Vascular Medicine

Hydrogen Peroxide Promotes Aging-Related Platelet Hyperactivation and Thrombosis

Sanjana Dayal, PhD; Katina M. Wilson, BS; David G. Motto, MD, PhD; Francis J. Miller, Jr, MD; Anil K. Chauhan, PhD; Steven R. Lentz, MD, PhD

Background—The incidence of thrombotic events increases during aging, but the mechanisms are not well understood. To investigate the prothrombotic role of oxidative stress during aging, we tested the hypothesis that aged mice overexpressing the antioxidant enzyme glutathione peroxidase-1 (Gpx1) are protected from experimental thrombosis.

Methods and Results—Susceptibility to carotid artery thrombosis was first examined in wild-type C57BL/6J mice. After photochemical injury of the carotid artery, the time to stable occlusion was significantly shorter in 12- and 18-month-old mice compared with 4-month-old mice (P<0.01). Unlike wild-type mice, transgenic mice overexpressing Gpx1 (Gpx1 Tg) did not exhibit shortened times to occlusion of the carotid artery at 12 or 18 months of age. Wild-type mice also exhibited increased susceptibility to venous thrombosis after inferior vena cava ligation at 12 or 18 months of age (P<0.05 versus 4 months of age). Gpx1 Tg mice were protected from this aging-related enhanced susceptibility to venous thrombosis. Age-dependent platelet hyperactivation, evidenced by increased hydrogen peroxide, fibrinogen binding, and activation of fibrinogen receptor αIIbβ3, was observed in thrombin-activated platelets from wild-type but not Gpx1 Tg mice (P<0.05). Enhanced platelet activation responses in aged mice were also prevented by polyethylene glycol–catalase or apocynin, an inhibitor of NADPH oxidase. Aged mice displayed increased intraplatelet expression of p47phox and superoxide dismutase-1, suggesting a mechanistic pathway for increased hydrogen peroxide generation.

Conclusions—Our findings demonstrate that hydrogen peroxide is a key mediator of platelet hyperactivity and enhanced thrombotic susceptibility in aged mice. (Circulation. 2013;127:1308-1316.)

Key Words: aging ■ blood platelets ■ thrombosis

Thrombotic events such as stroke, myocardial infarction, deep vein thrombosis, and pulmonary embolism are common causes of morbidity and mortality in the elderly. Despite the well-established clinical associations between aging and thrombosis, surprisingly little is known about the mechanisms by which aging increases susceptibility to thrombotic events.

Clinical Perspective on p 1316

Several lines of evidence indicate that aging is accompanied by a generalized increase in oxidative stress and that increased generation of reactive oxygen species may contribute to cardiovascular events, including thrombosis. A marked increase in oxidative stress has been observed with aging in multiple tissues in humans and experimental animals. Increased oxidative stress has been mechanistically implicated in several of the cardiovascular consequences that are also associated with aging such as myocardial dysfunction, myocardial ischemia/reperfusion injury, hypertension, and endothelial dysfunction. Moreover, there is abundant evidence that reactive oxygen species regulate several components of thrombotic processes, including platelet activation. To date, however, no studies have directly investigated the mechanistic contribution of reactive oxygen species to aging-related thrombosis.

The increased reactive oxygen species production associated with aging has been attributed in part to decreased expression and activity of antioxidant enzymes such as glutathione peroxidase, which reduces and detoxifies peroxides like hydrogen peroxide (H₂O₂). Glutathione peroxidase-1 (Gpx1) is the most abundant and widely expressed isoform of glutathione peroxidase in most tissues. A prospective human study demonstrated that the risk of cardiovascular events was inversely associated with increasing quartiles of erythrocyte Gpx1 activity. This observation suggests that peroxides or Gpx1 plays a role in the increased thrombotic susceptibility of aging.

In the present study, we compared thrombotic responses to arterial or venous injury in young versus aged mice and examined the protective effects of overexpression of Gpx1.
findings suggest that, in aging, peroxides mediate enhanced arterial and venous thrombosis and platelet hyperactivity. Our data also suggest that strategies to lower platelet H$_2$O$_2$ levels abrogate the enhanced activation of platelets from aged mice, providing mechanistic insights and a potential therapeutic approach to prevent aging-related thrombosis.

Methods

Mice
C57BL/6 mice were purchased from Jackson Laboratory at the age of 3 months and maintained in the animal care facilities of the University of Iowa. Gxp1 transgenic (Tg) mice were obtained from Dr Yi Shi Ho$^{25}$ and bred at least 15 generations with C57BL/6 mice before study. Genotyping for the Gpx1 transgene was performed with real-time polymerase chain reaction (PCR).$^{26}$ All animal protocols were approved by the University of Iowa Animal Care and Use Committee. Both male and female mice were included in the study. Mice at 4, 12, and 18 months of age were used for studies. These age groups were chosen in accordance with accepted principles for experiments on the biology of aging in mice.$^{27}$ Because the mean life span of C57BL/6 mice is $\approx 27$ to 29 months, the 4-, 12-, and 18-month-old mice are roughly equivalent to young adult (18–20 years of age), middle-aged ($\approx 45$–50 years of age), and older ($>70$ years of age) humans, respectively.

Carotid Artery Thrombosis
Carotid artery thrombosis was induced by photochemical injury as described previously.$^{28}$ Mice were anesthetized with sodium pentobarbital (70–90 mg/kg IP) and ventilated mechanically with room air and supplemental oxygen. To induce photochemical injury to the endothelial layer, the right common carotid artery was dissected free and transilluminated continuously with a 1.5-mW, 540-nm green laser (Melles Griot, Carlsbad, CA) from a distance of 6 cm, and rose bengal (35 mg/kg) was injected via a femoral vein catheter. Blood flow was monitored continuously for 90 minutes or until stable occlusion occurred, at which time the experiment was terminated. Stable occlusion was defined as the time at which blood flow remained absent for $\geq 10$ minutes.

Inferior Vena Cava Thrombosis
Susceptibility to thrombosis in the venous system was measured as described previously with minor modifications.$^{29}$ Briefly, mice were anesthetized with ketamine/xylazine (87.5 mg/kg ketamine and 12.5 mg/kg xylazine IP). A midline laparotomy was made, and the inferior vena cava was exposed directly via blunt dissection. The inferior vena cava was ligated inferiorly to the left renal vein with a 6-0 silk suture, and mice were allowed to recover. Two days later, the inferior vena cava was harvested for measurement of the length and weight of thrombus.

Platelet Activation
Washed platelets were isolated and resuspended in modified Tyrode buffer (134 mmol/L NaCl, 2.9 mmol/L KCl, 2.9 mmol/L CaCl$_2$, 0.34 mmol/L Na$_2$HPO$_4$, 12 mmol/L NaHCO$_3$, 20 mmol/L HEPES, 1.0 mmol/L MgCl$_2$, 5.0 mmol/L glucose, 0.05% [wt/vol] fatty acid–free BSA, pH 7.35) as described previously.$^{30}$ To assess platelet activation, washed platelets were activated with human thrombin (0.5 U/mL; Hematological Technologies, Essex Junction, VT) for 2 minutes at 37°C; incubated for 10 minutes with FITC-conjugated sheep anti-human fibrinogen antibody (Novus Biologicals, Littleton, CO), FITC-conjugated rat anti-mouse CD62P antibody for P-selectin (BD Biosciences), or PE-conjugated JON/A for activated cIIB/3 (Emret Analytics, Würzburg, Germany); fixed in 1% paraformaldehyde; diluted; and analyzed by a Becton Dickinson (San Diego, CA) FACScan flow cytometer. In some experiments, platelets were incubated with polyethylene glycol (PEG)-catalase (500 U/mL), apocynin (600 µmol/L), or N-nitro-l-arginine methyl ester (L-NAME; 100 µmol/L) for 15 minutes before stimulation with thrombin.

Measurement of Intracellular H$_2$O$_2$ in Platelets
Levels of platelet-derived intracellular H$_2$O$_2$ were measured as described previously with minor modifications.$^{31}$ Platelets were preincubated with 10 µmol/L 2’,7’-dichlorodihydrofluorescein diacetate (H$_2$DCF-DA; Molecular Probes, Gottingen, Germany) in either the presence or absence of PEG-catalase (500 U/mL) for 15 minutes. Platelets were then activated with 0.5 U/mL human thrombin, and the PEG-catalase–inhibitable fluorescent signal formed as a result of oxidation of H$_2$DCF-DA was measured by flow cytometry.

Real-time PCR
Levels of mRNA for Gpx1, Nox1, Nox2, Nox4, p47phox, Sod1, catalase, and 18S were measured by quantitative real-time PCR as described previously.$^{32}$ Total RNA was isolated from washed platelets with Trizol reagent (Invitrogen, Carlsbad, CA). Reverse-transcribed cDNA was incubated with TaqMan Universal PCR mix, PCR primers, and 6-carboxy fluorescein-labeled probes (Applied Biosystems) at 50°C for 2 minutes and then at 95°C for 10 minutes, followed by 40 cycles of 95°C for 15 seconds and 60°C for 1 minute. The comparative threshold cycle (ΔΔC$_t$) method was used for quantification with values normalized to 18S and expressed relative to levels in 4-month-old wild-type (WT) mice. Validation experiments were performed to confirm equal amplification efficiency for all primer sets. Platelet purity was confirmed by 2 methods: complete blood counting and real-time PCR with primers for CD45, a marker for leukocytes. All samples used for real-time PCR had undetectable levels of CD45 mRNA, and the leukocyte and red blood cell counts were equivalent to background.

Platelet Count
Blood was collected by retro-orbital bleeding into a 20-µL EDTA-coated glass capillary tube and immediately diluted 1:10 into PBS with 5% BSA and analyzed by a laser-based Bayer Advia 120 whole-blood analyzer.

Statistical Analysis
One-way ANOVA with the Tukey test for multiple comparisons was used to compare occlusion time, baseline blood flow, platelet counts, platelet activation responses before and after treatment with inhibitors, and mRNA levels in C57BL/6 or WT mice of different ages. Two-way ANOVA with multiple comparisons was used to compare time to occlusion, thrombus length and weight, H$_2$O$_2$ level, platelet activation response, platelet count, and baseline blood flow in Gxp1 Tg mice and WT littermate control mice. The paired Student t test was performed to compare platelet activation responses before and after treatment with inhibitors. All data were normally distributed except for the data in Figure 1; therefore, 1-way ANOVA was performed on log-transformed values for this data set. Statistical significance was defined as a value of P<0.05. All the values are reported as mean±SE.

Results

Susceptibility to Carotid Artery Thrombosis Is Increased in C57BL/6 Mice With Advancing Age
We first examined the relationship between age and susceptibility to arterial thrombosis in WT C57BL/6 mice (Figure 1). After chemical injury of the carotid artery, the time to stable occlusion in 4-month-old mice was 52.5±9.3 minutes, whereas the time to develop stable occlusion was significantly shorter for both 12- and 18-month-old mice (15.9±3.2 and 15.6±3.7 minutes, respectively; P<0.01).

Dayal et al
differences in baseline carotid artery blood flow were observed among 4-, 12-, and 18-month-old mice (Table 1). Platelet count did not differ significantly between 4- and 12-month-old mice but was ≈30% higher in 18-month-old mice (P<0.05 versus 4- and 12-month-old mice). These findings demonstrate that C57BL/6 mice exhibit increased susceptibility to carotid artery thrombosis with aging.

Overexpression of Gpx1 Protects Aged Mice From Accelerated Arterial and Venous Thrombosis

We next examined the role of oxidative stress in accelerated carotid artery thrombosis in aging using Gpx1 Tg mice and their WT littermates at 4, 12, and 18 months of age (Figure 2). Consistent with the findings in C57BL/6 mice, the time to develop stable occlusion of the carotid artery was significantly shorter in 12- and 18-month-old WT mice compared with 4-month-old WT mice (17.7±2.8, 16.9±4.2, and 56.2±1 minutes, respectively; P<0.05). At 4 months of age, the time to stable occlusion was similar in WT and Gpx1 Tg mice (56.2±10 and 41.7±9.6 minutes, respectively; P≈0.35), suggesting that, at a young age, overexpression of Gpx1 does not influence susceptibility to arterial thrombosis. However, the 12- and 18-month-old Gpx1 Tg mice were protected from the age-induced shortening of the time to stable occlusion (37.6±8.6 and 56.3±11.4 minutes, respectively, compared with age-matched WT littermates; P<0.05). Of note, the times to stable occlusion in the 12- and 18-month-old Gpx1 Tg mice did not differ significantly from that of 4-month-old Gpx1 Tg mice (P=0.9 and P=0.4, respectively). There was a significant overall interaction between age and genotype by 2-way ANOVA (P<0.017), which reflects the lack of effect of the Gpx1 Tg genotype on stable occlusion in 4-month-old mice.

No significant differences in baseline carotid artery blood flow were observed between 4-, 12-, and 18-month-old WT or Gpx1 Tg mice (Table 2). Platelet counts were again found to be higher in 18-month-old WT or Gpx1 Tg mice compared with 4- or 12-month-old mice of the same genotype (P<0.05). There were no significant differences in platelet count between WT and Gpx1 Tg mice at any age.

We next performed inferior vena cava ligation to examine whether Gpx1 overexpression provides protection from stasis-induced venous thrombosis (Figure 3). At 4 months of age, WT and Gpx1 Tg mice developed thrombi of similar length and weight (P>0.8). This observation suggests that overexpression of Gpx1 at a young age does not affect susceptibility to venous thrombosis. In contrast, the 12- and 18-month-old WT mice developed significantly larger and heavier inferior vena cava thrombi compared with 4-month-old WT mice (P<0.05). Overexpression of Gpx1 protected against this age-dependent effect, as evidenced by the significantly smaller thrombi developed in 12- and 18-month-old Gpx1 Tg mice compared with age-matched WT littermates (P<0.05). Collectively, these findings demonstrate that mice develop increased

Table 1. Baseline Carotid Artery Blood Flow and Platelet Count in C57BL/6 Mice

<table>
<thead>
<tr>
<th>Age Group</th>
<th>4 mo</th>
<th>12 mo</th>
<th>18 mo</th>
</tr>
</thead>
<tbody>
<tr>
<td>Baseline blood flow, mL/min</td>
<td>0.24±0.03</td>
<td>0.22±0.02</td>
<td>0.20±0.05</td>
</tr>
<tr>
<td>Platelet count, ×1000/µL</td>
<td>1393±66</td>
<td>1445±77</td>
<td>1835±146*</td>
</tr>
</tbody>
</table>

Eight to 12 mice were studied in each group. *P<0.05 versus 4-month-old mice by 1-way ANOVA.

Table 2. Baseline Carotid Artery Blood Flow and Platelet Count in WT and Gpx1 Tg Littermates

<table>
<thead>
<tr>
<th>Age Group</th>
<th>4 mo</th>
<th>12 mo</th>
<th>18 mo</th>
</tr>
</thead>
<tbody>
<tr>
<td>Baseline blood flow, mL/min</td>
<td>0.23±0.04</td>
<td>0.21±0.02</td>
<td>0.19±0.05</td>
</tr>
<tr>
<td>Gpx1 Tg</td>
<td>0.22±0.02</td>
<td>0.21±0.02</td>
<td>0.18±0.03</td>
</tr>
<tr>
<td>Platelet count, ×1000/µL</td>
<td>1360±53</td>
<td>1474±61</td>
<td>1747±112*</td>
</tr>
<tr>
<td>Gpx1 Tg</td>
<td>1286±47</td>
<td>1437±104</td>
<td>1880±72*</td>
</tr>
</tbody>
</table>

Tg indicates transgenic; and WT, wild-type. Six to 11 mice were studied in each group. *P<0.05 versus 4- and 12-month-old mice by 2-way ANOVA.
susceptibility to both arterial and venous thrombosis with advancing age and that overexpression of Gpx1 protects against these adverse effects.

**Platelets From Aged Mice Exhibit Increased Platelet Adhesion During Thrombus Formation In Vivo**
Because H$_2$O$_2$ has been reported to contribute to platelet activation responses, we hypothesized that the protective effect of overexpression of Gpx1 may be mediated through an effect on platelets. To address this hypothesis, platelets isolated from 4- or 18-month-old mice were differentially fluorescently labeled and infused into 4-week-old recipient mice. Thrombus formation in mesenteric arterioles was initiated with 5% ferric chloride, and the relative adhesion of young versus old platelets was quantified in real time by intravital microscopy. The results (Figure IA and IC and Movie I in the online-only Data Supplement) demonstrate that platelets from 18-month-old mice exhibit increased adhesion during thrombus formation in vivo compared with platelets from 4-month-old mice ($P<0.05$). There was also a trend toward decreased adhesion of infused platelets from aged (18-month-old) Gpx1 Tg mice compared with platelets from 18-month-old WT mice (Figure IB and ID and Movie II in the online-only Data Supplement).

**Activated Platelets From Aged Mice Generate Increased Levels of H$_2$O$_2$**
To further investigate the role of H$_2$O$_2$ in platelet activation during aging, platelets were stimulated with thrombin in vitro. We found that the intracellular concentration of H$_2$O$_2$ in thrombin-stimulated platelets was significantly higher in 18-month-old WT mice than in 4-month-old WT mice ($P<0.05$; Figure 4). In contrast, platelet H$_2$O$_2$ levels in 18-month-old Gpx1 Tg mice were similar to those in 4-month-old Gpx1 Tg mice ($P>0.8$). These data suggest that aging produces an elevation in levels of H$_2$O$_2$ within activated platelets and that overexpression of Gpx1 protects against this response. To confirm that Gpx1 is overexpressed in platelets from Gpx1 Tg mice, we measured levels of Gpx1 mRNA and Gpx1 protein in isolated platelets by quantitative PCR and Western blotting, respectively. Regardless of age, Gpx1 Tg mice had a large increase in both Gpx1 mRNA and Gpx1 protein levels in platelets compared with WT littermates ($P<0.05$; Figure IIA–IIC in the online-only Data Supplement).

**Aging Is Associated With H$_2$O$_2$-Dependent Inside-out Platelet Activation**
To determine whether increased generation of platelet H$_2$O$_2$ causes enhanced platelet activation in aged mice, we next examined surface P-selectin expression, fibrinogen binding, and integrin αIIbβ3 activation in thrombin-stimulated platelets from 4- and 18-month-old mice. No effects of advancing age or Gpx1 overexpression on P-selectin expression were observed (Figure III in the online-only Data Supplement), which suggests that the increased levels of platelet H$_2$O$_2$ in aged mice do not influence platelet granule release. In contrast, we observed an age-dependent increase in fibrinogen binding and αIIbβ3 activation in platelets from WT mice ($P<0.05$; Figure 5A and 5B). The increases in fibrinogen binding and αIIbβ3 activation were absent in 18-month-old Gpx1 Tg mice ($P<0.05$), suggesting a peroxide-dependent mechanism. To confirm the role of H$_2$O$_2$ versus other peroxides that are reduced by Gpx1, platelets from 4- and 18-month-old WT mice were treated with PEG-catalase, which selectively reduces H$_2$O$_2$. We found that PEG-catalase attenuated thrombin-stimulated fibrinogen binding and αIIbβ3 activation at both 4 and 18 months of age ($P<0.05$).
and completely abolished the age-dependent increased platelet activation in mice (Figure 5C and 5D). Taken together, these findings suggest that H$_2$O$_2$ is a critical mediator of increased inside-out activation of integrin αIIbβ3 (the major receptor for fibrinogen), leading to hyperactivation of platelets in aged mice.

**A Potential Mechanistic Pathway Leading to H$_2$O$_2$-Mediated Platelet Hyperactivation**

To identify potential mechanisms of H$_2$O$_2$-mediated platelet hyperactivation, we performed quantitative real-time PCR to measure mRNA levels of NADPH oxidase subunits and superoxide dismutase-1 (Sod1) in platelets. We found that platelets express significant levels of mRNA for the NADPH oxidase catalytic subunit Nox2 (Figure 6A) but do not express detectable levels of Nox1 or Nox4 mRNA at either 4 or 18 months of age (data not shown). We also observed a significant increase in platelet mRNA levels of the NADPH oxidase regulatory subunit p47$^{phox}$ ($P<0.05$; Figure 6B) in 18-month-old mice compared with 4-month-old mice. Furthermore, we observed an increase in Sod1 mRNA in platelets from aged mice ($P<0.05$; Figure 6C). These findings suggest a potential mechanistic pathway involving NADPH oxidase and SOD1 for increased generation of H$_2$O$_2$ in platelets from aged mice.

In the present study, we examined the role of H$_2$O$_2$ in the mechanism of increased susceptibility to arterial and venous thrombosis with aging. The main findings from this study are the following: (1) With advancing age, C57BL/6 mice become increasingly susceptible to both arterial and venous thrombosis; (2) overexpression of the peroxide-reducing enzyme Gpx1 protects against the accelerated thrombosis of aging; and (3) platelet activation responses are increased in aging, and platelet hyperresponsiveness is mediated by H$_2$O$_2$. Taken together, these findings suggest a mechanism in which increased H$_2$O$_2$ production leads to platelet hyperactivity and enhanced susceptibility to thrombosis in aging.

**Discussion**

In the present study, we examined the role of H$_2$O$_2$ in the mechanism of increased susceptibility to arterial and venous thrombosis with aging. The main findings from this study are the following: (1) With advancing age, C57BL/6 mice become increasingly susceptible to both arterial and venous thrombosis; (2) overexpression of the peroxide-reducing enzyme Gpx1 protects against the accelerated thrombosis of aging; and (3) platelet activation responses are increased in aging, and platelet hyperresponsiveness is mediated by H$_2$O$_2$. Taken together, these findings suggest a mechanism in which increased H$_2$O$_2$ production leads to platelet hyperactivity and enhanced susceptibility to thrombosis in aging.

One advantage of animal models of thrombosis, as opposed to human association studies, is that the contribution of aging can be assessed independently of other cardiovascular risk factors.
One novel finding of our study is that aged mice overexpressing Gpx1 were protected from enhanced thrombotic susceptibility, which implicates H$_2$O$_2$ and/or lipid peroxides in the prothrombotic phenotype of aging. Previous studies in humans have suggested that prothrombotic conditions such as diabetes mellitus are associated with increased platelet H$_2$O$_2$ production and enhanced platelet activation responses.$^{25,26}$ In concordance with these previous studies in humans, we found that intraplatelet H$_2$O$_2$ levels were elevated in activated platelets from aged mice with increased thrombotic susceptibility. We also found that platelets from aged mice exhibited increased adhesion to injured mesenteric arterioles in an in vivo model of thrombosis and enhanced activation responses to thrombin in vitro, in conjunction with increased accumulation of H$_2$O$_2$. Platelet hyperactivation was overcome not only by overexpression of Gpx1 but also by treatment with PEG-catalase, which implicate platelet H$_2$O$_2$ rather than lipid peroxides in the mechanism of enhanced platelet activation with aging.

We explored several possible mechanisms for the increased accumulation of H$_2$O$_2$ in activated platelets from aged mice. We first considered decreased Gpx1 expression in platelets as a potential mechanism because decreased expression of Gpx1 has been reported to be associated with advancing age in several other tissues.$^{20,21,23,37}$ and because decreased erythrocyte Gpx1 activity is associated with increased cardiovascular risk in patients with coronary artery disease.$^{24}$ However, we did not observe any differences in platelet Gpx1 mRNA or protein levels between young and old mice (Figure II in the online-only Data Supplement). We also did not observe a decrease in the expression of catalase mRNA in platelets from aged mice (data not shown). These findings suggest that the increase in platelet H$_2$O$_2$ in aged mice is not caused by decreased expression of Gpx1 or catalase.

We next considered the potential roles of NADPH oxidase and superoxide dismutase as enzymatic sources of elevated H$_2$O$_2$ in platelets from aged mice. NADPH oxidase-dependent generation of superoxide has been reported to regulate platelet integrin $\alpha$I$\beta$3 activation.$^{31}$ Additionally, platelets from patients with chronic granulomatous disease resulting from a deficiency of the NADPH oxidase catalytic subunit Nox2 have almost complete suppression of platelet superoxide production.$^{38,39}$ Because H$_2$O$_2$ can be generated from superoxide by superoxide dismutase, we hypothesized that increased generation of H$_2$O$_2$ in platelets from aged mice results from increased NADPH oxidase-dependent production of superoxide that is subsequently converted to H$_2$O$_2$ by superoxide dismutase. In agreement with this hypothesis, we found that preincubation of platelets from aged mice with the NADPH oxidase inhibitor apocynin resulted in a significant decrease in thrombin-induced activation of $\alpha$I$\beta$3 and surface fibrinogen binding. We also observed increased expression of mRNA for the NADPH oxidase regulatory subunit p47$^\text{phox}$, as well as $\text{Sod1}$, in platelets from aged mice. Furthermore, we considered the possibility that platelet NADPH oxidase might directly generate H$_2$O$_2$ because isoforms of NADPH oxidase that contain the Nox4 catalytic subunit can generate significant amounts of H$_2$O$_2$.$^{40,41}$ We consider this to be unlikely, however,
because we did not detect any significant expression of Nox4 mRNA in platelets from young or old mice. Unlike Nox4-containing NADPH oxidases, Nox2-containing NADPH oxidases usually generate superoxide with little or no direct generation of H$_2$O$_2$.\textsuperscript{3,4} Future studies using mice deficient in p47\,p22 or Nox2 might provide additional mechanistic insights into the role of platelet NADPH oxidases in platelet hyperactivity and thrombosis.

Interestingly, previous work by Freedman and colleagues\textsuperscript{42} and Jin et al\textsuperscript{43} has demonstrated that deficiency of plasma glutathione peroxidase (glutathione peroxidase-3, Gpx3) causes platelet hyperactivation in both humans and mice. The proposed mechanism was that deficiency of Gpx3 may lead to increased extracellular levels of H$_2$O$_2$, resulting in decreased bioavailability of NO and decreased NO-mediated inhibition of platelet activation. Our findings suggest that regulation of intracellular H$_2$O$_2$ by Gpx1 also affects platelet activation responses. The protective effect of Gpx1 is likely independent of platelet-derived NO because we did not observe any inhibition of platelet activation in the presence of the NO synthase inhibitor L-NAME (Figure 7). Although our data suggest a direct, NO-independent effect of aging on H$_2$O$_2$-mediated platelet activation, we recognize that aging is also associated with increased oxidative stress in the vessel wall and that decreased endothelium-derived NO may contribute in part to increased thrombotic susceptibility. In addition, other vascular cell-derived thrombotic and inflammatory factors such as plasminogen activator inhibitor-1, P-selectin, and tissue factor\textsuperscript{49} also may play a role in the mechanism by which increased oxidative stress enhances susceptibility to thrombosis during aging.

Given the well-established role of platelets in arterial thrombosis as opposed to venous thrombosis, it is perhaps surprising that we observed similar effects of aging and Gpx1 overexpression in experimental models of both arterial and venous thrombosis. However, the paradigm that platelets are more important in arterial than venous thrombosis has been challenged by recent work in mouse models\textsuperscript{54,55} and by human studies in which aspirin has proven to have benefit in the prevention of recurrent venous thromboembolism.\textsuperscript{6,7} Our work presented here is consistent with these recent findings and suggests a possible mechanism for enhanced platelet-dependent thrombosis in both arterial and venous systems during aging.

Finally, although there was a modest but significant increase in the platelet count in mice at 18 months of age compared with 4 or 12 months of age (Tables 1 and 2), it is unlikely that this difference contributed appreciably to the increased thrombotic susceptibility with aging for 2 reasons: First, the platelet count was significantly elevated only at 18 months of age, whereas increased susceptibility to arterial and venous thrombosis was apparent at both 12 and 18 months of age; and second, a similar elevation of the platelet count was observed in WT and Gpx1 Tg mice at 18 months of age (Table 2), and Gpx1 Tg mice were nevertheless protected from platelet hyperactivation and thrombosis. We previously reported that a similar increase in platelet count was not associated with increased thrombotic susceptibility in mice with hyperhomocysteinemia.\textsuperscript{48} We conclude therefore that the prothrombotic phenotype of aged mice is likely to be mediated by platelet hyperactivity rather than thrombocytosis.

**Conclusions**

We demonstrate here that aged mice develop increased susceptibility to both arterial and venous thrombosis and that H$_2$O$_2$-mediated platelet hyperactivation is a likely mechanism leading to this prothrombotic phenotype. These findings suggest that therapeutic strategies targeted toward lowering platelet H$_2$O$_2$ levels may have the potential to decrease thrombotic complications of aging. One potential strategy is to identify drugs that increase platelet glutathione peroxidase or catalase activity, perhaps by upregulating or allosterically increasing their activity. Another approach might be to develop inhibitors of the platelet NADPH oxidase/SOD1 pathway. Inhibition of SOD1 may prove to be problematic, however, because SOD1 inhibitors might be expected not only to decrease H$_2$O$_2$ but also to increase superoxide, which also may produce platelet hyperactivation. Targeting NADPH oxidase may prove to be a more promising approach because several small-molecule and peptide-based inhibitors of Nox2-containing NADPH oxidases subunits are currently in development.\textsuperscript{49,50}

**Acknowledgments**

We thank Li Wang for her help in measuring platelet count in whole-blood samples. We also thank Kristina W. Thiel for assistance in manuscript preparation and editing.

**Sources of Funding**

This work was supported by American Heart Association Beginning Grant-in-Aid 0860052Z to Dr Dayal, American Heart Association Scientist Development Grant SDG7520015 to Dr Chauhan, and by National Institutes of Health grants HL.063943 and NS024621 (to Dr Lentz) and HL.106495 (to Dr Motto).

**Disclosures**

None.

**References**

8. Oudot A, Martin C, Busseau D, Vergely C, Demaison L, Rochette L. NADPH oxidases are in part responsible for increased...
27. Dayal et al Platelet Activation and Thrombosis in Aging 1315

CLINICAL PERSPECTIVE
Aging is associated with an increased incidence of thrombotic events such as stroke, myocardial infarction, deep vein thrombosis, and pulmonary embolism. However, the mechanisms by which aging increases susceptibility to thrombosis remain elusive. One potential causal link between aging and thrombotic susceptibility may be oxidative stress, which increases with aging and is hypothesized to play a pivotal role in cardiovascular disease. The objective of this study was to use a mouse model to investigate the mechanistic role of oxidative stress in aging-related thrombosis. To do so, we tested the specific hypothesis that aged mice overexpressing an antioxidant enzyme, glutathione peroxidase-1, are protected from experimental thrombosis. Our data demonstrate that susceptibility to both arterial and venous thrombosis is enhanced with aging by a mechanism involving hydrogen peroxide and that peroxide-mediated hyperactivation of platelets in aged mice may be particularly important for this prothrombotic phenotype. These findings suggest that targeted approaches to prevent the accumulation of peroxide in activated platelets may represent an alternative to general antioxidant therapy as a therapeutic strategy to prevent thrombotic complications of aging.
Hydrogen Peroxide Promotes Aging-Related Platelet Hyperactivation and Thrombosis
Sanjana Dayal, Katina M. Wilson, David G. Motto, Francis J. Miller, Jr, Anil K. Chauhan and Steven R. Lentz

_Circulation_. 2013;127:1308-1316; originally published online February 20, 2013; doi: 10.1161/CIRCULATIONAHA.112.000966
_Circulation_ is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2013 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circ.ahajournals.org/content/127/12/1308

Data Supplement (unedited) at:
http://circ.ahajournals.org/content/suppl/2013/02/20/CIRCULATIONAHA.112.000966.DC1

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in _Circulation_ can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to _Circulation_ is online at:
http://circ.ahajournals.org//subscriptions/
SUPPLEMENTAL MATERIAL

Hydrogen peroxide promotes aging-related platelet hyperactivation and thrombosis

Sanjana Dayal, PhD, Katina M. Wilson, BS, David G. Motto, MD, PhD, Francis J. Miller, Jr., MD, Anil K. Chauhan, PhD, and Steven R. Lentz, MD, PhD.
**Supplemental Methods**

**Platelet immunoblotting**

Platelets were isolated from citrated blood and washed in modified Tyrode’s buffer. Platelet lysates were prepared in 1M Tris buffer (pH 7.4) containing 1% triton X-100, 150 mM NaCl, 1 mM EDTA, 1 mM EGTA (pH 8.0), 0.5% NP-40, and protease inhibitor cocktail (Complete™ Mini EDTA-free, Roche, USA). Immunoblotting was performed as described previously.¹ Briefly, 30 µg of protein was run on a 12.5 % Tris-HCl reducing gel (Bio-Rad), and membranes were probed with either 1.0 µg/ml rabbit polyclonal mouse anti-Gpx1 (ab22604, Abcam, Cambridge, MA) or 0.5 µg/ml of mouse monoclonal anti-β-actin as a loading control (ab8226, Abcam, Cambridge, MA). Immunoreactive bands were visualized using Supersignal West femto (Pierce, IL, USA) detection system. Results were quantified with NIH Image J, and expressed as percent of levels observed in 4 month old wild-type (Wt) mice. Two-way ANOVA was used for comparison between the groups.

**Platelet adhesion during thrombus formation in vivo**

The relative adhesion of isolated platelets from donor mice (4 vs. 18 month old Wt mice, or 18 month old Wt vs. 18 month old Gpx1 Tg mice) was assessed during ferric chloride-induced thrombosis formation in mesenteric arterioles of 4 week old Wt recipient mice. Blood was collected from the retro-orbital venous plexus of donor mice into a polypropylene tube containing 0.3 mg/ml enoxaparin. Platelets were isolated, washed, and differentially fluorescently labeled with acetomethoxy (AM) derivative of calcein green or calcein red-orange (Invitrogen) as previously described.² The host mouse was anesthetized with avertin, and differentially labeled platelets (2.5 x 10⁹ platelets/kg) were infused retro-orbitally. Mesenteric
arterioles (between 80-100 µm in diameter) of the host mouse were topically injured with 5% FeCl₃. Real-time imaging was performed using intravital microscopy to record the number of fluorescent platelets adhering to the injured vessel wall during the initial 3 minutes of thrombus formation. One to three mesenteric arterioles were studied in each recipient mouse. At the completion of each experiment, blood was collected and the percentage of circulating fluorescently labeled platelets was determined by flow cytometry. Relative platelet adhesion was defined as the number of adherent fluorescent platelets, normalized to the percentage of circulating fluorescent platelets, and presented relative to either 4 month old Wt mice or 18 month old Gpx1 Tg mice. One-way analysis of variance (ANOVA) was performed on log-transformed values to compare the relative adhesion. Statistical significance was defined as a P value <0.05. Values are reported as mean±SE.
Supplemental Figure I. Platelet adhesion during thrombus formation is increased with aged platelets. Differentially fluorescently labeled platelets were infused into 4 week old recipient C57Bl6/J mice, mesenteric arterioles were injured with 5% FeCl$_3$ and relative platelet adhesion during initial thrombus formation was assessed by intravital microscopy. (A) Relative adhesion of platelets from 4 month old vs. 18 month old Wt mice. (B) Relative adhesion of platelets from 18 month old Gpx1 Tg vs. 18 month old Wt mice. 12-20 vessels were examined in 5-8 host mice per group. Values are mean ± SE. *p<0.05 vs. 4 month old C57Bl6/J mice by one-way ANOVA. (C) Representative image of adhesion of platelets from 4 month old (calcein red-orange AM-labeled; red arrow) vs. 18 month old (calcein green AM-labeled; green arrow) Wt
mice. Also see Supplemental Video S1. (D) Representative image of adhesion of platelets from 18 month old Gpx1 Tg (calcein red-orange AM-labeled; red arrow) vs. 18 month old Wt (calcein green AM-labeled; green arrow) mice. Also see Supplemental Video S2.
Supplemental Figure II. Gpx1 Tg mice have higher Gpx1 mRNA and protein levels in platelets. Gpx1 mRNA and protein levels were measured in platelets from 4 or 18 month old wild-type (Wt) or Gpx1 Tg (Tg) mice by real time PCR and Western blotting. (A) Platelet mRNA levels of Gpx1 expressed as the percent of Gpx1 levels observed in 4 month old Wt mice. Platelets from 9-15 mice were studied in each group. (B) Representative immunoblot of platelet Gpx1 protein levels. (C) Summary of quantitation of 5-9 immunoblotting experiments, with Gpx1 protein levels normalized to β-actin and expressed as percent of levels observed in 4 month old Wt mice. *P<0.05 compared with age-matched Wt littermates by two-way ANOVA.
Supplemental Figure III. Expression of P-selectin on platelet surface is not influenced by aging or Gpx1 genotype. Expression of P-selectin was examined in thrombin-activated platelets from either wild-type (Wt) or Gpx1 Tg (Tg) mice at 4 or 18 months of age using flow cytometry. Platelets from 6-8 mice were studied in each group. Values are mean ± SE.
Supplemental References


Video legends

**Supplemental Video S1**: Relative adhesion of platelets from 4 and 18 month old C57Bl6/J mice. Representative video of adhesion of platelets from 4 month old (calcein red-orange AM-labeled) vs. 18 month old (calcein green AM-labeled) C57Bl6/J mice. Differentially fluorescently labeled platelets from 4 and 18 months old C57Bl6/J mice were infused into 4 week old recipient C57Bl6/J mice, mesenteric arterioles were injured with 5% FeCl$_3$ and relative platelet adhesion during initial thrombus formation was assessed by intravital microscopy.

**Supplemental Video 2**: Relative adhesion of platelets from 18 month old wild-type (Wt) and Gpx1 Tg (Tg) mice. Representative video of adhesion of platelets from 18 month old Tg (calcein red-orange AM-labeled) vs. Wt (calcein green AM-labeled) mice. Differentially fluorescently labeled platelets from 18 months old Wt or Tg mice were infused into 4 week old
recipient C57Bl6/J mice, mesenteric arterioles were injured with 5% FeCl₃ and relative platelet adhesion during initial thrombus formation was assessed by intravital microscopy.