A Quantitative Trait Locus (LSq-1) on Mouse Chromosome 7 Is Linked to the Absence of Tissue Loss After Surgical Hindlimb Ischemia

Ayotunde O. Dokun, MD, PhD*; Sehoon Keum, MS*; Surovi Hazarika, MD, PhD; Yongjun Li, MD; Gregory M. Lamonte, MS; Ferrin Wheeler, PhD; Douglas A. Marchuk, PhD†; Brian H. Annex, MD†

**Background**—Peripheral arterial disease (PAD) caused by occlusive atherosclerosis of the lower extremity has 2 major clinical manifestations. Critical limb ischemia is characterized by rest pain and/or tissue loss and has a ≥40% risk of death and major amputation. Intermittent claudication causes pain on walking, has no tissue loss, and has amputation plus mortality rates of 2% to 4% per year. Progression from claudication to limb ischemia is infrequent. Risk factors in most PAD patients overlap. Thus, we hypothesized that genetic variations may be linked to presence or absence of tissue loss in PAD.

**Methods and Results**—Hindlimb ischemia (murine model of PAD) was induced in C57BL/6, BALB/c, C57BL/6BALB/c (F1), F1/BALB/c (N2), A/J, and C57BL/6J-Chr7A/J/Nal chromosome substitution strains. Mice were monitored for perfusion recovery and tissue necrosis. Genome-wide scanning with polymorphic markers across the 19 murine autosomes was performed on the N2 mice. Greater tissue loss and poorer perfusion recovery occurred in BALB/c than in the C57BL/6 strain. Analysis of 105 N2 progeny identified a single quantitative trait locus on chromosome 7 that exhibited significant linkage to both tissue necrosis and extent of perfusion recovery. Using the appropriate chromosome substitution strain, we demonstrate that C57BL/6-derived chromosome 7 is required for tissue preservation.

**Conclusions**—We have identified a quantitative trait locus on murine chromosome 7 (LSq-1) that is associated with the absence of tissue loss in a preclinical model of PAD and may be useful in identifying gene(s) that influence PAD in humans. *(Circulation. 2008;117:1207-1215.)*

**Key Words:** atherosclerosis ■ genetics ■ peripheral vascular disease

Peripheral arterial disease (PAD) is caused by atherosclerosis in arterial beds other than the coronary arteries. The lower extremity is the most common site for PAD, where atherosclerotic occlusive disease impairs perfusion to the leg(s). The 2 major clinical manifestations of lower-extremity PAD are (1) intermittent claudication (IC), which is defined as leg pain during exercise that is relieved by rest and the absence of either rest pain or tissue loss, and (2) critical limb ischemia (CLI), which is defined as leg pain at rest with or without tissue loss (nonhealing leg ulcers or gangrene). Patients with IC infrequently progress to CLI and have a combined annual amputation and mortality rate of 2% to 4% per patient per year, whereas patients with CLI have a 6-month amputation risk of 25% to 40% and an annual mortality rate as high as 20%.1–3
morphism of a gene (or genes) can be associated with an increased risk for the development of lower-extremity atherosclerosis, with or without involvement of other vascular beds; examples of this include apolipoprotein E, interleukin-6, and thrombin. Second, in the setting of occlusive atherosclerosis, a polymorphism in a gene (or genes) can influence the disease severity, such as the presence or absence of tissue loss. To date, no information is available on the latter.

We took advantage of prior findings in a preclinical model of PAD in which, after surgically induced hindlimb ischemia, the C57BL/6 strain displayed good blood flow recovery with little or no tissue loss, whereas the BALB/c strain showed poor recovery and was prone to develop tissue loss or necrosis of the affected limb. This contrast suggests that poor recovery and was prone to develop tissue loss or absence of tissue loss. To date, no information is available on the latter.

We took advantage of prior findings in a preclinical model of PAD in which, after surgically induced hindlimb ischemia, the C57BL/6 strain displayed good blood flow recovery with little or no tissue loss, whereas the BALB/c strain showed poor recovery and was prone to develop tissue loss or necrosis of the affected limb.13,14 This contrast suggests that genes (or genes) can influence the disease severity, such as the presence or absence of tissue loss. To date, no information is available on the latter.

We took advantage of prior findings in a preclinical model of PAD in which, after surgically induced hindlimb ischemia, the C57BL/6 strain displayed good blood flow recovery with little or no tissue loss, whereas the BALB/c strain showed poor recovery and was prone to develop tissue loss or necrosis of the affected limb.13,14 This contrast suggests that sequence variation in specific gene(s) expressed in C57BL/6 and BALB/c strains may confer the observed phenotypic difference. Therefore, we sought to map the genetic loci involved in this differential recovery and test the role of the identified chromosomal region in the observed phenotypic outcome.

**Methods**

**Mice**

All mice (C57BL/6, BALB/c, A/J, C57BL/6J-Chr7^NaJ/NaJ) were obtained from the Jackson Laboratory (Bar Harbor, Me) either directly or bred internally (C57BL/6×BALB/c [F1] and F1×BALB/c [N2]) from a parental strain. N2 animals were derived from 1-way mating with F1 males crossed to BALB/c females. The C57BL/6J-Chr7^NaJ/NaJ (chromosome substitution strains [CSS]) strain is genetically identical to C57BL/6 except at chromosome 7, which is derived from the A/J strain. Mice were allowed to acclimate for a minimum of 3 days. Mice were age matched (12 to 18 weeks old for hindlimb ischemia and 8 to 12 weeks old for wound healing) and sex matched for all experiments. Studies were performed under protocols approved by the Duke University Animal Care and Use Committee.

**Hindlimb Ischemia, Perfusion Recovery, Necrosis Score, and Histology**

Unilateral femoral artery ligation and excision, resulting in hindlimb ischemia, was performed as described previously. Perfusion flow in the ischemic and contralateral nonischemic limbs was measured as described previously with the use of a laser Doppler perfusion imaging system (Perimed, Stockholm, Sweden). Perfusion was expressed as the ratio of the left (ischemic) to right (nonischemic) hindlimb and was performed immediately after surgery and weekly or biweekly up to 21 or 35 days postoperatively. Perfusion recovery was determined either by the absolute perfusion ratio at follow-up or as the ratio at follow-up minus the ratio immediately after surgery. In mice that developed autoamputation, the perfusion ratio obtained from the limb before autoamputation was used. The extent of necrosis was scored as follows: grade I, involving only toes; grade II, extending to dorsum pedis; grade III, extending to crus; and grade IV, extending to thigh or complete necrosis. In the ischemic and nonischemic gastrocnemius muscles, vascular density was analyzed by immunohistochemistry with a rat anti-mouse CD31 antibody by counting 3 random high-power (magnification ×200) fields for a minimum of 200 fibers and was expressed as the number of CD31^+ cells per fiber, as described previously.

**Wound Healing**

Full-thickness skin punch biopsies were made, as described previously by others. Briefly, mice were anesthetized with ketamine (50 mg/kg) and xylazine (5 mg/kg). The skin over the dorsum surface was shaved, and a 0.7- to 1.0-cm-diameter wound was generated with a disposable 0.8-cm-diameter skin punch biopsy tool (Acuderm Inc, Fort Lauderdale, Fla). Wound dressing was achieved with the use of a spray-on liquid bandage solution (Nexcare, St Paul, Minn). Wound area was measured weekly. The healing rate was defined as a percentage of original wound area or totally healed.

**Genotyping**

A standard high-salt procedure was used to isolate genomic DNA from mouse tails. A total of 105 N2 animals were used to identify quantitative trait locus (QTL) in the linkage scan. Single-nucleotide polymorphism (SNP) genotyping was performed with the use of the GoldenGate genotyping assay and the Illumina Bead Station (San Diego, Calif). The Illumina mouse low-density linkage panel was used, consisting of 377 SNPs covering the entire murine genome, chosen to be maximally informative in crosses between C57BL/6 and other inbred strains. In our cross between C57BL/6 and BALB/c, 239 autosomal SNP markers were informative. The average spacing per informative SNP was 6.8±1.6 cM, with the largest gap located on chromosome 2 (24.6 cM). Reported genetic map positions for the markers were retrieved from the SNP database (build 36.1) of the National Center for Biotechnology Information (NCBI).

**Linkage Analysis**

The MapManager QTX program (version b20) was used to localize the QTL responsible for the variability of recovery phenotypes from surgically induced hindlimb ischemia (necrosis and perfusion ratio). The source codes are available online at http://www.mapmanager.org/mmQTX.html. The significance thresholds for each phenotypic trait were determined empirically by permutation tests performed with all the informative markers and 10,000 permutation tests of data. This logarithmic index can be converted to a logarithm of odds (LOD) score by dividing it by 4.61. Suggestive (P=0.07), significant (P=0.05), and highly significant (P=0.001) thresholds were established on the basis of the guidelines suggested by Lander and Kruglyak. Single-locus association tests were performed between each marker and the phenotype to identify regions of interest in our initial genome scan. The genome-wide scans were plotted with the use of the J/QTML mapping program (version 0.8) obtained from the Jackson Laboratory.

In Silico Haplotype Analysis

To assess the possibility that QTLs can be identified only in genomic regions that differ between 3 strains of interest (C57BL/6, BALB/c, and A/J), SNP data were used to construct the haplotype map of the 31.2-Mb interval of mouse chromosome 7. SNP data were obtained from the Mouse Phenome Database (http://phenome.jax.org/pub-cgi/phenome/mpdcgi?rtn=snp/door). Physical map position for markers was based on the mouse genomic sequence from the NCBI build 36.1. Only SNPs polymorphic between the 3 strains were used to refine the haplotype map. SNPs missing data on 1 or 2 strains as well as SNPs that were not polymorphic between the 3 strains were omitted from the haplotype map because of lack of information.

**Statistical Analysis**

All continuous measures were expressed as mean±SEM. Statistical comparisons of perfusion or capillary density between multiple strains of mice were performed with the use of 1-way ANOVA; between 2 strains, the independent Student t test was used. Comparison of perfusion within each strain at different time points was analyzed by repeated-measures ANOVA. Differences in necrosis score between mice with a different genetic background were analyzed by nonparametric Kruskal-Wallis test. In all cases, P<0.05 was considered statistically significant. Correlation of perfusion recovery with necrosis was performed with the use of the Spearman correlation.

The authors had full access to and take full responsibility for the integrity of the data. All authors have read and agree to the manuscript as written.
Results

Early Identification of a QTL

C57BL/6 mice showed robust perfusion recovery after ligation, whereas far less recovery was shown by the BALB/c mice (Figure 1A). Necrosis was quite rare in the C57BL/6 strain and when present was at a low grade, whereas BALB/c mice were prone to develop necrosis (Figure 1B and the Table). The F1 mice exhibited no significant difference in perfusion recovery or necrosis from the C57BL/6 parental strain (Figure 2). This suggested that a dominant allele(s) in C57BL/6 could be responsible for the lack of tissue loss.

To map the allele in C57BL/6 mice, we performed an F1 backcross with BALB/c to generate 105 N2 progeny. A total of 105 N2 progeny were genotyped for the 239 SNPs that were informative in this cross. The average spacing between SNPs was \(6.8 \pm 1.6\) cM, affording full coverage of the mouse genome. Figure 3 illustrates the interval maps of each of the autosomes for the phenotypes of necrosis and perfusion ratio at day 21 after surgery. All but 1 of the chromosomes showed linkage profiles that were essentially flat, providing evidence for the exclusion of any strongly acting loci mapping to these chromosomes. However, we identified a single locus on

<table>
<thead>
<tr>
<th>Mouse Strain</th>
<th>n</th>
<th>Mice With Any Necrosis</th>
<th>Necrosis Grade</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Grade 0</td>
</tr>
<tr>
<td>BALB/c</td>
<td>11</td>
<td>10/11 (90.1)</td>
<td>1/11 (9)</td>
</tr>
<tr>
<td>C57BL/6(B/6)</td>
<td>41</td>
<td>1/41 (2.4)</td>
<td>40/41 (97.6)</td>
</tr>
<tr>
<td>B/6×BALB/cF1</td>
<td>18</td>
<td>3/18 (16.7)</td>
<td>15/18 (83)</td>
</tr>
<tr>
<td>A/J</td>
<td>15</td>
<td>9/15 (60)</td>
<td>6/15 (40)</td>
</tr>
<tr>
<td>CSS</td>
<td>10</td>
<td>8/10 (80)</td>
<td>2/10 (20)</td>
</tr>
</tbody>
</table>

Necrosis grade is from 0 through 4, as described in Methods. Values are expressed as number of mice affected in each category/total number of mice in the category (%). Distribution is to the right.
chromosome 7 spanning ∼31 Mb that exhibited significant linkage to tissue necrosis and day 21 perfusion ratios, with a peak LOD score of 7.96 and 3.71, respectively, at marker rs13479513 (Figure 4). We showed the whole genome linkage profile for reperfusion ratio at day 21 because at this time point, chromosome 7 exhibited the highest LOD score. Perfusion ratio was also measured at days 7 and 14 after surgery. For both of these time points, the chromosome 7 linkage reached the P=0.05 significance threshold (data not shown). Otherwise, across the genome, the linkage profiles for other time points are similar to that seen for the day 21 measurement. Furthermore, the linkage peak for necrosis was even more significant, far surpassing the P<0.001 threshold. The linkage peaks for perfusion ratio (Figure 3B) and necrosis (Figure 3A) were almost identical in chromosomal position, differing only in the magnitude of the strength of linkage. Because the LOD score for the necrosis trait greatly exceeded that of the perfusion ratio trait, we examined the possibility that the linkage score for the necrosis trait was artificially inflated because of a nonnormal distribution of necrosis scores between heterozygous (C57BL/6×BALB) and homozygous (BALB×BALB) animals at the chromosome 7 locus. To exclude this possibility, we recoded the necrosis scores of all the N2 mice on a scale of 0, I, or II (where all scores >II received a value of II) and again as a dichotomous trait (where any level of necrosis received a score of I). Under both recoding schemes, the linkage scores for necrosis were essentially unchanged from the original (data not shown). Because the MapManager QTX program employs means and not variance components in the analysis, the linkage scores are not unduly influenced by the nonnormal distribution of the values for the trait. Instead, it is likely that these 2 phenotypes are biologically interrelated, with necrosis being the stronger measure of the underlying pathogenesis. The coincident linkage peaks for both traits further support this possibility. No significant gender differences were present in either tissue necrosis or perfusion ratio in either parental background (data not shown) or in the progeny of the backcross.
Genotype–Phenotype Correlations

To determine the allelic contribution to the effect of the locus between genotype and phenotype data, necrosis and perfusion ratio were evaluated against genotype at the chromosome 7 locus that showed the highest LOD score among all of our SNP markers. The N2 animals with a heterozygous genotype (C57BL/6×BALB) at marker rs13479513 showed minimal necrosis and higher perfusion ratio on average than their homozygous (BALB×BALB) littermates (0.37 versus 1.60 and 0.71 versus 0.47, respectively; Figure 5A and 5B). Because the necrosis score was not normally distributed, we used the nonparametric Kruskal-Wallis test, and both necrosis score and perfusion ratio were significantly different between BALB×BALB and C57BL/6×BALB animals (P<0.0001). Moreover, when we examined the correlation between necrosis and perfusion recovery in the N2 progeny, an association was present between poor final perfusion and greater necrosis in these mice (r=-0.628, P<0.001; Figure 5C). Results were similar if the perfusion recovery was measured as perfusion at follow-up minus the immediate postoperative value. Taken together, these data support the findings that a region of the C57BL/6 strain is contributing a dominant protective allele (or alleles) to the recovery phenotype from hindlimb ischemia, and this allele is protective via a mechanism contributing to perfusion recovery.

Requirement of QTL on C57BL/6 Chromosome 7 for Absence of Tissue Loss

We sought to formally test whether the C57BL/6 chromosome 7 was required for the absence of tissue loss. We took advantage of the CSS strain C57BL/6J-Chr7a/aNaJ in which the C57BL/6 chromosome 7 has been replaced by chromosome 7 from the A/J strain. To determine whether this strain will be useful in testing our hypothesis, it was important to first determine the recovery phenotype of the parental A/J strain in surgically induced hindlimb ischemia. If the A/J strain shows poor recovery, then the CSS can be used to evaluate the role of C57BL/6 chromosome 7. Hence, we performed unilateral femoral ligation and excision on A/J mice to determine its recovery phenotype; indeed, the A/J strain showed poor recovery compared with C57BL/6 (Figure 6A) and, similar to BALB/c, was prone to developing limb necrosis (Table). In light of these findings, we proceeded to test the phenotype in the CSS strain and discovered that this strain also showed poor recovery and tendency to develop limb necrosis (Table). Furthermore, although no significant difference was observed in capillary density between C57BL/6 (0.99±0.07) and A/J (0.91±0.04; P=0.38) strains in nonischemic tissues, 5 weeks after induction of limb ischemia, C57BL/6 mice showed higher capillary density (1.30±0.06) than the A/J or the CSS strain (0.97±0.10, P=0.03 or 1.01±0.11, P=0.04, n=3 per group). Therefore, these data show that the C57BL/6 chromosome 7 is required for the lack of tissue loss and greater perfusion recovery phenotype after hindlimb ischemia.

C57BL/6 Chromosome 7 QTL Has No Effect on Wound Healing

Next, we sought to determine the role of this C57BL/6-derived chromosome 7 locus in a nonischemic tissue injury (ie, wound healing). Wounding healing rates were determined over a 2-week period. No significant difference was observed between the 2 strains in the rate of wound healing 1 week after injury (79.8±2.6% in BALB/c versus 83.9±3.2% in C57BL/6; P=0.29; n=6 per group), and all mice from both strains achieved complete wound closure by week 2 (100% in both strains). Similar results were observed in the CSS strain (90.4±4.1% at 1 week; n=4; all mice were healed at 2
Polymorphism. Approximately 97% of the genetic variation within a QTL interval that are likely to harbor the causal gene(s) can be identified by quickly identifying high-priority regions in the inbred mouse lineage. Haplotype analysis can decrease confidence intervals by taking advantage of ancestral haplotype blocks that are shared between strains, with only a minority of the sequence variation being unique to any 1 strain. There-fore, we hypothesize that the allelic variation at the chromosome 7 recovery locus is most likely due to a gene that maps within ancestral haplotype blocks that are shared between BALB/c and A/J but differ from C57BL/6 strains. These regions are most likely to contain the causative gene for the differential recovery after ischemic tissue injury.

Using a dense microsatellite and SNP map of the broad chromosome 7 locus, we were able to identify the haplotype blocks that consisted of the same SNP alleles between BALB/c and A/J but were different from C57BL/6. As shown in Figure 7, 2 of the haplotype blocks in which the C57BL/6 allele differed from the shared BALB/c and A/J allele lie directly under the central region of the linkage peak (2-LOD support interval). Of primary interest are the 2 haplotype blocks of ≈1.7 and 1.9 Mb, which contain 21 and 16 known or predicted genes, respectively (Figure 7).

**Discussion**

The results of our study demonstrate, for the first time, that inherited genetic variations strongly influence the phenotypic outcome after femoral arterial occlusion, which is a preclinical model of PAD. We took advantage of known strain-specific differences in recovery after hindlimb ischemia. First, using classic genetic tools, we demonstrated that the genetic background of C57BL/6 mice was associated with greater limb perfusion and less limb loss after hindlimb ischemia compared with BALB/c. We then used 2 complementary approaches to demonstrate the pivotal role of C57BL/6-derived genetic sequence in conferring limb preservation (when present) or tissue loss (when absent). We identified an initial genetic locus covering an area of ≈19.8 Mb (Figure 4; highly significant) to 31.2 Mb (Figure 4; significant) containing 274 to 495 genes, respectively. Then, by taking advantage of ancestral haplotype blocks that are similar in strains with poor recovery but different in a strain with good recovery, we were able to quickly refine this genetic interval to 2 regions containing a total of 37 genes.

Aside from the information contained in our present report, little to no information is available on genetic influences in PAD. The overall approach used in our study has successfully led to the identification of genes associated with human disease. Paigen et al took advantage of differences in mouse strain susceptibility to the development of atherosclerosis to describe a region on chromosome 1, termed the 

**Haplotype Analysis: Toward Gene Identification**

We employed ancestral haplotype sharing pattern within the inbred mouse lineage. Haplotype analysis can decrease confidence intervals by quickly identifying high-priority regions within a QTL interval that are likely to harbor the causal polymorphism. Approximately 97% of the genetic variation between mouse inbred strains is found in ancestral haplotype blocks that are shared between strains, with only a minority of the sequence variation being unique to any 1 strain. Therefore, we hypothesize that the allelic variation at the chromosome 7 recovery locus is most likely due to a gene that maps within ancestral haplotype blocks that are shared between BALB/c and A/J but differ from C57BL/6 strains. These regions are most likely to contain the causative gene for the differential recovery after ischemic tissue injury.
is not surprising that associations were found between PAD and polymorphisms in genes such as apolipoprotein-B and thrombin. However, outside of genes related to atherosclerosis or thrombosis risk, genetic association studies for PAD are quite limited. We are aware of only 1 study, a family-based linkage study that identified a genetic locus conferring susceptibility to PAD. Gudmundsson et al identified a locus mapping to human chromosome 1p31 termed PAOD1. One of the strengths of this study is that it took an unbiased approach, starting with the PAD phenotype, to identify novel genetic risk factors rather than starting with a potentially biased list of candidate genes. Using a similarly unbiased genetic mapping approach, we have identified a genetic locus conferring limb preservation after murine hindlimb ischemia. Our locus (LSq-1) is not syntenic to human chromosome 1p31 but rather shows synteny to regions of human chromosomes 10, 11, and 16. It is therefore highly unlikely that PAOD1 encodes the same gene (or genes) as LSq-1, although it is plausible that genes in both species may ultimately be shown to play a role in the same physiological pathway.

Recently, Faber and colleagues proposed that differences in expression of vascular endothelial growth factor-A (VEGF-A) may account for the differences in collateral vessel density observed between C57BL/6 and BALB/c and further suggested that this anatomic difference may underlie the different recovery phenotypes after surgically induced hindlimb ischemia. They determined that these 2 strains do not show differential expression of VEGF-A in skeletal muscle at baseline (nonischemic muscle), but expression levels differ as a result of the PAD phenotype. Using QTL analysis, they also determined that the cause of this differential expression maps to murine chromosome 17, and we did not find any evidence of a dominantly acting C57BL/6 allele on this chromosome in our cross (Figure 3A and 3B). Thus, we have not genetically linked the VEGF-A gene locus.

### Table 1

<table>
<thead>
<tr>
<th>Position (bp)</th>
<th>Gene Symbol ID</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>12783216</td>
<td>LOCC66875</td>
<td>itergrin alpha M</td>
</tr>
<tr>
<td>127852379</td>
<td>Itgam</td>
<td>integrin X</td>
</tr>
<tr>
<td>127918726</td>
<td>Itgax</td>
<td>integrin alpha D</td>
</tr>
<tr>
<td>127963443</td>
<td>Itgad</td>
<td>cytochrome c oxidase subunit VI a, polypeptide 2</td>
</tr>
<tr>
<td>128026980</td>
<td>Armc5</td>
<td>armadillo c repeat containing 5</td>
</tr>
<tr>
<td>128027554</td>
<td>9130023H24Rik</td>
<td>RIKEN cDNA 9130023H24 gene</td>
</tr>
<tr>
<td>128036360</td>
<td>Tgbill</td>
<td>transforming growth factor beta 1 induced transcript 1</td>
</tr>
<tr>
<td>128055419</td>
<td>Slc5a2</td>
<td>solute carrier family 5 (sodium/glucose co-transporter) member 2</td>
</tr>
<tr>
<td>128062043</td>
<td>BC017158</td>
<td>cDNA sequence BC017158</td>
</tr>
<tr>
<td>128129969</td>
<td>LOC628781</td>
<td>regulator of G protein signaling 10</td>
</tr>
<tr>
<td>128164773</td>
<td>Rgs10</td>
<td>Tila 1 cytotoxic granule-associated RNA binding protein-like 1</td>
</tr>
<tr>
<td>128233450</td>
<td>Tial1</td>
<td>Tila 1 cytotoxic granule-associated RNA binding protein-like 1</td>
</tr>
<tr>
<td>128314383</td>
<td>Bag3</td>
<td>Bcl2-associated anagathogene 3</td>
</tr>
<tr>
<td>128314383</td>
<td>LOC639030</td>
<td>regulator of G protein signaling 10</td>
</tr>
<tr>
<td>128402198</td>
<td>Inpp5f</td>
<td>inositol polyphosphate-5-phosphatase F</td>
</tr>
<tr>
<td>128487163</td>
<td>1110007A13Rik</td>
<td>RIKEN cDNA 1110007A13 gene</td>
</tr>
<tr>
<td>128534855</td>
<td>Sec23ip</td>
<td>Sec23 interacting protein</td>
</tr>
<tr>
<td>128788002</td>
<td>LOC210714</td>
<td>phosphatidic acid phosphatase type 2 domain containing 1A</td>
</tr>
<tr>
<td>129046580</td>
<td>Ppapdc1a</td>
<td>phosphatidic acid phosphatase type 2 domain containing 1A</td>
</tr>
<tr>
<td>129381624</td>
<td>Brwd2</td>
<td>bromodomain and WD repeat domain containing 2</td>
</tr>
</tbody>
</table>

![Figure 7](https://example.com/figure7.png)
The surgical hindlimb ischemia model is complex, and multiple mechanisms are involved in recovery, including but not limited to ischemic injury, tissue regeneration, inflammation, angiogenesis, and arteriogenesis. Although our data demonstrate that genetic variations influence perfusion recovery, the specific gene(s) and mechanism(s) by which LSq-1 contributes to strain specific recovery in this preclinical model are beyond the scope of a single study. We found that LSq-1 did not affect nonischemic wound healing, and we showed a correlation between lower perfusion recovery and greater necrosis, suggesting that the locus effect may be via a mechanism contributing to restoration of perfusion. This suggests that C57BL/6-derived genetic sequences may be important in the response to ischemia rather than for generalized tissue regeneration.

Classic studies by Schaper and colleagues showed that differences in the extent of preexisting collateral vessels and arteriogenesis play a role in perfusion recovery after hindlimb ischemia. Studies by Fukino et al. and other groups found a differential angiogenic response in the C57BL/6 and BALB/c strains after limb ischemia, suggesting a role for arteriogenesis in this phenotypic outcome. Our data showing higher vascular density in C57BL/6 tissue at late time points after ligation compared with the CSS and A/J strains are also consistent with a role for arteriogenesis in strain-specific recovery. Our data do not exclude the possibility that differences in preexisting collateral vessels arteriogenesis, development of new collateral vessels via angiogenesis, adaptation to tissue ischemia, or some combination of these factors contribute to differential recovery after ischemia or that these genetic differences could work through mechanisms not yet postulated. At present, the mechanism(s) by which LSq-1 contributes to strain-dependent recovery after hindlimb ischemia is not understood completely. However, future studies that identify the specific gene(s) within the LSq-1 locus that mediate the beneficial effect, as well as their human orthologs, will likely lead to a better understanding of the pathogenesis of PAD and may also provide insight into novel therapeutic options.

Sources of Funding
Dr Dokun is a Postdoctoral Fellow supported by grant T32 HL007101 from the National Institutes of Health. Dr Annex was supported in part by grant R01 HL755752 from the National Institutes of Health.

Disclosures
None.

References


CLINICAL PERSPECTIVE

When occlusions are present in the large arteries to the lower extremity, the result is the most common form of peripheral arterial disease (PAD). PAD, a major healthcare problem, has 2 major clinical presentations: intermittent claudication and critical limb ischemia. Despite the significant overlap in risk factors leading to the development of both clinical presentations, patients with critical limb ischemia frequently have ongoing or imminent tissue loss and thus have a higher rate of amputation and mortality. Patients with intermittent claudication rarely develop tissue loss. Therefore, we hypothesized that underlying genetic differences may influence the clinical outcomes in PAD. In the present study, we used a preclinical model of PAD and identified a chromosomal region in mice, termed LSq-1, that, when present, was associated with absence of tissue loss. This is the first identification of such a quantitative trait locus in PAD. This study can serve as a valuable foundation for future studies to identify specific gene(s) involved in PAD in patients. We describe how these results can be used to rapidly identify human orthologs of the mouse genes. Such information has the potential to advance our understanding of the pathophysiology of PAD, as well as new treatment options.
A Quantitative Trait Locus (LSq-1) on Mouse Chromosome 7 Is Linked to the Absence of Tissue Loss After Surgical Hindlimb Ischemia
Ayotunde O. Dokun, Sehoon Keum, Surovi Hazarika, Yongjun Li, Gregory M. Lamonte, Ferrin Wheeler, Douglas A. Marchuk and Brian H. Annex

Circulation. 2008;117:1207-1215; originally published online February 19, 2008;
doi: 10.1161/CIRCULATIONAHA.107.736447
Circulation is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2008 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circ.ahajournals.org/content/117/9/1207

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Circulation can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Circulation is online at:
http://circ.ahajournals.org//subscriptions/