Signal Transducer and Activator of Transcription 1 Is Required for Optimal Foam Cell Formation and Atherosclerotic Lesion Development

Sudesh Agrawal, PhD; Maria Febbraio, PhD; Eugene Podrez, MD, PhD; Martha K. Cathcart, PhD; George R. Stark, PhD; Guy M. Chisolm, PhD

Background—Signal transducer and activator of transcription 1 (Stat1) potently regulates gene expression after stimulation by certain cytokines involved in tumorigenesis and host defenses. The present study investigated a novel role for Stat1 in foam cell formation and atherosclerosis.

Methods and Results—Inhibition of Stat1 activity by a Stat1-specific DNA “decoy” oligomer transfected into differentiated human THP-1 cells, and deficiency of stat1 in mouse macrophages significantly inhibited foam cell formation assessed by lipid staining and cholesteryl ester accumulation compared with control cells. The mechanism of Stat1 regulation of foam cell formation was uniquely dependent on the scavenger receptor CD36. Blunted Stat1 activity and stat1 deficiency significantly decreased expression of CD36 but not of scavenger receptor-A compared with controls, as assessed by immunoblotting and flow cytometry. Deficiency of CD36 but not scavenger receptor-A in mouse macrophages removed any dependency of foam cell formation on Stat1. In an intraperitoneal model of foam cell formation in which foam cells form in vivo independently of the model ligands used in vitro, stat1 deficiency significantly inhibited foam cell formation and CD36 expression. Transplantation of bone marrow from apolipoprotein e⁻/⁻×stat1⁻/⁻ mice into lethally irradiated, atherosclerosis-susceptible apolipoprotein e⁻/⁻ recipients significantly reduced both en face aortic lesion coverage and aortic root lesions compared with recipients of bone marrow from genetically matched apolipoprotein e⁻/⁻ mice.

Conclusions—Stat1 regulations CD36 expression and foam cell formation in macrophages in vitro; the Stat1 regulation of foam cell formation requires CD36. The regulation of CD36 expression by Stat1 may be important in other pathophysiological CD36-dependent events. Stat1 deficiency reduces atherosclerosis in an apolipoprotein e⁻/⁻ atherosclerosis-susceptible bone marrow transplantation model. (Circulation. 2007;115:2939-2947.)

Key Words: atherosclerosis ■ CD36 antigens ■ foam cells ■ lipoproteins ■ macrophage ■ signal transduction ■ STAT1 transcription factor

The janus kinase-signal transducers and activators of transcription (Stat) signaling pathway regulates certain biological events in development, differentiation, apoptosis, proliferation, and immune responses. Stat1 is a transcription factor that is a downstream target of interferon (IFN)-α and -γ and certain other cytokines. Binding of IFN-γ to its receptor evokes a cascade of events that involves phosphorylation of the receptor and associated janus kinase proteins. The activated receptor–janus kinase complex serves as a docking site for Stat1, which leads to its phosphorylation at tyrosine-701. Homodimers of activated Stat1 can translocate to the nucleus where they bind consensus DNA γ-activated sites, which leads to activation or repression of the transcription of numerous genes. Although IFN-γ has been shown to be proatherogenic, a role for Stat1 in atherosclerosis has not been previously reported.

Clinical Perspective p 2947

In early fatty streak lesions of human and mouse atherosclerosis, lipoprotein accumulation and oxidation occur. This is followed by activation of the endothelium, recruitment of circulating monocytes and their adhesion to the endothelium, diapedesis into the intima, and differentiation to macrophages. Macrophage uptake of oxidized lipoproteins via scavenger receptors such as CD36 and scavenger receptor-A (SR-A) is hypothesized to be an early event in atherosclerotic lesion formation, which leads to the formation of lipid-engorged “foam cells.” The studies herein tested the hypothesis that Stat1 is proatherogenic. The role of Stat1 was studied with in vitro and in vivo foam cell formation model systems and an
atherosclerosis-susceptible mouse model. The results revealed that Stat1 positively influences lesion formation in vivo and is required for optimal progression of foam cell formation in vitro and in vivo. In addition, Stat1 regulation of foam cell formation in vitro was mediated by CD36. These roles of Stat1 may be important not only for the understanding of atherosclerosis, but also of other CD36-dependent biological events.

Methods

THP-1 Cell Differentiation and Foam Cell Formation
THP-1 cells obtained from American Type Culture Collection (ATCC, Manassas, Va.) were differentiated by stimulation with 15 nM phorbol 12-myristate 13-acetate (PMA) for 2 hours and then washed with PBS and cultured in RPMI-1640 with 10% fetal bovine serum for multiple days. Foam cell formation was induced by incubation of differentiated THP-1 cells (grown on glass coverslips) for 48 hours with cupric ion-oxidized low-density lipoprotein (LDL) (Cu-oxLDL; 50 μg/mL) or other forms of modified LDL. Cells were stained with Oil Red O as described. Photographs of foam cells were taken with a phase-contrast microscope, and at least 10 microscopic fields were counted from 4 different slides for the same treatment for quantification of foam cells.

Flow Cytometry
Washed THP-1 cells were incubated with phycoerythrin-conjugated anti-human CD11b or fluorescein isothiocyanate–conjugated anti-human CD36 antibodies for 60 minutes on ice in the dark. Mouse bone marrow macrophages (BMMs) were incubated first with a mouse anti-mouse CD36 polyclonal antibody and second with a phycoerythrin-conjugated anti-mouse IgA, and mouse IgA was used as isotype control (all antibodies were from BD Biosciences, San Jose, Calif). At least 25,000 cells were analyzed on a Becton-Dickinson FACScan with Cell-Quest software.

Transfection of a Stat1 Decoy and Missense Oligodeoxyribonucleotides Into THP-1 and Mouse Primary Macrophages
A double-stranded DNA oligomer, used successfully as a DNA “decoy” in previous studies, was used to specifically inhibit Stat1 activity. The phosphorothioated oligodeoxyribonucleotides, the decoy and missense control oligomers, were purchased from Sigma-Genosys Biotechnologies, Inc (Woodlands, Texas).

Gel Shift Assay
For detection of Stat1 DNA binding to protein, a gel shift assay was performed as described. The probe used for Stat1 DNA binding was the consensus binding site for Stat1, double-stranded DNA 5'-CAT GTT AGT CAT ATT CCT GTA AGT G-3', purchased from Santa Cruz Biotechnology (Santa Cruz, Calif).

Isolation of Mouse Peritoneal Macrophages and Bone Marrow–Derived Macrophages
Elicited mouse peritoneal macrophages (MPMs) were harvested by peritoneal lavage with ice-cold PBS 2 to 3 days after intraperitoneal thioglycollate stimulation in female C57BL/6 mice. Primary cultures were prepared at a density of 10⁶ cells/16 mm diameter well in RPMI-1640 that contained 10% fetal bovine serum and used 48 hours after plating. For BMMs, femurs were dissected under sterile conditions as described. BMMs were cultured for 24 hours, washed with PBS, and incubated for at least 4 days before experiments in Dulbecco modified Eagles medium with 30% conditioned media from L-929 cells (media rich in granulocyte-macrophage colony-stimulating factor), supplemented with 10% fetal bovine serum and 50 U/mL of both penicillin and streptomycin.

Immunoblot Analysis
BMMs were lysed with radioimmunoprecipitation buffer (Roche Diagnostics, Penzberg, Germany). Protein was transferred to PVDF membrane for immunoblotting using a polyclonal anti-mouse CD36 antibody generated and characterized by M.F. (unpublished observations, 2006).

LDL Preparation, Oxidation, Uptake and Binding
LDL was isolated from human plasma as described. Cu-oxLDL was prepared as described. LDL was alternatively oxidized with myeloperoxidase-generated nitrating intermediates (NO₂-oxLDL), or modified by acetylation (Ac-LDL), as described. Lipoprotein iodination and their binding, uptake, and degradation by cultured cells were performed as described previously.

Intraperitoneal Foam Cell Formation
Sixteen-week old apolipoprotein e⁻/⁻ (apo e⁻/⁻) or apo e⁻/⁻/stat1⁻/⁻ male mice were fed a high-fat diet (42% calories from fat; Harlan Teklad TD.88137, Madison, Wis) for 4 weeks, and peritoneal cells were harvested and counted according to published protocols. Cells were washed with ice-cold PBS and plated for protein expression or foam cell formation measurements.

Bone Marrow Transplantation
The stat1⁻/⁻ mouse (SV129 background) was bred into the apo e⁻/⁻ mouse (C57Bl/6 background) for 5 generations, and genetically matched littermates that were either apo e⁻/⁻/stat1⁻/⁻ or apo e⁻/⁻ were used in intraperitoneal foam cell formation experiments or as bone marrow donors (4 males each of apo e⁻/⁻/stat1⁻/⁻ and apo e⁻/⁻) by previously described methods. Eight-week-old apo e⁻/⁻ male mice (C57 Bl/6, Jackson Laboratory, Bar Harbor, Me) (12 to receive apo e⁻/⁻ bone marrow and 12 to receive apo e⁻/⁻/stat1⁻/⁻ bone marrow) were lethally irradiated as per animal facility guidelines and previously published protocols. Polymerase chain reaction (PCR) for stat1⁻/⁻ or stat1⁻/⁻ alleles was performed on DNA isolated from whole blood samples from the recipient mice posttransplantation (see Animal Genotyping and PCR below). PCR analysis performed as previously described, revealed 75% to 100% engraftment of apo e⁻/⁻ or apo e⁻/⁻/stat1⁻/⁻ bone marrow into lethally irradiated apo e⁻/⁻ recipient mice (2 mice that received stat1⁻/⁻ bone marrow did not show significant engraftment of the stat1⁻/⁻ allele; data from these 2 mice were excluded).

Animal procedures were approved by the Cleveland Clinic Institutional Animal Care and Use Committee and performed in facilities accredited by the Association for Assessment and Accreditation of Laboratory Animal Care International.

Animal Genotyping and PCR
Genotyping was performed on ear clip–derived DNA. For apo e genotype, the PCR protocol described on the Jackson Laboratory Web site was used. For stat1 genotype PCR analysis, 2 different reactions were used: one that used the stat1 sense primer 5'-CTACCAAGTAGATCTGCTTAGAC-3' and antisense primer 5'-CCTCTCAACCCTCTGGACACC-3' to detect the wild-type allele, and one that used the stat1 sense primer 5'-CTACCAAGTAGATC TGCTTAGAC-3' and neoprimer 5'-CGCCGCTCCGATTCGCA-GGCCATCGC-3'. The reaction contained 3 μL of genomic DNA and 0.5 μL of the pooled primers at 50 pM/μL each. The Expand High Fidelity PCR system kit (Roche Diagnostics) was used. The reaction mixture was preheated to 94°C for 3 minutes, then run for 31 cycles at 94°C for 1 minute, 60°C for 1 minute, 72°C for 3 minutes, and then again at 72°C for 3 minutes at the end of the reaction. The products were run on a 2% agarose gel to distinguish the 285-bp wild-type apo e allele product from the 570-bp knockout allele product.

Atherosclerosis Lesion Measurements
Atherosclerotic lesions were quantified in bone marrow transplanted mice by 2 independent and blinded assessments: en face aortic coverage measured by computer-assisted planimetry, as described by
The authors had full access to and take full responsibility for the integrity of the data. All authors have read and agree to the manuscript as written.

Results

Blunting Stat1 Binding to DNA Inhibited Foam Cell Formation in Differentiated Human Monocytoid THP-1 Cells

To determine if Stat1 can regulate foam cell formation, a frequently studied in vitro foam cell model system was used: the human monocyte-like leukemic cell line THP-1. Stat1 had been shown to be involved early in cell differentiation; therefore, the effects of Stat1 on foam cell formation were only tested after differentiation. The time course and degree of Stat1 activation were determined after differentiation was initiated with PMA. Differentiation was monitored by CD11b/MAC-1 expression with use of fluorescence-activated cell-sorting analysis. CD11b reached a maximum and a plateau at 3 days (Figure 1A). Stat1 activation was assessed by immunoblot analysis with use of antibodies specific for Stat1 phosphorylation at serine-727 and tyrosine-701 and by measurement of binding of a radiolabeled DNA Stat1 consensus binding element to proteins in nuclear extracts. In both assays, adherent THP-1 cells expressed a relatively low but detectable basal Stat1 activity. This increased markedly in response to treatment with PMA; however, after 2 days Stat1 activation returned to basal levels (data not shown). Thus, to test the effect of Stat1 on foam cell formation independently of the effect of Stat1 on differentiation, experiments were conducted after 3 days.

Foam cell formation was induced postdifferentiation by exposure of THP-1-derived macrophages to Cu-oxLDL for 2 days. A double-stranded DNA decoy oligomer was transfected into differentiated THP-1 cells to block Stat1 binding to its target genes. The effectiveness of this decoy was assessed by measurement of the binding of a radiolabeled DNA Stat1 consensus binding element to proteins in the nuclear extract. Transfection of the Stat1 decoy reduced Stat1 binding markedly (80% to 85%) compared with differentiated THP-1 cells that were untreated, treated with Superfectin alone, or transfected with missense double-stranded DNA (Figure 1B). Foam cell formation, as identified by Oil Red O staining, was readily apparent in cells treated with Cu-oxLDL alone, Cu-oxLDL plus Superfectin, and Cu-oxLDL plus missense oligomer. Foam cell formation was reduced in cells treated with Cu-oxLDL plus the Stat1 decoy compared with the above-mentioned control cells (Figure 1C). Cholesteryl ester accumulation, also used to quantify foam cell formation, was significantly decreased by ~50% in Cu-oxLDL–incubated cells treated with the Stat1 decoy compared with Cu-oxLDL–incubated control, missense-treated, or Superfectin-treated cells (Figure 1D).

Stat1 Deficiency Inhibited Foam Cell Formation in Mouse Bone Marrow Macrophages

In an independent approach, BMMs from wild-type and stat1/−/− mice (SV129 genetic background) were obtained by harvesting of marrow cells and inducing differentiation. After 5 days, macrophages were treated with Cu-oxLDL for 48 hours to induce foam cell formation. Lipid accumulation, assessed with Oil Red O (see Methods), was 60% to 70% less in BMMs from stat1/−/− mice than in those from wild-type mice (Figure 2A). Cholesteryl ester accumulation after Cu-oxLDL treatment was reduced by ~50% in Cu-oxLDL–incubated cells treated with the Stat1 decoy compared with Cu-oxLDL–incubated control, missense-treated, or Superfectin-treated cells (Figure 1D).
oxLDL treatment was also attenuated in BMMs from stat1−/− mice compared with those from wild-type mice (5.8±0.5 μg/mg versus 9.8±1.7 μg/mg of protein; P<0.008) (Figure 2B). Similar results were obtained with use of MPMs from stat1−/− and wild-type mice (data not shown).

CD36 Expression Was Inhibited in THP-1 Cells After Stat1 Activity Was Blunted

Macrophages recognize and selectively ingest modified lipoproteins through multiple cell surface scavenger receptors to become foam cells. Deficiencies of either of 2 such receptors, SR-A and CD36, have been reported to reduce lesions in atherosclerosis-susceptible mouse models.29,30 We hypothesized that the mechanism of decreased foam cell formation after interference with Stat1 could be a result of Stat1 regulation of 1 or both receptors. Immunostaining for CD36 by flow cytometry revealed marginally reduced basal and significantly reduced Cu-oxLDL–induced CD36 expression on Stat1-decoy–treated and differentiated THP-1 cells compared with sham-treated control cells (Figure 3A).

CD36 Expression Was Inhibited in Bone Marrow Macrophages From stat1−/− Mice Compared With Those From Wild-Type Mice

BMMs from stat1−/− mice exposed to Cu-oxLDL showed significantly reduced basal expression of CD36 compared with those from wild-type mice by flow cytometry (Figure 3B) and Western blot analysis (Figure 3C). Cu-oxLDL enhanced CD36 protein expression in a time-dependent manner in BMMs from stat1−/− mice and wild-type mice, but the level of induction was significantly less in stat1−/− macrophages. Expression of SR-A was not affected by the absence of Stat1, as analyzed by immunoblotting and flow cytometry (data not shown). These data showed that at least a correlation existed between reduced Stat1 activity or Stat1 deficiency, reduced CD36 expression, and reduced foam cell formation.

To test whether decreased CD36 expression after interference with Stat1 resulted in reduced CD36 function (ie, decreased binding and cell association of CD36 ligands), particular forms of modified LDL that have been shown to be recognized by distinct scavenger receptors were used.12 It has been shown that LDL does not bind SR-A or CD36; Ac-LDL is a preferential ligand for SR-A; Cu-oxLDL is a ligand for both SR-A and CD36; and NO2-oxLDL is a preferential ligand for CD36.12 Thioglycollate-elicited MPMs from stat1−/− or wild-type mice were incubated with 75 μg/mL of the 125I-labeled ligands, LDL, Ac-LDL, or NO2-oxLDL, or with 50 μg/mL 125I-labeled Cu-oxLDL. Ligand binding (incubation at 4°C) and cell association (ie, binding and uptake...
after incubation at 37°C) were quantified as described. Binding of 125I-labeled Ac-LDL did not differ between wild-type and stat1/2/2 macrophages. In contrast, binding of 125I-labeled NO2-oxLDL and binding of 125I-labeled Cu-oxLDL were both significantly decreased in macrophages from stat1/2/2 mice compared with macrophages from wild-type mice (Figure 4A). Analogous results were obtained with BMMs from stat1/2/2 and wild-type mice (data not shown). The relationship between Stat1 regulation of CD36 expression, CD36 ligand recognition, and foam cell formation was probed further by treatment of BMMs from stat1/2/2 and wild-type mice with 50 μg/mL unlabeled Cu-oxLDL, NO2-oxLDL, or Ac-LDL for 48 hours and stained with the neutral lipid stain Oil Red O to visualize cytoplasmic lipid accumulation. Scale bar=10 μm. C, Foam cell formation, as measured by cholesteryl ester accumulation, was reduced in MPMs from wild-type or sr-a/2/2 mice compared with their respective missense-treated control cells after transfection of the cells with the Stat1 DNA oligomer “decoy.” Foam cell formation was unchanged by decoy treatment of macrophages from cd36/2/2 or cd36/2/2sr-a/2/2 mice. Specificity of the decoy was evaluated as shown in Figure 1B. Results shown are expressed as cholesteryl ester:protein ratios and are means±SD of data combined from 2 experiments.

Figure 4. CD36 deficiency diminished the Stat1 dependency of foam cell formation; Stat1 dependency of foam cell formation occurred for lipoprotein ligands recognized by CD36 (Cu-oxLDL, NO2-oxLDL), but not those recognized by SR-A (Ac-LDL). A, Specific ligand binding of 125I-labeled LDL, Ac-LDL, Cu-oxLDL, or NO2-oxLDL was assessed in MPMs from wild-type and stat1−− mice. Ac-LDL is a preferential ligand for SR-A, Cu-oxLDL is a ligand for both SR-A and CD36, and NO2-oxLDL is a preferential ligand for CD36. LDL does not bind either SR-A or CD36. Results are expressed as means±SD of triplicate measurements of specific μg 125I-labeled lipoprotein bound per mg cell protein after 5 hours. Asterisks indicate statistically significant differences compared with wild-type mice (P<0.001). B, MPMs from wild-type and stat1−− mice were treated with unlabeled LDL, Ac-LDL, Cu-oxLDL, or NO2-oxLDL for 48 hours and stained with the neutral lipid stain Oil Red O to visualize cytoplasmic lipid accumulation. Scale bar=10 μm. C, Foam cell formation was unchanged by decoy treatment of macrophages from cd36−− or cd36−−sr-a−− mice. Specificity of the decoy was evaluated as shown in Figure 1B. Results shown are expressed as cholesteryl ester:protein ratios and are means±SD of data combined from 2 experiments.
formation was significantly inhibited by the Stat1 decoy in MPMs from wild-type and sr-α− mice (Figure 4C). In contrast, no significant inhibition of foam cell formation was observed in Stat1 decoy-treated MPMs from cd36−/− or double-knockout mice (Figure 4C); the absence of CD36 removed the Stat1 dependency of foam cell formation. Similar results were observed in Stat1 decoy-treated BMMs from these animals (data not shown). These data demonstrated a requirement for CD36 in Stat1 regulation of foam cell formation and revealed that, in the absence of CD36, blockade of Stat1 activity did not inhibit foam cell formation by CD36-independent means.

Foam Cell Formation and CD36 Expression Were Inhibited In Vivo by Stat1 Deficiency
To determine whether foam cell formation in vivo was limited by stat1 deficiency, and whether CD36 expression was reduced in vivo by stat1 deficiency, a variation of the technique reported by Li et al31 was adapted (see Methods). Six days after peritoneal thioglycollate injection into either apoε−/− mice or apoε−/−×stat1−/− mice, peritoneal cells were harvested and assessed with Oil Red O staining and cholesteryl ester:protein measurement after allowing in vivo foam cell formation. Cell surface CD36 expression and plasma cholesterol were also quantified. Cholesteryl ester accumulation was significantly reduced in peritoneal cells from apoε−/−×stat1−/− mice compared with those from apoε−/− mice (1.29±0.1 μg/mg cell protein (n=12) versus 2.65±0.1 μg/mg (n=11); P<0.003) (Figure 5A), as was CD36 cell surface expression (546.3±79.2 relative units of intensity (n=12) versus 1102±164.0 (n=11); P<0.005) (Figure 5B). No significant difference existed in the total number of peritoneal cells harvested (27±1.7×10⁶ versus 28.2±2.0×10⁶; P<0.15) or plasma cholesterol levels (1869±239 μg/mL [n=12] versus 2194±120 μg/mL [n=11]; P<0.25) between the apoε−/− and apoε−/−×stat1−/− mice. Thus, Stat1 deficiency is linked to reduced CD36 expression on foam cells formed in vivo, analogous to the results obtained in vitro. In addition, the role of Stat1 in foam cell formation that was observed with the model of lipoproteins oxidized in vitro was recapitulated when foam cells were produced from ligands formed in vivo.

Atherosclerotic Lesion Formation Was Inhibited in apoε−/− Mice That Received Bone Marrow From apoε−/−×stat1−/− Compared With apoε−/− Mice That Received Bone Marrow From apoε−/− Mice
Eight-week-old apoε−/− mice were irradiated and received bone marrow transplants. After 4 weeks, they were fed a high-fat diet for an additional 14 week-old. apoε−/− mice that received bone marrow from apoε−/−×stat1−/− mice had significantly reduced en face aortic lesion area coverage compared with recipients of bone marrow from apoε−/− mice (1.41±0.4% (n=10) versus 4.33±0.8% (n=12), which is a 69% decrease (P<0.004) (Figure 6, A to C). Aortic root lesions were reduced from 274 400±24 360 μm² (n=12) in apoε−/− recipients to 165 100±18 740 μm² (n=10) in apoε−/−×stat1−/− recipients (Figure 6D through 6F), which is a ∼45% decrease (P<0.0026). Surprisingly, compared with apoε−/− mice that received apoε−/− bone marrow, mice that received apoε−/−×stat1−/− bone marrow were statistically significantly heavier (Figure 6H) and had significantly higher plasma cholesterol levels (Figure 6G). Immunohistochemistry with anti–phospho-Stat1 revealed that phosphorylated Stat1 was present in aortic root lesions from apoε−/− mice but not in aortic root lesions from apoε−/−×stat1−/− mice (data not shown).

Discussion
Our results reveal 2 novel findings. First, in both in vitro and in vivo macrophage cell systems, interference with Stat1-dependent pathways downregulated CD36 expression. Reduced foam cell formation in vitro by Stat1 pathway disruption depended on CD36. Second, our in vivo results showed that Stat1 deficiency reduced foam cell formation in an intraperitoneal inflammation model and reduced atherosclerosis in an atherosclerosis-susceptible bone marrow transplantation mouse model.

Our discovery that Stat1 helps regulate CD36 and foam cell formation is reinforced by the consistency of the findings in 3 distinct in vitro models (differentiated human THP-1 cells, BMMs, and MPMs), with use of different inducers of macrophage differentiation (PMA, granulocyte-macrophage colony-stimulating factor–rich L929-conditioned media, and thioglycollate recruitment and differentiation in vivo) and use of different means of Stat1 pathway interference (blockade of Stat1 binding to DNA and Stat1 deficiency). Consistent results were also obtained in an in vivo peritoneal inflammation model of foam cell formation, in which foam cell formation occurred in vivo, independent of in vitro–modified model lipoproteins.
responses to infections.9,33,34 CD36 has been implicated in the pathological and physiological processes, such as long-chain fatty acid transport, recognition and clearance of apoptotic cells, sequestration of the malarial parasite, and immune inflammatory response.35,36 But its role in atherosclerosis is a novel observation. Our findings are of particular interest in light of multiple prior observations. Stat1 is a downstream target in IFN-γ signaling, and several studies have identified a role for IFN-γ in atherosclerosis, such as in atherosclerosis-susceptible mice.4–6 Our data could be construed to suggest that Stat1 represents a step in the intracellular pathway by which IFN-γ or other cytokines worsen fatty streak formation and that CD36 might mediate this process. However, in vitro reports of the effect of IFN-γ on CD36 are inconclusive. IFN-γ has been shown to enhance and to decrease CD36 in various monocyte and macrophage-like cell systems. In addition, there are other possible explanations. Stat1 may be proatherogenic by an alternative mechanism. Stat1 activity was recently shown to contribute to 15-lipoxygenase expression,14 and 15-lipoxygenase expression is believed to be atherogenic.39,40

On the basis of our in vitro findings that CD36 mediates the reduced foam cell formation caused by interference with Stat1 pathways, it is tempting to speculate that the reduction in atherosclerosis we observed in Stat1 deficiency was also mediated by CD36. However, this would be premature. Although reduced foam cell lesions in atherosclerosis-susceptible mice deficient in either CD3690 or SR-A290 have been reported, a recent paper has questioned the role of scavenger receptors in atherosclerosis.41 In that study, in the absence of CD36 en face aortic analysis showed significant protection from atherosclerosis in female mice, but only a
trend toward reduced lesion area in males. Furthermore, the aortic root lesion data were discordant; females that lacked CD36 had larger lesions, and no significant difference was found in males. Thus, our data could reflect other CD36-independent and antiatherogenic influences exerted by the deficiency in Stat1.

Our data highlight the idea that Stat1 inhibition could represent a target to reduce inflammation. Certain substances known to inhibit Stat1, albeit nonselectively, namely the 3-hydroxy-3-methylglutaryl coenzyme A reductase—known to inhibit Stat1, albeit nonselectively, namely the 3-hydroxy-3-methylglutaryl coenzyme A reductase-inhibiting statins and epigallocatechin-3-gallate, have also been shown to inhibit atherosclerosis in humans and animals. Statins are known to have antiatherosclerotic effects in excess of that predicted from their effects on LDL lowering; our results invite speculation that a part of these pleiotropic effects of statins could be linked to Stat1 inhibition.

Acknowledgments
The authors wish to thank Charlie Kaul and Richard Cole for their technical assistance and Dr Jonathan D. Smith, whose laboratory performed aortic root lesion analysis.

Sources of Funding
This work was supported by National Institutes of Health grants PO1 HL29582 to Dr Chisolm, PO1 CA062220 to Dr Stark, RO1 HL70083-01 to Dr Febbraio, and RO1 HL077213 to Dr Podrez.

Disclosures
None.

References


**CLINICAL PERSPECTIVE**

Signal transducer and activator of transcription 1 (Stat1) is a very well-studied transcription factor in the janus kinase–Stat signaling pathway. It is a downstream target of interferon-γ and other specific cytokines and regulates numerous genes involved in immune responses, tumorigenesis, and inflammation. The results reported here reveal for the first time that Stat1 regulates the conversion of macrophages to lipid-engorged “foam cells,” an early cellular event in the pathological sequence of atherosclerotic lesion development. Furthermore, mice deficient in Stat1 developed significantly less atherosclerosis than did control mice in a bone marrow transplantation model of atherosclerosis. The in vitro data presented here show, also for the first time, that Stat1 regulates the scavenger receptor CD36, a protein linked to fatty acid transport, apoptotic cell recognition, metabolic syndrome, and responses to infection. The potential significance of these findings includes the possibility that further studies of Stat1 regulation of CD36 and its influence on atherosclerosis will identify novel therapeutic targets in pathological Stat1-dependent signaling pathways. Interestingly, certain substances known to inhibit Stat1, albeit nonselectively (eg, 3-hydroxy-3-methylglutaryl coenzyme A reductase-inhibiting statins and epigallocatechin-3-gallate) have also been shown to inhibit atherosclerosis in humans and animals. Because statins are known to have antiatherosclerotic effects in excess of that predicted from their effects on lowering low-density lipoprotein, it is tempting to speculate that a part of the pleiotropic effects of statins could be through Stat1 inhibition.
Signal Transducer and Activator of Transcription 1 Is Required for Optimal Foam Cell Formation and Atherosclerotic Lesion Development

Sudesh Agrawal, Maria Febbraio, Eugene Podrez, Martha K. Cathcart, George R. Stark and Guy M. Chisolm

_Circulation_. 2007;115:2939-2947; originally published online May 28, 2007; doi: 10.1161/CIRCULATIONAHA.107.696922

_Circulation_ is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2007 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circ.ahajournals.org/content/115/23/2939

Data Supplement (unedited) at:
http://circ.ahajournals.org/content/suppl/2007/05/18/CIRCULATIONAHA.107.696922.DC1

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in _Circulation_ can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to _Circulation_ is online at:
http://circ.ahajournals.org//subscriptions/