana kit (Applied Biosystems) was used to extract total RNA from PASMC. Stem-loop qRT-PCR for mature miRNAs was performed on a real-time PCR system (Applied Biosystems). For immunoblots, protein expression of PARP-1 (Santa Cruz, 1:500), γH2AX (Cell signaling and
Abcam, 1:500) and HIF-1α (Novus Biological, 1:1000) were quantified and normalized to amidoblack as previously described\(^4\). All immunoblots were performed in triplicate.

**PARP activity assay**

PARP activity was assayed using the Trevigen Universal chemiluminescent PARP assay kit according to the manufacturer’s instruction, with modifications. Cell lysate (40µg/well) was added to the wells containing PARP buffer and PARP cocktail. Activated DNA and PARP enzyme were not added to cell lysate, as previously described\(^5\).

**Proliferation and apoptosis measurements**

To study the effect of PARP-1 on PASMC proliferation and apoptosis in vitro, cultured human PAH-PASMC were exposed to 10% FBS (a condition that is known to promote proliferation\(^1,4\)) or 0.1% FBS (a starvation condition that promotes apoptosis\(^1,4\)). PASMC proliferation was measured using Ki67 antibody (Millipore) and PCNA (Dako) and apoptosis by using Apoptag apoptosis detection kit (TUNEL; Millipore) and AnnexinV (Clontech). Percent of positive PASMC (Ki67, TUNEL) were determined and divided by the total amount of cells, calculated with DAPI (total nuclei). WST1 assay, also to assess proliferation, was used according to the manufacturer’s instruction with modifications. Briefly, 2000 cells/well were plated onto 24-well plates. After stabilization, cells were treated for 48 hours under different conditions. The reagent was added to the medium and after 2h incubation; the absorbance at 440 nm was measured using a microplate reader (Synergy H1, BioTek). Fluo-3AM (Invitrogen) and TMRM (Tetramethylrodamine methyl ester perchlorate, Invitrogen) were measured in PASMC treated with different activators and inhibitors to evaluate the effect of modulating our studied pathway
on intracellular calcium and mitochondrial membrane potential (n=20 to 50 cells/experiment in 3 experiments).

**DNA damage**

Immunofluorescence for γ-H2AX and immunohistochemistry for 53BP1 were performed as previously reported in Rodrigue et al.⁶ using a monoclonal antibody directed against γ-H2AX (Cell Signaling Canada) and a polyclonal antibody against 53BP1 (Bethyl Canada).

**Confocal Microscopy/Immunofluorescence**

Sections of lungs and human PASMC were used for immunofluorescence staining. PASMC were fixed with 1% paraformaldehyde and permeabilized with 0.2% Triton X-100. For lung samples, only small and distal pulmonary arteries (<100 microns) were investigated. Rat lungs were fixed with 4% paraformaldehyde. Immunofluorescence was performed on 5µm lung slices. PARP-1 (Santa Cruz, 1:100), γ−H2AX (Cell signaling, 1:50), NFATc2 (Abcam; 1:250), HIF-1α (Novus Biologicals, 1:250), smooth muscle actin (Sigma, 1:250) primary antibodies were used and Alexa Fluor 488 and 594 were used as secondary antibodies. Co-localization studies were performed using the Zen system from Zeiss. Briefly, stack imaging (10 pictures taken at random on 4 micrometer thick slides) was analyzed and all co-localizations detected throughout the stack were shown in yellow. When measuring DNA damage in vitro, cells were classified based on the number of foci in the nucleus: cells with greater than 10 foci were counted as positive according to the standard procedure² and graphs represent the percent of positive cells (n=20 to 50 cells/experiment in 3 experiments).
Animal models

Male Sprague-Dawley rats (250–350g) (strain 400, Charles River) were randomly distributed between the groups and all the hemodynamic measurements were performed blinded to the condition. For the Sugen-hypoxia model (Sugen), rats were injected with 20mg/kg in of SU5416 (Sigma) and put in hypoxia (10% O₂) for 3 weeks. Chambers were opened twice a week for cleaning and replenishment of food and water. Oxygen concentrations were continuously monitored with blood gas analyzers. For the monocrotaline-induced PAH model (MCT), rats were injected s.c. with 60 mg/kg of crotaline (Sigma). PAH progression was monitored every week by non-invasive echocardiography measuring the pulmonary artery acceleration times (PAAT) and the right ventricular (RV) hypertrophy. At the end of the treatment period, all rats underwent right heart catheterizations (closed chest) using SciScence catheters to measure pulmonary arterial pressure, as previously described⁴,⁷. Histology measurements were performed as previously described³.⁸. Pulmonary artery wall thickness was measured using 2 measurements/artery in 10 arteries/animal, in at least 5 animals per group. We also performed experiments on MCT-induced PAH in rats to evaluate the therapeutic potential of ABT-888 (6mg/kg/day) compared to actual treatments used in PAH (Bosentan (100mg/kg/day) and Tadalafil (10mg/kg/day))⁹ and all these treatments started 2 weeks after MCT injection (when PAH was established) and were diluted in water to be administered per os. Drinking water was used in all conditions as the vehicle group.
Supplemental Figure Legends

Supplemental Figure 1: DNA damage and PARP-1 expression

a) We confirmed that no staining appeared on our immunohistochemistry negative control. b) In PAH-PASMC, AG14361 stimulates PARP-1 expression (as ABT-888 does). We also confirmed siPARP-1 (20nM) efficiency, as no PARP-1 expression is seen on the immunoblot. c) We demonstrated that other pro-PAH factors, IL-6 (100µM) and PDGF (30ng/mL), also induce DNA damage in control cells (measured by γ-H2AX staining; n=3, **p<0.001).

Supplemental Figure 2: PARP-1 upregulation is limited to the lungs

a) PARP-1 is overexpressed in human lung tissues (n=5 controls and 6 PAH patients), but not in human kidney or brain (n=6 controls and 4 PAH patients). b) PARP-1 expression is increased in lungs of both PAH models used: the monocrotaline (MCT) and the Sugen (with hypoxia)-induced PAH (n=5 rats per group). We confirmed that ABT-888 administration per os decreases PARP-1 activity in the lungs, using the modified PARP-1 activity assay (n=5 rats per group). c) This increased PARP-1 expression is located in the lungs, as no increase is observed in left ventricle or in the liver (n=5 rats per group). *p<0.05 and ***p<0.001

Supplemental Figure 3: PARP inhibition decreases proliferation and increases apoptosis

a) PARP-1 inhibition in PAH-PASMC with ABT-888 (10µM for 48 hours) decreases proliferation, measured by PCNA (n=4). b) Similarly, ABT-888 restores apoptosis levels in serum-starved media measured by AnnexinV (n=4). *p<0.05; **p<0.01 and ***p<0.001

Supplemental Figure 4: DNA damage and inflammation are present in PAH animal models
a) DNA damage (53BP1) was quantified in distal PAs (8 rats per group, 5 PAs/lung). Cells with positive nuclear staining were considered positive and this was divided by the total amount of cells (total nuclei). As in human PAH distal PAs express more DNA damage and, as *in vitro*, DNA damage in maintained with ABT-888 treatment. b) Both animal models show increased inflammation, measured by circulating IL-6 levels (a pro-inflammatory cytokine) and visualized in H&E staining. (n=3 per group). Furthermore, NFκB protein expression, an inflammatory marker, is increased in Sugen-PAH lungs and decreased when rats were treated with ABT-888. *p<0.05; **p<0.01 and ***p<0.001

**Supplemental Figure 5: Clinically available PARP inhibitor ABT-888 reverses PAH in two experimental rat models**

a) ABT-888 (6mg/Kg/day) *per os* given for 2 weeks once PAH established (2 weeks post MCT injection and 5 weeks post Sugen injection) efficiently decreases PA pressure (right heart catheterization) and RV hypertrophy (RV/LVS ratio) (n= a minimum of 6 rats per group). B) Echography and treadmill data taken at the end of protocols also demonstrate a beneficial effect of ABT-888 treatment of lung and heart function (echography) as well as exercise capacity (treadmill). b) Creatinine and aspartate aminotransferase (AST) levels were measured in plasma to assess liver and kidney toxicity (n=5 per group). Since variance of these results increases with mean value (unequal variances), statistical analyses were performed on log transformation of the data. *p<0.05; **p<0.01 and ***p<0.001

**Supplemental Figure 6: Comparison between ABT-888 and actual therapies**
Different treatments were administered starting at week 2 in the MCT-PAH model. ABT-888 (6mg/kg/day), Bosentan (100mg/kg/day) and Tadalafil (10mg/kg/day) were all diluted in water and administered per os (oral administration) on a daily basis. Right heart catheterization performed after two weeks of treatment shows that ABT-888 add to the beneficial effect of the current cocktail therapy. Since variance of these results with mean value (unequal variances), statistical analyses were performed on log transformation of the data. *p<0.05; **p<0.01 and ***p<0.001
Supplemental References


Supplemental Figure 1

A  Immunohistochemistry negative control

53BP1 in brown and nuclei in blue (100X)

B  AG14361 and siPARP-1 effect on PAH-PASMC

PARP-1

Amidoblock

116 kDa
45 kDa

C  Pro-PAH factors induce DNA damage

\( \gamma H2AX \) in green and DAPI in blue (63x)
A PARP-1 expression is not increased elsewhere than lungs

B PARP-1 expression is increased PAH lungs and ABT treatment decreases its activity

C PARP-1 expression is not increased elsewhere than lung tissues (MCT model)
**A** PARP-1 inhibition inhibits the pro-proliferative phenotype

Control  | PAH  | PAH+ABT-888
---|---|---
PCNA in red and DAPI in blue (20x)

% PCNA positive cells

- Control
- PAH
- PAH+ABT-888

**B** PARP-1 inhibition restores apoptosis levels

Control  | PAH  | PAH+ABT-888
---|---|---
Annexin V in green and DAPI in blue (20x)

% AnnexinV positive cells

- Control
- PAH
- PAH+ABT-888
Supplemental Figure 4

A DNA damage is also present in PAH animal models regardless of ABT-888 treatment

B Inflammation is present in both animal models

NFκB protein expression in lungs

Control | Sugen-PAH | Sugen-PAH + ABT-888

NFκB p65
Stain free

kDa
65
45

Control | Sugen-PAH | Sugen-PAH + ABT-888

H&E (10X)
Supplemental Figure 5

A Follow-up measurements for both animal models

**Pulmonary artery acceleration time**

**RV free wall**

**Treadmill**

B Indirect measurements of renal and liver health

**Creatinine**

**Aspartate aminotransferase (AST)**
Supplemental Figure 6

A Adding ABT-888 to actual therapies reduces pulmonary pressures