Overexpression of Mitochondrial Peroxiredoxin-3 Prevents Left Ventricular Remodeling and Failure After Myocardial Infarction in Mice

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Background—Mitochondrial oxidative stress and damage play major roles in the development and progression of left ventricular (LV) remodeling and failure after myocardial infarction (MI). We hypothesized that overexpression of the mitochondrial antioxidant, peroxiredoxin-3 (Prx-3), could attenuate this deleterious process.

Methods and Results—We created MI in 12- to 16-week-old, male Prx-3–transgenic mice (TG+MI, n=37) and nontransgenic wild-type mice (WT+MI, n=39) by ligating the left coronary artery. Prx-3 protein levels were 1.8 times higher in the hearts from TG than WT mice, with no significant changes in other antioxidant enzymes. At 4 weeks after MI, LV thiobarbituric acid–reactive substances in the mitochondria were significantly lower in TG+MI than in WT+MI mice (mean±SEM, 1.5±0.2 vs 2.2±0.2 nmol/mg protein; n=8 each, P<0.05). LV cavity dilatation and dysfunction were attenuated in TG+MI compared with WT+MI mice, with no significant differences in infarct size (56±1% vs 55±1%; n=6 each, P=NS) and aortic pressure between groups. Mean LV end-diastolic pressures and lung weights in TG+MI mice were also larger than those in WT+ sham-operated mice but smaller than those in WT+MI mice. Improvement in LV function in TG+MI mice was accompanied by a decrease in myocardial oxidative stress including the antioxidant Prx-3 might be beneficial in preventing cardiac failure. (Circulation. 2006;113:1779-1786.)

Conclusions—Overexpression of Prx-3 inhibited LV remodeling and failure after MI. Therapies designed to interfere with mitochondrial oxidative stress including the antioxidant Prx-3 might be beneficial in preventing cardiac failure.

Key Words: antioxidants ■ free radicals ■ heart failure ■ myocardial infarction ■ remodeling
either a large transmural MI induced by coronary artery ligation or sham operation.

**Methods**

**Generation of TG Mice**

The rat Prx-3 cDNA fragment including the entire open reading frame from nucleotide 5 to 802 was amplified by polymerase chain reaction (PCR) and cloned into pCR II (Invitrogen, Carlsbad, Calif.). An expression vector for Prx-3 was constructed with pQBI25 (TaKaRa), and the gene for green fluorescent protein was removed at the site of NheI-BamHI. A cytomegalovirus promoter–driven expression cassette containing rat Prx-3 cDNA in the sense orientation was purified by ultracentrifugation with CsCl. The pronuclei of fertilized eggs from hypervolulted C57BL/6j mice were randomly microinjected with this DNA construct. Tail clips and a PCR protocol to confirm the genotype were performed by one group of investigators. Homozygous TG mice and C57BL/6j WT mice were used at 12 to 16 weeks of age. The study was approved by our institutional animal research committee and conformed to the animal care guidelines of the American Physiological Society.

**Creation of MI**

We created MI in 12- to 16-week-old, male TG mice (TG+MI) and nontransgenic WT littermates (WT+MI) by ligation of the left coronary artery. Sham operation without coronary artery ligation was also performed in WT (WT+sham) and TG (TG+sham) mice. This assignment procedure was performed with the use of numeric codes to identify the animals.

**Prx-3 Protein**

Prx-3 protein levels were analyzed in cardiac tissue homogenates by Western blot analysis with a monoclonal antibody against rat Prx-3. Our preliminary studies revealed that this antibody against rat Prx-3 cross-reacted with mouse Prx-3 as a single band of 25 kDa. In brief, the LV tissues were homogenized with lysis buffer (20 mmol/L Tris-HCl, 1 mmol/L EDTA, 1 mmol/L EGTA, and 1 mmol/L phenylmethylsulfonylfluoride; pH 7.4). After centrifugation, equal amounts of protein (5 μg protein/lane), estimated by the Bradford method with a protein assay (Bio-Rad, Hercules, Calif.), were electrophoresed on a 15% sodium dodecyl sulfate–polyacrylamide gel and then electrophoretically transferred to a nitrocellulose membrane (Millipore, Billerica, Mass). After being blocked with 5% nonfat milk in phosphate-buffered saline (PBS) containing 0.05% Tween 20 at 20°C for 1 hour, the membrane was incubated with the first antibody and then with the peroxidase-linked secondary antibody (Amersham Pharmacia, Uppsala, Sweden). Chemiluminescence was detected with an enhanced chemiluminescence Western blot detection kit (Amersham Pharmacia) according to the manufacturer’s recommendation.

To further assess the subcellular localization of Prx-3 protein, mitochondrial and cytoplasmatic fractions were prepared from LVs and subjected to Western blot analysis. In brief, the LV tissues were homogenized at 4°C for 1 minute in 6 volumes of buffer consisting of 10 mmol/L HEPES-NaOH (pH 7.4), 1 mmol/L disodium EDTA, and 250 mmol/L sucrose. The homogenate was centrifuged at 4°C and 3000g for 10 minutes to remove any nuclear and myofibrillar debris, and the resultant supernatant was centrifuged at 10 000g for 10 minutes to separate any cardiac subcellular fractions. The supernatant was used for the cytoplasmatic fraction assay. To isolate the mitochondrial fraction, the pellet was resuspended at 4°C in a buffer consisting of 10 mmol/L HEPES-NaOH (pH 7.4), 1 mmol/L sodium EDTA, and 250 mmol/L sucrose and was washed 3 times with the same buffer. Mitochondria antibodies directed toward glyceraldehyde-3-phosphate dehydrogenase (GAPDH) and cytochrome oxidase (COX) subunit I were also used to verify the integrity of these subcellular fractions.

**Immunohistochemistry**

Frozen sections of cardiac tissues were incubated in the presence of 100 mmol/L MitoTracker Red CMXRos (Molecular Probes, Eugene, Ore) at 37°C for 20 minutes. We did not repeat the freeze/thaw procedure to avoid the loss of mitochondrial integrity. After being washed with PBS (10 mmol/L sodium phosphate, pH 7.4, and 150 mmol/L NaCl), the sections were fixed with 3.7% formaldehyde for 5 minutes. After being washed, the fixed sections were incubated with 100-fold–diluted anti-rat Prx-3 antibody (10 μg/mL) in PBS at 4°C overnight. Fluorescence images were taken with a confocal laser scanning microscope (Bio-Rad MRC 1024) with laser beams of 488 and 568 nm for excitation.

**Myocardial Antioxidant Enzyme Activities and Lipid Peroxidation**

For the subsequent biochemical studies, the myocardial tissues with MI were carefully dissected into 3 parts: one consisting of the infarcted LV, one consisting of the border zone LV with the peri-infarct rim (a 1-mm rim of normal-appearing tissue), and one consisting of the remaining noninfarcted (remote) LV. The antioxidant enzymatic activities of superoxide dismutase (SOD), catalase, and glutathione peroxidase (GSHPs) were measured in the noninfarcted LV.11 The formation of lipid peroxides was measured in the mitochondrial fraction isolated from the LV myocardium with use of a biochemical assay with thiobarbituric acid–reactive substances (TBARS).4

**Survival**

A survival analysis was performed in WT+sham (n=15), TG+sham (n=14), WT+MI (n=39), and TG+MI (n=37) mice. During the study period of 4 weeks, the cages were inspected daily for deceased animals. All deceased mice were examined for the presence of MI as well as pleural effusion and cardiac rupture.

**Echocardiographic and Hemodynamic Measurements**

At 4 weeks after surgery, echocardiographic studies were performed under light anesthesia with tribromoethanol/amylene hydrate (2.5% wt/vol, 8 μL/g IP) and spontaneous respiration. Two-dimensional-targeted M-mode tracings were recorded at a paper speed of 50 mm/s. Under the same anesthesia with Avertin, a 1.4F micrometer-tipped catheter (Millar Instruments, Houston, Tex) was inserted into the right carotid artery and then advanced into the LV to measure LV pressures. One subset of investigators who were not informed of the experimental group assignments performed the in vivo LV function studies.

**Infarct Size**

To measure infarct size 28 days after MI, the heart was excised and the LVs were cut from apex to base into 3 transverse sections. Five-micron sections were cut and stained with Masson's trichrome. Infarct length was measured along the endocardial and epicardial surfaces in each of the cardiac sections, and the values from all specimens were summed. Infarct size (as a percentage) was calculated as total infarct circumference divided by total cardiac circumference.12

In addition, to measure infarct size after 24 hours (when most animals were still alive), a separate group of animals including WT+MI (n=5) and TG+MI (n=5) mice was created by ligating the left coronary artery according to the same methods described earlier. After 24 hours of coronary artery ligation, Evans blue dye (1%) was perfused into the aorta and coronary arteries, and tissue sections were weighed and then incubated with a 1.5% triphenyltetrazolium chloride solution. The infarct area (pale), the area at risk (not blue), and the total LV area from each section were measured.13 In our preliminary study, we confirmed excellent reliability of infarct size measurements, in which a morphometric method similar to that performed in this study was used. The intrare observer and interob-
server variabilities between 2 measurements divided by these means, expressed as a percentage, were each <5%.

Myocardial Histopathology and Apoptosis
Myocyte cross-sectional area and collagen volume fraction were determined by quantitative morphometry of tissue sections from the mid-LV. To detect apoptosis, tissue sections from the mid-LV were stained with terminal deoxynucleotidyl transferase–mediated dUTP nick end-labeling (TUNEL) staining. The number of TUNEL-positive cardiac myocyte nuclei was counted, and the data were normalized per 10^5 total nuclei identified by hematoxylin-positive staining in the same sections. The proportion of apoptotic cells was counted in the noninfarcted LV. We further examined whether apoptosis was present by the more sensitive ligation-mediated PCR fragmentation assays (Maxim Biotech, Inc, Rockville, Md).

**mtDNA Copy Number**
DNA was extracted from cardiac tissues, and a Southern blot analysis was performed to measure the mtDNA copy number, as described earlier. Primers for the mtDNA probe corresponded to nucleotides 2424 to 3605 of the mouse mitochondrial genome, and those for the nuclear-encoded mouse 18S rRNA probe corresponded to nucleotides 435 to 1951 of the human 18S rRNA genome. The mtDNA levels were normalized to the abundance of the 18S rRNA gene run on the same gel.

Mitochondrial Complex Enzyme Activity
The specific activity of mitochondrial electron transport chain complex I (rotenone-sensitive NADH-ubiquinone oxidoreductase), complex II (succinate-ubiquinone oxidoreductase), complex III (ubiquinol–cytochrome c oxidoreductase), and complex IV (cytochrome c oxidase) was measured in myocardial tissues according to methods described previously. All enzymatic activities were expressed as nanomoles per minute per milligram protein.

**Plasma TBARS**
The formation of TBARS in peripheral blood samples from WT+MI and TG+MI mice was measured by a fluorometric assay, as described previously. In brief, 100 μL of whole blood was mixed with 1 mL of saline and centrifuged at 3000 g for 15 minutes. The supernatant was mixed with 1/50 H_2SO_4 and 10% phosphogluconic acid, and the mixture was centrifuged. The sediment was suspended in distilled water, 0.3% thiobarbituric acid, and 0.1% butylated hydroxytoluene. The reaction mixture was then heated at 100°C for 60 minutes in an oil bath. After being cooled with tap water, the mixture was extracted with n-butanol and centrifuged at 1600 g for 15 minutes. The fluorescence intensity of the organic phase was measured by spectrophotoflurometry with a wavelength of 510-nm excitation and 550-nm emission. Malondialdehyde standards (Sigma-Aldrich, St. Louis, Mo) were included with each assay batch, and plasma TBARS were expressed as micromoles per gram of plasma protein in reference to these standards.

**Statistical Analysis**
Data are expressed as mean±SEM. Survival analysis was performed by the Kaplan-Meier method, and between-group differences in survival were tested by the log-rank test. A between-group comparison of means was performed by 1-way ANOVA, followed by t tests. The Bonferroni correction was applied for multiple comparisons of means. *P*<0.05 was considered statistically significant.

The authors had full access to the data and take full responsibility for their integrity. All authors have read and agreed to the manuscript as written.

**Results**
We investigated 4 groups of mice, WT+sham (n=15), TG+sham (n=14), WT+MI (n=39), and TG+MI (n=37), in the present study. A survival analysis was performed for all of these mice. Subsequent echocardiographic and hemodynamic measurements were performed in the 4-week survivors: 15 WT+sham, 14 TG+sham, 25 WT+MI, and 31 TG+MI mice. These measurements could not be accomplished in 4 WT+MI and 5 TG+MI mice owing to technical difficulties. Survivor mice were further divided into 2 groups: those studied for subsequent histological analysis, including infarct size, myocyte size, and collagen volume fraction measurements as well as TUNEL staining (5 WT+sham, 5 TG+sham, 8 WT+MI, and 8 TG+MI), and those for the biochemical assay, including antioxidant enzyme activity, Prx-3 protein levels, mitochondrial lipid peroxidation, mtDNA copy number, and mitochondrial complex enzyme activities (8 WT+sham, 8 TG+sham, 8 WT+MI, and 8 TG+MI). Infarct size was not measured in the mice that died.

**Myocardial Antioxidants and TBARS**
First, baseline differences in Prx-3 proteins as well as other antioxidant enzyme activities between WT and TG mice were determined. In TG+sham, there was a significant increase in Prx-3 protein levels in the LV compared with that of WT+sham (Figure 1A). Importantly, the antioxidants, in-
Survival
There were no deaths in the sham-operated groups. Early operative mortality (within 6 hours) was comparable between WT+MI and TG+MI animals (15% versus 7%; P=NS). The survival rate up to 4 weeks tended to be higher in TG+MI compared with WT+MI mice, but this difference did not reach statistical significance (P=0.06 by log-rank test; Figure 3A). Death was suspected to be attributable to heart failure and/or arrhythmia. Five WT+MI (15%) and 2 TG+MI (5%) mice died of LV rupture (P=NS).

**Infarct Size**
Infarct size as determined by morphometric analysis 28 days after MI was comparable (55±1% versus 56±1%; P=0.83) between WT+MI (n=6) and TG+MI (n=6) groups. To further confirm that overexpression of Prx-3 did not alter infarct size, both the area at risk and infarct area were

**Figure 2.** Mitochondrial TBARS in 4 experimental groups of animals (n=8 each). Values are mean±SEM. *P<0.05 for difference from the WT+sham value. †P<0.05 for difference from the WT+MI value.

**TABLE 1.** Characteristics of Animal Models

<table>
<thead>
<tr>
<th></th>
<th>WT+Sham</th>
<th>TG+Sham</th>
<th>WT+MI</th>
<th>TG+MI</th>
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<tr>
<td><strong>Antioxidant enzymes</strong></td>
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<tr>
<td>n</td>
<td>7</td>
<td>7</td>
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<tr>
<td>SOD, U/mg protein</td>
<td>26.4±1.1</td>
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<td>25.1±1.7</td>
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<td>GSHPx, nmol/min per mg protein</td>
<td>74.1±3.2</td>
<td>77.7±6.7</td>
<td>87.8±4.8</td>
<td>86.1±4.2</td>
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<td>Catalase, nmol/mg protein</td>
<td>79.9±6.4</td>
<td>85.0±6.2</td>
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<td>81.4±5.8</td>
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<td><strong>Echocardiographic data</strong></td>
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<tr>
<td>n</td>
<td>15</td>
<td>14</td>
<td>21</td>
<td>26</td>
</tr>
<tr>
<td>Heart rate, bpm</td>
<td>481±11</td>
<td>451±8</td>
<td>463±13</td>
<td>458±8</td>
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<td>LVESD, mm</td>
<td>3.47±0.05</td>
<td>3.37±0.08</td>
<td>5.51±0.13†</td>
<td>4.9±0.10†§</td>
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<td>Fractional shortening, %</td>
<td>35.3±0.8</td>
<td>37.0±1.1</td>
<td>13.1±0.6†</td>
<td>16.9±0.6†§</td>
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<td><strong>Hemodynamic data</strong></td>
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<td>n</td>
<td>15</td>
<td>14</td>
<td>21</td>
<td>26</td>
</tr>
<tr>
<td>Heart rate, bpm</td>
<td>447±14</td>
<td>455±14</td>
<td>453±9</td>
<td>466±7</td>
</tr>
<tr>
<td>Mean aortic pressure, mm Hg</td>
<td>83±3</td>
<td>78±2</td>
<td>76±2</td>
<td>77±3</td>
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<tr>
<td>LVESD, mm</td>
<td>2.7±0.5</td>
<td>2.5±0.3</td>
<td>11.4±1.5†</td>
<td>7.6±1.0*‡</td>
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<td><strong>Organ weight data</strong></td>
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<tr>
<td>n</td>
<td>15</td>
<td>14</td>
<td>21</td>
<td>26</td>
</tr>
<tr>
<td>Body wt, g</td>
<td>26.9±0.5</td>
<td>27.0±0.8</td>
<td>27.0±0.3</td>
<td>26.4±0.4</td>
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<td>LV wt/body wt, mg/g</td>
<td>3.2±0.1</td>
<td>3.0±0.1</td>
<td>4.6±0.3†</td>
<td>4.4±0.1†</td>
</tr>
<tr>
<td>Lung wt/body wt, mg/g</td>
<td>5.0±0.1</td>
<td>5.2±0.1</td>
<td>7.6±0.5†</td>
<td>6.4±0.3†‡</td>
</tr>
<tr>
<td>Pleural effusion, %</td>
<td>0</td>
<td>0</td>
<td>43</td>
<td>15†</td>
</tr>
</tbody>
</table>

EDD indicates end-diastolic diameter; ESD, end-systolic diameter; and wt, weight. Values are mean±SEM.

*P<0.05, †P<0.01 vs WT+Sham. ‡P<0.05, §P<0.01 vs WT+MI.
measured in mice 24 hours after coronary artery ligation. Percentages of the LV at risk (risk area/LV, 51±3% versus 52±2%; \(P=0.89\)) and infarct size (infarct/risk area, 79±1% versus 78±1%; \(P=0.13\)) were also comparable between WT (n=5) and TG (n=5) animals.

### Echocardiography and Hemodynamics

The echocardiographic and hemodynamic data of surviving mice at 28 days are shown in Figure 3B and Table 1. LV diameters were significantly larger in WT MI mice at 28 days compared with sham-operated animals. TG MI mice displayed less attenuation in TG MI mice but was significantly attenuated in TG MI mice.

### Organ Weights and Histomorphometry

Coincident with an increased LVEDP, lung weight/body weight was larger in WT+MI mice, and this increase was attenuated in TG+MI mice (Table 1). The prevalence of pleural effusion was also lower in TG+MI than in WT+MI groups. Histomorphometric analysis of noninfarcted LV sections showed that myocyte cross-sectional area was greater in WT+MI mice but was significantly attenuated in TG+MI mice (Figure 4). Collagen volume fraction was greater in WT+MI mice, but this change was inhibited in TG+MI mice.

### Myocardial Apoptosis

There were rare TUNEL-positive nuclei in sham-operated mice. The number of TUNEL-positive myocytes in the noninfarcted LV was larger in WT+MI mice but was significantly smaller in TG+MI animals (Figure 5A). In addition, the intensity of the DNA ladder indicated that apoptosis in TG+MI animals was decreased compared with that in WT+MI mice (Figure 5B).

### mtDNA and Mitochondrial Complex Enzymes Activity

Consistent with our previous studies, \(^4\) mtDNA copy number in the noninfarcted LV from WT+MI animals showed a 36% decrease (\(P<0.05\)) compared with that in sham-operated mice, which was significantly prevented and was preserved at normal levels in TG+MI animals (Figure 6). To determine the effects of mtDNA alterations on mitochondrial function, we next measured the mitochondrial electron transport chain complex enzyme activities. The enzymatic activities of complexes I, III, and IV were significantly lower in the noninfarcted LV from WT+MI than in those from WT+sham animals (Table 2). Most important, no such decrease was observed in TG+MI mice. The enzymatic activity of complex II, exclusively encoded by nuclear DNA, was not altered in either group. These results indicate that mtDNA copy number and mitochondrial complex enzymatic activities are downregulated in the post-MI heart and that Prx-3 gene overexpression efficiently counteracts these mitochondrial deficiencies.
intramitochondrial Mn-SOD. Although Mn-SOD relieves matrix. ·O₂ mitochondrial is vectorially released into the mitochondrial Fe-S centers of complex enzymes. In addition, ·O₂ failing myocardium. 1 Most of ·O₂ but occurred secondary to more adaptive remodeling. Dysfunction. They were not due to its MI size-sparing effect but occurred secondary to more adaptive remodeling.

Dysfunction. They were not due to its MI size-sparing effect but occurred secondary to more adaptive remodeling.

Prx-5 was formerly known as SP-22, or MER5, is currently identified as a mitochondrial member of the novel antioxidant proteins designated as Prxs. Among 6 known mammalian Prxs, Prx-1 to -4 require the small redox protein thioredoxin (Trx) as an electron donor to remove H₂O₂, whereas Prx-5 and -6 can use other cellular reductants, such as GSH, as their electron donor. Prx-1, -2, and -6 are found in the cytoplasm and nucleus, whereas Prx-3 contains a mitochondrial localization sequence, is found exclusively in the mitochondria, and uses mitochondrial Trx-2 as the electron donor for its peroxidase activity. It functions not only by removing H₂O₂ formed after the SOD-catalyzed dismutation reaction but also by detoxifying peroxynitrite. Therefore, the great efficiency of Prx-3 as an antioxidant shown in the present study may be attributable to the fact that it is located in the mitochondria and can utilize lipid peroxides as well as H₂O₂ for substrates. In fact, overexpression of Prx-3 has been shown to protect thymoma cells from apoptosis induced by hypoxia, a bolus of peroxide, or an anticancer drug. Moreover, Prx-3 overexpression has been reported to protect rat hippocampal neurons from excitotoxic injury. Prx-5 is also associated with the mitochondria in addition to the peroxisomes and nucleus. Recently, increased expression of Prx-5 was found to have protected Chinese hamster ovary cells from mtDNA damage induced by oxidative stress. Therefore, Prx-5 may also exert beneficial effects against mitochondrial oxidative stress in post-MI hearts.

**TABLE 2. Mitochondrial Complex Enzyme Activities**

<table>
<thead>
<tr>
<th>Complex</th>
<th>WT + Sham</th>
<th>TG + Sham</th>
<th>WT + MI</th>
<th>TG + MI</th>
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<tr>
<td>Complex I, nmol/min per mg protein</td>
<td>282±26</td>
<td>265±38</td>
<td>159±25*</td>
<td>287±16†</td>
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<tr>
<td>Complex II, nmol/min per mg protein</td>
<td>770±70</td>
<td>718±93</td>
<td>711±85</td>
<td>726±128</td>
</tr>
<tr>
<td>Complex III, nmol/min per mg protein</td>
<td>505±11</td>
<td>470±31</td>
<td>367±20*</td>
<td>451±21†</td>
</tr>
<tr>
<td>Complex IV, nmol/min per mg protein</td>
<td>1223±37</td>
<td>1175±34</td>
<td>744±68*</td>
<td>939±54†</td>
</tr>
</tbody>
</table>

Values are mean±SEM.

*P<0.05 vs WT + sham; †P<0.05 vs WT + MI.
inhibited LV remodeling and failure after MI. However, GSHPx is located predominantly in the cytosol, and only a small proportion (≈10%) is present in the mitochondria. Therefore, it remains unclear whether the beneficial effects of GSHPx overexpression on post-MI hearts were attributable to an increase of this enzyme in the cytosol, the mitochondria, or both. The specific localization of Prx-3 in the mitochondria suggests that mitochondrial oxidative stress plays an important role in the development and progression of heart failure, and antioxidants localized specifically within the mitochondria provide a primary line of defense against this disease process.

A growing body of evidence suggests that ROS play a major role in the pathogenesis of cardiac failure. Furthermore, antioxidants have been shown to exert protective and beneficial effects against heart failure. A previous study from our laboratory demonstrated that dimethylthiourea improved survival and prevented LV remodeling and failure after MI. However, the most effective way to evaluate the contribution of any specific antioxidant and obtain direct evidence of an adverse role for ROS in heart failure is through gene manipulation. Therefore, the present study not only extends the previous observation that used antioxidants but also reveals the major role of mitochondrial oxidative stress in the pathophysiology of post-MI remodeling and failure.

The beneficial effects of Prx-3 overexpression shown in the present study were not due to its MI size-sparing effect, because there was no statistically significant difference in infarct size between WT + MI and TG + MI mice. Furthermore, its effects were not attributable to hemodynamics because blood pressures and heart rates were not altered (Table 1). Importantly, it is also unlikely that these effects were caused by the altered expression of antioxidant enzymes other than Prx-3 (Table 1). Moreover, the beneficial effects of Prx-3 overexpression were not due to systemic induction of antioxidant defenses. This possibility is less likely because plasma TBARS were comparable between WT + MI and TG + MI mice. Nevertheless, we cannot completely exclude the possibility that the systemic effects of Prx-3 induction might also have contributed to this phenotype because this TG is not heart-specific.

There may be several factors contributing to the protective effects conferred by Prx-3 overexpression on post-MI remodeling and failure. First, recent studies have demonstrated that a Trx-related antioxidant system is closely associated with the regulation of apoptosis, probably through quenching of ROS and redox control of the mitochondrial permeability transition pores that release cytochrome c. A subtle increase in ROS caused by partial inhibition of SOD results in apoptosis in isolated cardiac myocytes. Previous studies have demonstrated that apoptosis appears not only in infarcted but also in noninfarcted myocardium after MI. Specifically, apoptosis occurs in the noninfarcted LV late after MI. This is an intriguing observation, in light of the remodeling process known to occur within the noninfarcted area, which is characterized by the loss of myocytes and hypertrophy. In fact, recent studies have suggested cardiac myocyte apoptosis contributes to LV remodeling after MI. Importantly, increased oxidative stress occurs concomitantly with increased cardiac myocyte apoptosis within the noninfarcted area. This is a provoking observation, because oxidative stress is a powerful inducer of apoptotic cell death. The present study suggests that the coexistence of oxidative stress and myocyte apoptosis in the noninfarcted LV after MI is causally related. Oxidative stress may mediate myocyte apoptosis, which may lead to myocardial remodeling and failure. Therefore, the decreased mitochondrial oxidative stress due to Prx-3 overexpression could contribute to the amelioration of apoptosis (Figure 5) and eventual post-MI cardiac failure. Second, Prx-3 overexpression prevented the decrease in mtDNA copy number (Figure 6) as well as mitochondrial complex enzyme activities (Table 2). Our previous studies have demonstrated an intimate link between mtDNA damage, increased lipid peroxidation, and a decrease in mitochondrial function, which might play a major role in the development and progression of cardiac failure.

There are several issues to be acknowledged as limitations of this study. First, the differences between WT + MI and TG + MI groups in their echocardiographic measurements are not remarkable, even though they are statistically significant (Table 1). However, our previous study showed that the intraobserver and interobserver variabilities in our echocardiographic measurements for LV dimensions were small, and measurements made in the same animals on separate days were highly reproducible. Therefore, these values are considered to be valid. Second, longer-term follow-up data are not available for the animals in the current study. We therefore could not determine whether the differences between WT + MI and TG + MI groups seen in the present study were more or less significant at later time points, when additional LV remodeling would have been expected to occur.

In conclusion, Prx-3 overexpression inhibited the development of LV remodeling and failure after MI, which was associated with an attenuation of myocyte hypertrophy, apoptosis, and interstitial fibrosis. It also ameliorated mitochondrial oxidative stress as well as mtDNA decline and mitochondrial dysfunction in post-MI hearts. Therapies designed to interfere with mitochondrial oxidative stress could be beneficial to prevent heart failure after MI.

Acknowledgments

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Disclosures

None.

References


8. Hattori F, Murayama N, Noshita T, Oikawa S. Mitochondrial peroxiredoxin-3 might also beneficial in preventing clinical heart failure. Extends previous investigations that used antioxidants but also reveals a major role for mitochondrial oxidative stress in the pathogenesis of heart failure. Furthermore, antioxidants have been shown to exert protective and beneficial effects against this process. Recent studies have suggested that mitochondria are the predominant source of ROS in the failing heart, and mitochondrial antioxidants are expected to be the first line of defense against mitochondrial oxidative stress–mediated myocardial injury. The present study demonstrated that overexpression of peroxiredoxin-3 (Prx-3) inhibited cardiac remodeling and failure after myocardial infarction (MI) created by ligation of the left coronary artery in mice. Prx-3 contains a mitochondrial localization sequence, is found exclusively in the mitochondria, and uses mitochondrial thioredoxin (Trx)-2 as the electron donor for its peroxidase activity. It functions not only by removing H₂O₂ formed after the superoxide dismutase (SOD)–catalyzed dismutation reaction but also by detoxifying peroxynitrite. The great efficiency of Prx-3 as an antioxidant shown in the present study may be attributable to the fact that it is located in the mitochondria and can utilize lipid peroxides as well as H₂O₂ for substrates. The present study not only extends previous investigations that used antioxidants but also reveals a major role for mitochondrial oxidative stress in the pathophysiology of postinfarct heart failure. Therapies designed to interfere with mitochondrial oxidative stress by using antioxidant Prx-3 might also beneficial in preventing clinical heart failure.


Overexpression of Mitochondrial Peroxiredoxin-3 Prevents Left Ventricular Remodeling and Failure After Myocardial Infarction in Mice

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