5-Methyltetrahydrofolate Rapidly Improves Endothelial Function and Decreases Superoxide Production in Human Vessels

Effects on Vascular Tetrahydrobiopterin Availability and Endothelial Nitric Oxide Synthase Coupling

Charalambos Antoniades, MD, PhD*; Cheerag Shirodaria, MRCP*; Nicholas Warrick, BSc, PhD; Shijie Cai, MD, PhD; Joseph de Bono, MRCP; Justin Lee, MRCP; Paul Leeson, PhD, MRCP; Stefan Neubauer, MD, FRCP; Chandi Ratnatunga, FRCS; Ravi Pillai, FRCS; Helga Refsum, MD, PhD; Keith M. Channon, MD, FRCP

Background—The circulating form of folic acid, 5-methyltetrahydrofolate (5-MTHF), may have beneficial effects on endothelial function; however, its mechanisms of action remain uncertain. Decreased nitric oxide (NO) bioavailability and increased vascular superoxide production in vascular disease states are due in part to endothelial NO synthase (eNOS) uncoupling related to deficiency of the eNOS cofactor tetrahydrobiopterin (BH4), but whether this mechanism is important in human atherosclerosis and represents a rational therapeutic target remains unclear. We hypothesized that 5-MTHF would improve endothelial function by decreasing superoxide and peroxynitrite production and by improving eNOS coupling, mediated by BH4 availability.

Methods and Results—Vascular superoxide/peroxynitrite production and vasomotor responses to acetylcholine and bradykinin were determined in saphenous veins and internal mammary arteries from 117 patients undergoing CABG. The effects of 5-MTHF were examined ex vivo (n=61) by incubating vessels with 5-MTHF (1 to 100 μmol/L) and in vivo by intravenous infusion of 5-MTHF or placebo before vessel harvest (n=56). 5-MTHF improved NO-mediated endothelium-dependent vasomotor responses and reduced vascular superoxide, both ex vivo and in vivo. These changes were not explained by direct superoxide scavenging by 5-MTHF in vitro or by changes in plasma total homocysteine in vivo. Rather, 5-MTHF was a strong peroxynitrite scavenger and increased vascular BH4 and the BH4/total biopterin ratio. Furthermore, 5-MTHF reversed eNOS uncoupling, as assessed by Nω-nitro-L-arginine methyl ester–inhibitable superoxide production, increased the eNOS dimer:monomer ratio, and enhanced eNOS activity.

Conclusions—5-MTHF has beneficial effects on endothelial function and vascular superoxide production in human atherosclerosis, by preventing peroxynitrite-mediated BH4 oxidation and improving eNOS coupling. (Circulation. 2006;114:1193-1201.)

Key Words: acetylcholine ■ antioxidants ■ coronary disease ■ endothelium ■ free radicals ■ nitric oxide ■ nitric oxide synthase

Increased plasma total homocysteine (tHcy) has been associated with increased cardiovascular risk, but it is still unclear whether lowering tHcy has antiatherogenic effects. Recent studies have suggested that folic acid, through its circulating form, 5-methyltetrahydrofolate (5-MTHF), may have antioxidant properties and exert biological effects in vascular cells that are not directly related to changes in plasma tHcy. Previous studies have suggested that folates may have direct effects on nitric oxide (NO)–mediated endothelial function, possibly through changes in endothelial NO synthase (eNOS) regulation mediated by the eNOS cofactor, tetrahydrobiopterin (BH4). However, the effects of 5-MTHF on vascular superoxide production and eNOS function in patients with atherosclerosis are unknown. Furthermore, the effects of folic acid supplementation on clinical outcomes remain uncertain; understanding the mechanism of

Received January 10, 2006; revision received June 14, 2006; accepted June 23, 2006.


*Drs Antoniades and Shirodaria contributed equally to the study.

Correspondence to Professor Keith M. Channon, Department of Cardiovascular Medicine, University of Oxford, John Radcliffe Hospital, Oxford, OX3 9DU, United Kingdom. E-mail keith.channon@cardiov.ox.ac.uk

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Circulation is available at http://www.circulationaha.org DOI: 10.1161/CIRCULATIONAHA.106.612325

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action of folic acid supplementation in human vascular disease is critical to guide future clinical studies.

Oxidative stress plays an important role in the pathogenesis of atherosclerosis, through effects of reactive oxygen species (ROS) on NO bioavailability and through interactions with numerous redox-sensitive signaling pathways.9,10 Indeed, the kinetics and appears to be an important mechanism linking oxidative stress to endothelial dysfunction.9,10 Indeed, the kinetics and progression of BH4 oxidation by peroxynitrite have been well defined,10 whereas the reaction rate between BH4 and superoxide is much smaller than that with peroxynitrite in vivo, which suggests that superoxide is not a major contributor of BH4 oxidation in vivo.9,11 Therefore, restoration of endothelial BH4 availability by scavenging peroxynitrite radicals in atherosclerosis may improve endothelial function and reduce atherosclerotic progression.12

In the present study, we explored the effects of 5-MTHF on endothelial function and vascular superoxide/peroxynitrite production in human vessels both ex vivo and in vivo. We hypothesized that 5-MTHF acts as an effective intracellular antioxidant and sought to evaluate the effects of 5-MTHF on eNOS “coupling” and on vascular BH4 availability in human atherosclerosis.

Methods

Patients

We studied 117 patients with coronary artery disease undergoing routine CABG surgery at the John Radcliffe Hospital, Oxford, United Kingdom. Exclusion criteria were the existence of any inflammatory, infective, liver, or renal disease or malignancy. Patients receiving nonsteroidal antiinflammatory drugs or any di-inhibitors or blockers or inhibitors of PDE, PPAR, or HMG-CoA reductase were excluded. Demographic characteristics of the patients are presented in the Table. The study protocol was approved by the local Research Ethics Committee, and each patient gave written informed consent.

Vessel Harvesting and Ex Vivo Studies

Samples of saphenous vein (SV; n=38) and internal mammary artery (IMA; n=46) from a total of 61 patients were harvested during CABG operation, as we have described previously.15,16 Vessel segments were transferred to the laboratory within 30 minutes in ice-cold Krebs-Henseleit buffer. Segments of SV and IMA were incubated with 0 (control), 1, 10, or 100 μmol/L 5-MTHF added to the organ equilibrating buffer and calculating the difference in superoxide was measured by adding L-NAME (100 μmol/L) to the equilibrating buffer and calculating the difference in superoxide signal compared with basal conditions. Vascular peroxynitrite was determined with luminol (100 μmol/L) instead of lucigenin and by subtracting the remaining signal after the addition of the specific peroxynitrite scavenger uric acid (1 mmol/L), as described previously.9

Oxidative Fluorescent Microtopography

In situ superoxide production was determined in vessel cryosections with the oxidative fluorescent dye dihydroethidium (DHE).16 Paired cryosections (30 μmol/L) from the same vessel were incubated with DHE (2 μmol/L) in PBS, with or without L-NAME (100 μmol/L). Fluorescence images (×40, Zeiss LSM 510 META laser scanning confocal microscope, Carl Zeiss GmbH, Jena, Germany) were obtained from each vessel quadrant, with the luminal side of the vessel incorporated to quantify endothelial cell fluorescence. In each case, segments of vessel rings (with and without L-NAME) were analyzed in parallel with identical imaging parameters. DHE image analysis was performed in a blinded fashion by 2 independent investigators using Image-Pro Plus software (Media Cybernetics, Silver Spring, Md).

Superoxide and Peroxynitrite Scavenging Assays

The direct superoxide scavenging effect of 5-MTHF was evaluated with the use of the xanthine/xanthine oxidase system. Superoxide production was induced by the addition of xanthine oxidase (6.67 MU/mL) in Krebs HEPES buffer (2 mL, pH 7.4; 37°C) containing xanthine (0.133 mmol/L) and lucigenin (5 μmol/L) in the presence of 5-MTHF (0 to 100 μmol/L) or equal concentrations of ascorbic acid as positive control. Peroxynitrite production was induced by addition of 3-morpholinosydnonimine (SIN-1, 1 μmol/L) in Krebs HEPES buffer (2 mL, pH 7.4; 37°C) containing luminol (100 μmol/L) in the presence of different concentrations of 5-MTHF (0 to 100 μmol/L) or equal concentrations of uric acid as positive control, as described previously.10 The scavenging effects of 5-MTHF,
vitamin C, or uric acid were calculated as the percent inhibition of chemiluminescence compared with control.

**Determination of 5-MTHF and tHcy**

Blood samples were collected before administration of 5-MTHF or placebo and at the time of vessel harvesting. tHcy was measured by a fluorescence polarization immunoassay adapted to the IMx analyzer (Abbott Diagnostics, Abbott Park, Ill). Plasma and tissue levels of 5-MTHF were determined by high-performance liquid chromatography (HPLC), as described previously.17

**Determination of Biopterin Levels**

Biopterin levels in human vessels were determined by HPLC, as described previously,18 and expressed as pmol/g of tissue. To examine the effect of 5-MTHF on peroxynitrite-induced oxidation of BH4, we used SIN-1 (2 μmol/L) to oxidize BH4 (0.1 μmol/L) in the presence or absence of 5-MTHF (1 μmol/L) at 37°C for 14 minutes.19

**Evaluation of eNOS Activity in Intact Vessels**

The activity of eNOS was estimated by HPLC quantification of radiolabeled arginine to citrulline conversion from intact vessel rings, as described recently.20

**Statistical Analysis**

Analyses were performed with the SPSS 12.0 statistical package for Windows (SPSS Inc, Chicago, Ill). Normally distributed data are presented as mean±SEM, whereas nonnormally distributed variables (such as vascular superoxide) are presented as median (25th to 75th percentile values). Baseline comparisons between groups were performed with 1-way ANOVA for multiple comparisons, followed by Bonferroni correction. The effects of 5-MTHF on vasomotor responses in each vessel ring were assessed by 2-way ANOVA for repeated measurements. The effects of 5-MTHF incubations on superoxide production and levels of 5-MTHF or tHcy were assessed by Mann-Whitney U tests, Wilcoxon signed rank tests, or t tests for unpaired or paired data, as appropriate. A 2-tailed P<0.05 was considered statistically significant.

### Western Blot Analysis

The eNOS dimer:monomer ratio in paired IMA samples incubated for 45 minutes in the presence or absence of 5-MTHF 1 μmol/L was measured by Western blotting as described previously.21 Bands were visualized with chemiluminescence and quantified with NIH Image software (National Institutes of Health, Bethesda, Md).

### Demographic Characteristics of the Participants

<table>
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<tr>
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<th>Ex Vivo Study</th>
<th>Intravenous 5-MTHF Study</th>
<th>Placebo</th>
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<td>Males/females, n</td>
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<td>22/2</td>
<td>27/5</td>
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<td>SVs obtained, n</td>
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<tr>
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<td>25</td>
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<tr>
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<td>1.69±0.25</td>
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<td>1.09±0.06</td>
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<td>Plasma tHcy, μmol/L</td>
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<td>11.5±1.07</td>
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<td>Plasma 5-MTHF, nmol/L</td>
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<td>32.4±9.01</td>
<td>35.4±8.10</td>
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<td>2-Vessel disease</td>
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<td>6 (19)</td>
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<td>20 (83)</td>
<td>26 (81)</td>
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<td>28 (88)</td>
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<td>9 (28)</td>
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<td>2 (8)</td>
<td>2 (6)</td>
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<td>β-Blocker</td>
<td>53 (87)</td>
<td>20 (83)</td>
<td>28 (88)</td>
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<td>Nitrates</td>
<td>28 (46)</td>
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<td>Aspirin</td>
<td>47 (79)</td>
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<td>24 (75)</td>
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<tr>
<td>Clopidogrel</td>
<td>18 (30)</td>
<td>8 (33)</td>
<td>10 (31)</td>
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CAD indicates coronary artery disease. Values are mean±SEM or n (%). There were no significant differences between groups in clinical characteristics or medication.

<table>
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<tr>
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<th>Ex Vivo Study</th>
<th>Intravenous 5-MTHF Study</th>
<th>Placebo</th>
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<tr>
<td>CAD indicates coronary artery disease. Values are mean±SEM or n (%). There were no significant differences between groups in clinical characteristics or medication.</td>
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</table>
The authors had full access to the data and take full responsibility for its integrity. All authors have read and agree to the manuscript as written.

Results

Patient Characteristics
Vessels were obtained from 117 patients (61 included in the ex vivo part of the study and 56 in the in vivo part). There were no significant differences in demographic characteristics or baseline plasma tHcy levels between the treatment groups (Table).

Effects of 5-MTHF on Endothelial Function and on Superoxide and Peroxynitrite Production in Human Vessels Ex Vivo
We first investigated the effects of 5-MTHF on vasomotor responses to ACh and BK in vessel segments at baseline and after 45-minute incubation in organ chambers with 0 to 100 μmol/L 5-MTHF. Relaxations in response to ACh and BK were similar between the 4 rings from the same vessel at baseline. Maximum relaxations to ACh and BK were similar between the 4 rings from the same vessel at baseline and remained unchanged after incubation with 5-MTHF. Absolute contractions in response to phenylephrine were 7.9±0.8g at baseline and remained unchanged after incubation. *P<0.05, **P<0.01 vs baseline.

Figure 1. Isometric tension studies in segments of SVs from 30 patients incubated with increasing concentrations of 5-MTHF for 45 minutes. Baseline relaxations were identical between the 4 groups and are presented as a single curve. Vessel relaxations to the endothelium-dependent agonists ACh (A) or BK (Bk; B) did not change in control vessels but increased significantly after incubation with 5-MTHF. Absolute contractions in response to phenylephrine were 7.9±0.8g at baseline and remained unchanged after incubation.

Correspondingly, tissue 5-MTHF levels in SV (9.35±1.72 nmol/L; P<0.01 vs control) were increased at 45 minutes of incubation of 25 SVs with 5-MTHF at 1, 10, and 100 μmol/L, from 0.40±0.71, 21.3±8.10, and 57.2±18.9 nmol/g tissue after incubation of 25 SVs with 5-MTHF at 1, 10, and 100 μmol/L, respectively (P<0.05 for each versus control). There were no differences in vasomotor responses to the endothelium-dependent agonist SNP between control vessel segments and those incubated with 5-MTHF (data not shown), which indicates a specific effect of 5-MTHF on NO-mediated endothelial function.

Vascular superoxide and peroxynitrite production from SV and IMA were determined after 45 minutes’ incubation with either buffer alone or with 5-MTHF at 1, 10, or 100 μmol/L. Incubation with 1 μmol/L 5-MTHF significantly reduced both vascular superoxide and peroxynitrite production in SV and IMA, with no further reductions at higher 5-MTHF concentrations (Figure 2).

Effects of 5-MTHF on Endothelial Function and Superoxide Production in Human Vessels In Vivo
On the basis of our findings of improved endothelial function and reduced superoxide and peroxynitrite production after ex vivo incubation with 1 μmol/L 5-MTHF, but no further effects at higher concentrations, we aimed to test the effects of similar 5-MTHF concentrations in vivo. In pilot studies, 0.13 mg/kg body weight 5-MTHF administered intravenously increased plasma 5-MTHF from 74.1±22.2 nmol/L to 2.48±0.47 μmol/L (n=4) 5 minutes after infusion and to 1.71±8.28 μmol/L at 45 minutes.

Patients were randomized to receive either intravenous 5-MTHF or placebo in a double-blind fashion. Baseline plasma levels of 5-MTHF were not significantly different between patients who received intravenous 5-MTHF (32.4±9.01 nmol/L) and those who received placebo (26.4±4.68 nmol/L, P=NS). In contrast, plasma 5-MTHF levels were increased significantly at the time of vessel harvesting in the 5-MTHF–treated group (2.28±0.21 μmol/L; P<0.001 versus baseline) but remained unchanged in the placebo-treated group (25.8±6.85 nmol/L, P=NS versus baseline, P<0.001 versus 5-MTHF–treated group). Correspondingly, tissue 5-MTHF levels in SV (9.35±1.72
Capacity of 5-MTHF

Direct Superoxide/Peroxynitrite Scavenging Capacity of 5-MTHF

To investigate whether direct superoxide scavenging by 5-MTHF could account for its effects on vascular superoxide production and endothelial function, we assessed the ability of 5-MTHF to scavenge superoxide in comparison with a known superoxide scavenger, ascorbic acid. When added to a xanthine/xanthine oxidase system that generated superoxide at similar levels to those observed in vascular tissues, ascorbic acid had a potent scavenging effect, reducing measurable superoxide by 50% at 1 μmol/L (Figure 5). In contrast, low concentrations of 5-MTHF (1 to 10 μmol/L) had no detectable effect on measurable superoxide (Figure 5), with only modest superoxide scavenging observed even at very high 5-MTHF concentrations (100 μmol/L).

The direct peroxynitrite scavenging capacity of 5-MTHF was assessed in comparison with a known peroxynitrite scavenger, uric acid. When added to an SIN-1 system, 5-MTHF had a potent scavenging effect, reducing measurable peroxynitrite by 75% at 1 μmol/L (Figure 5), comparable to the effect of uric acid at the same concentration. These observations suggest that 5-MTHF, at concentrations used in vivo, has a strong direct peroxynitrite scavenging capacity.

<table>
<thead>
<tr>
<th>Concentration (μmol/L)</th>
<th>5-MTHF</th>
<th>Placebo</th>
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<tbody>
<tr>
<td>0.01</td>
<td>0.92</td>
<td>0.94</td>
</tr>
<tr>
<td>0.1</td>
<td>0.88</td>
<td>0.90</td>
</tr>
<tr>
<td>1.0</td>
<td>0.84</td>
<td>0.86</td>
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</table>

Results from Figure 5 demonstrate that 5-MTHF has a strong direct peroxynitrite scavenging effect at concentrations as low as 1 μmol/L, comparable to that of equal concentrations of uric acid. Values are mean ± SEM of 3 separate experiments. *P<0.01 vs vitamin C; †P<0.05 and ‡P<0.01 vs 0 μmol/L.
the present in vivo studies, exerts a significant peroxynitrite but not superoxide-scavenging effect.

Effects of 5-MTHF on eNOS-Derived Vascular Superoxide Production and eNOS Activity

Because the direct superoxide scavenging capacity of 5-MTHF did not account for the observed reduction in vascular superoxide production, we next examined whether the antioxidant effect of 5-MTHF was due to an improvement in eNOS coupling. NOS-derived superoxide production was estimated in paired samples of SV and IMA by quantifying the effects of NOS inhibition with L-NAME. Vascular superoxide production was decreased by L-NAME in both SV and IMA in the placebo group, which suggests a net contribution to vascular superoxide production by NOS. In contrast, L-NAME increased superoxide production in vessels from 5-MTHF–treated patients, which suggests net NO production (Figure 6). Importantly, L-NAME–inhibitable superoxide production in SV or IMA was correlated with plasma 5-MTHF (r=0.511, P=0.006 and r=0.690, P=0.0001, respectively) but not with plasma tHcy levels (r=0.040, P=0.829 and r=−0.286, P=0.106, respectively).

To visualize vascular superoxide production and to specify the changes in eNOS-mediated endothelial superoxide production, we used oxidative fluorescent microtopography with DHE (Figure 6). L-NAME decreased endothelial DHE fluorescence in vessels from placebo patients, which indicates eNOS uncoupling. Conversely, LNAME increased endothelial DHE fluorescence in vessels from 5-MTHF–treated patients, which suggests an improvement in eNOS coupling in these vessels (Figure 6). Importantly, DHE fluorescence in other regions of the vessel wall was unaffected by 5-MTHF, which provides a within-section control and demonstrates the endothelium-specific effect of NOS inhibition.

To further evaluate the effects of 5-MTHF on eNOS-activity, we used HPLC to measure the conversion of radiolabeled arginine to citrulline in samples of IMA from 12 patients, incubated with or without 5-MTHF 1 μmol/L for 45 minutes. We observed a significant increase in citrulline production in vessels incubated with 5-MTHF (0.20±0.03%/g tissue) compared with paired control vessels from the same patients (0.14±0.02%/g, P<0.05), which suggests that 5-MTHF directly increases eNOS activity in human vessels. Taken together, these observations suggest that 5-MTHF reduces vascular superoxide production and improves NO-mediated endothelial function through effects on eNOS coupling.

Effects of 5-MTHF on BH₄ Levels and eNOS Dimer:Monomer Ratio in Human Vessels

To further investigate the mechanisms by which 5-MTHF improves eNOS coupling, we examined the effects of 5-MTHF on vascular BH₄ levels. Incubation of SV for 45 minutes ex vivo with 1 μmol/L 5-MTHF significantly increased vascular BH₄ and the BH₄/tBio ratio (Figure 7). More strikingly, both the absolute levels of BH₄ and the BH₄/tBio ratio were significantly elevated in patients who received intravenous infusion of 5-MTHF in vivo compared with placebo-treated patients (Figure 7). Because we observed that 5-MTHF is a strong peroxynitrite scavenger, we then examined the effects of 5-MTHF on peroxynitrite-induced BH₄ oxidation. 5-MTHF significantly reduced the decrease of both BH₄ and BH₄/tBio ratio after exposure to peroxynitrite generated by SIN-1 (Figure 7). Furthermore, the dimer:monomer ratio was also increased after exposure of IMA rings to 5-MTHF 1 μmol/L (Figure 7). We also found that the vascular BH₄/tBio ratio was significantly correlated with the L-NAME–induced change in superoxide production (Figure 8) in both SV (r=0.495, P=0.002) and IMA (r=0.621, P=0.001) from individual patients. Finally, the BH₄/tBio ratio in SV and IMA was significantly correlated with plasma 5-MTHF (r=0.498, P=0.01 and r=0.656, P=0.0001, respectively) but not with plasma tHcy (r=0.033, P=0.836 and r=−0.165, P=0.374, respectively).
respectively), which suggests that 5-MTHF itself rather than plasma tHcy is the critical parameter that prevents the oxidation of BH4 in human vessels in vivo. These observations suggest a direct functional relationship between vascular BH4 availability and eNOS coupling in human vessels that is a major determinant of both NO-mediated endothelial function and vascular superoxide production.

**Discussion**

In the present study, we show that 5-MTHF, the circulating form of folic acid, has striking direct effects on vascular function. 5-MTHF acutely improves NO-mediated endothelial function and decreases superoxide production, through mechanisms independent of direct superoxide scavenging. Rather, the effects of 5-MTHF on the vascular endothelium appear to be mediated through restoration of the enzymatic coupling of eNOS due to increased vascular BH4 availability. This effect may be due to the protective effect of 5-MTHF on peroxynitrite-induced oxidation of BH4. Taken together, these findings demonstrate that BH4-mediated regulation of eNOS is an important determinant of endothelial function and superoxide production in human atherosclerosis. More generally, they provide proof-of-principle that targeting specific redox mechanisms in human atherosclerosis can provide a rational basis for therapeutic intervention.

Plasma levels of tHcy1 and folate22 are related to cardiovascular risk1 and endothelial function.4,7,23 Early clinical trials suggested that lowering tHcy with folic acid may retard progression of atherosclerosis,24 but these findings were not confirmed by more recent studies.25 Whereas high-dose folic acid treatment may be beneficial specifically in patients with high tHcy levels,2 benefits in other patient groups have been inconsistent.26 Three recent larger trials in patients with stroke,2 myocardial infarction,27 or stable coronary artery disease28 found that folic acid treatment did not improve clinical outcome. In part, the continued uncertainty over the effects of folate status and tHcy-lowering relates to incomplete understanding of the mechanistic relationships between folate, homocysteine, and vascular disease. Evidence suggests that mechanisms other than homocysteine-lowering may be important factors in the relationship between folate and atherosclerosis. Recent studies have reported beneficial effects of folic acid treatment on endothelial function in patients with advanced atherosclerosis that are discordant with changes in plasma tHcy.29 More specifically, 5-MTHF appears to be most strongly associated with endothelial function, independent of tHcy levels.30 Indeed, direct intra-

Figure 7. Tissue biopterin levels in SVs and IMAs. Ex vivo incubations for 45 minutes with 5-MTHF (1 μmol/L) increased BH4 levels (A) and BH4/tBio ratio (B) compared with control in both SV (n=20 pairs) and IMA (n=12 pairs). Similarly, vascular BH4 (C) and BH4/tBio ratio (D) were higher in vessels obtained after infusion of 5-MTHF (0.13 mg/kg, n=23 for SV and n=14 for IMA) than in vessels from patients who received placebo (n=19 for SV and n=19 for IMA). There was no significant difference in demographic characteristics or medication use between patients in the 2 groups. Shaded bars indicate control or placebo; open bars, 5-MTHF. *P<0.05; **P<0.01 vs control or placebo. 5-MTHF (1 μmol/L) prevented the decrease in BH4 (E) and BH4/tBio ratio (F) induced by exposure of BH4 (0.1 μmol/L) to SIN-1 (2 μmol/L) for 14 minutes. †P<0.05 and ‡P<0.01 vs BH4 alone; *P<0.05 and **P<0.01 vs BH4+SIN-1. Ex vivo incubation of IMA (n=5 pairs) with 5-MTHF significantly increased eNOS dimer:monomer ratio as evaluated by immunoblotting (G) after quantification of eNOS band intensity (H; *P<0.05 vs control).

Figure 8. Vascular BH4/tBio ratio was correlated with the L-NAME–induced change in superoxide production in both SVs (r=0.495, P=0.002) and IMAs (r=0.621, P=0.001) from patients who received either placebo or 5-MTHF in vivo. Shown is the combined correlation for SV and IMA. Red open dots indicate placebo-treated IMA; red solid dots, 5-MTHF–treated IMA; black open dots, placebo-treated SV; black solid dots, 5-MTHF–treated SV; and RLU/sec/mg, relative light units per second per milligram.
arterial infusion of 5-MTHF improved flow-mediated dilation in patients with coronary artery disease before any detectable reduction in plasma tHcy. Because 5-MTHF is a reduced form of folic acid that does not require conversion by dihydrofolate reductase, some direct effects may be attributable to redox mechanisms that are not seen when oral folic acid is used to increase plasma folate levels.

We now demonstrate that 5-MTHF has a rapid, direct effect on the human vascular endothelium in a concentration range that is readily achievable in vivo. Specifically, incubation of vessels with 1 μmol/L 5-MTHF for 45 minutes improved vasomotor responses to both ACh and BK, with no further improvement at higher concentrations. Accordingly, we used an intravenous infusion of 5-MTHF in patients undergoing CABG that was sufficient to increase circulating 5-MTHF levels to 1 to 3 μmol/L. These concentrations also improved vasomotor responses to both ACh and BK in SV segments, which suggests an improvement in NO bioavailability in these vessels. The present study adds further weight to the notion that the salutary effects of 5-MTHF on vascular function are not mediated solely through changes in plasma tHcy levels. Although we observed a small decrease in plasma tHcy levels 45 minutes after 5-MTHF administration in vivo, plasma tHcy levels were not related to vasomotor responses to ACh or BK or vascular superoxide production.

Our findings suggest that 5-MTHF has specific rather than general antioxidant effects in human atherosclerosis. It is likely that chronic treatment with folic acid does exert some biological effects by decreasing homocysteine levels. At very high concentrations (100 to 500 μmol/L), 5-MTHF may have also direct antioxidant properties, as confirmed in our own studies. However, we observed striking effects on endothelial function and a decrease of superoxide production at 5-MTHF concentrations as low as 1 μmol/L, both ex vivo and in vivo, with no evidence of significant direct superoxide scavenging. The assays of superoxide production used in clinical material have limited quantitative specificity and could be improved, for example, by HPLC analysis of DHE oxidation. However, we also found that 5-MTHF was a strong scavenger for peroxynitrite in vitro and reduced peroxynitrite production from vessel rings. These observations now add major new clinical importance to the previous key finding that 5-MTHF can interact with purified eNOS in vitro, preventing its “uncoupling.” We observed that 5-MTHF could enhance eNOS activity and qualitatively restore eNOS coupling in human vessels, from a net producer of superoxide (as revealed by inhibition of superoxide production by L-NAME) to a net producer of NO (leading to increased superoxide release by L-NAME). We also observed that 5-MTHF increased the dimer:monomer ratio in human vessels, which suggests an improvement in eNOS protein dimerization.

A major determinant of eNOS function and enzymatic coupling is the cofactor BH₄. Recent studies in both transgenic mice and endothelial cells have shown that an imbalance between intracellular eNOS and BH₄ availability levels leads to uncoupling of eNOS, which results in superoxide production. BH₄ deficiency in vascular disease states appears to result principally from its increased intracellular oxidation by peroxynitrite. The present findings provide evidence that although 5-MTHF is a rather weak superoxide scavenger, it may be acting to increase intracellular BH₄ by preventing intracellular oxidation by peroxynitrite. Previous observations suggested that 5-MTHF could directly increase NO production by recombinant eNOS in the presence of BH₄, and it may also interact with the pterin-binding site of eNOS in a fashion analogous to BH₄.

In summary, we describe a striking effect of 5-MTHF on NO-mediated endothelial function and on superoxide production in human vessels from patients with atherosclerosis, both in vivo and ex vivo. We propose that 5-MTHF may prevent the peroxynitrite-induced oxidation of BH₄, leading to an improvement in eNOS coupling and an enhancement of eNOS activity. These findings show, in general, that targeting of specific redox mechanisms that regulate endothelial function in human atherosclerosis is a rational therapeutic approach. Specifically, BH₄ availability and eNOS function may be a key mechanism that can be targeted with 5-MTHF, or other therapies, to improve endothelial function in patients with vascular disease. Finally, the present study adds mechanistic insights to enable us to further understand the complex relationships between homocysteine, folate, and vascular disease. Identifying the mechanisms of action of folate in vascular disease states will allow more appropriate selection of patient groups and therapeutic regimens for future clinical trials.

Acknowledgments

We are grateful to Dr R. Moser, Merck Eprova AG, Switzerland, an affiliate of Merck KGaA, Darmstadt, Germany, for providing 5-MTHF. We are also grateful to David P. Taggart for allowing his patients to participate in this study.

Sources of Funding

Dr Antoniades is supported by a Marie Curie Intra-European Fellowship. This work was also supported by grants from the British Heart Foundation (RG/02/006 to Professor Channon and FS/03/105/16340 to Dr Shirodaria).

Disclosures

None.

References

The importance of homocysteine-lowering treatment with folic acid remains controversial, because initial reports that suggest a beneficial effect of folic acid in human atherosclerosis have not been confirmed by recent large-scale clinical trials. Understanding the mechanisms by which folic acid affects vascular function is critical to the design of future therapeutic strategies. We evaluated the effects of 5-methyltetrahydrofolate (5-MTHF), the circulating form of folic acid, in patients with coronary artery disease. We first studied 5-MTHF incubation of vessel rings ex vivo, then conducted a double-blind, placebo-controlled clinical trial of intravenous 5-MTHF. We demonstrate that 5-MTHF rapidly improves endothelial function and decreases superoxide production in vessels from patients with coronary artery disease by mechanisms that appear independent of homocysteine lowering. The effects of 5-MTHF are due in part to direct scavenging of the oxidant radical peroxynitrite. By this mechanism, 5-MTHF improves bioavailability of the endothelial nitric oxide synthase (eNOS) cofactor, tetrahydrobiopterin (BH4), which leads to an improvement in eNOS “coupling” and a decrease in eNOS-derived superoxide production. Thus, 5-MTHF has direct effects on vascular endothelium by improving BH4-mediated eNOS coupling in vessels from patients with coronary artery disease. Future clinical studies are required to evaluate how these novel effects of 5-MTHF, rather than homocysteine-lowering alone, may be related to clinical benefits in vascular disease.
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Charalambos Antoniades, Cheerag Shirodaria, Nicholas Warrick, Shijie Cai, Joseph de Bono, Justin Lee, Paul Leeson, Stefan Neubauer, Chandi Ratnatunga, Ravi Pillai, Helga Refsum and Keith M. Channon

Circulation. 2006;114:1193-1201; originally published online August 28, 2006; doi: 10.1161/CIRCULATIONAHA.106.612325
Circulation is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2006 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the World Wide Web at:
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