Deficiency of the 5-Hydroxytryptamine Transporter Gene Leads to Cardiac Fibrosis and Valvulopathy in Mice

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Background—Serotonin (5-hydroxytryptamine; 5-HT) overproduction is responsible for cardiac valvular disease in patients with carcinoid tumors. Reduced 5-HT inactivation is one proposed mechanism of the valvulopathy observed in individuals treated with the appetite suppressants fenfluramine and phentermine. One key protein limiting systemic availability of 5-HT is the 5-HT transporter (5-HTT) expressed by platelets and pulmonary vascular cells; 5-HTT is responsible for 5-HT uptake and subsequent inactivation of the amine passing through the lung. Here we investigated whether 5-HTT–deficient (5-HTT-KO) mice developed structural and/or functional cardiac abnormalities and valvulopathy.

Methods and Results—Cardiac endothelial cells expressed large amounts of 5-HTT in wild-type mice. 5-HTT deficiency appeared to be associated with marked interstitial, perivascular, and valvular fibrosis as evidenced by staining of cardiac collagen in 5-HTT-KO mice. Histological analysis provided evidence for valvulopathy characterized by valvular hyperplasia and prominent fibrosis at the attachment site and base of the leaflets. Echocardiography revealed an increase in left ventricular lumen diameter and a decrease in left ventricular diameter fractional shortening. Although 5-HT1B receptors mediated the 5-HT–induced collagen secretion by human cardiac myofibroblasts, the contribution of this receptor type to valvulopathy was ruled out because double-KO mice deficient in both 5-HTT and 5-HT1B receptors showed the same cardiac alterations as 5-HTT-KO mice.

Conclusions—The present results establish a link between 5-HTT and the development of cardiac fibrosis and valvulopathy in vivo. 5-HTT-KO mice represent an especially relevant model for studying the mechanisms by which 5-HT induces valvulopathy. (Circulation. 2006;113:81-89.)

Key Words: heart diseases ■ pathology ■ valves ■ fibrosis ■ serotonin

Serotonin (5-hydroxytryptamine; 5-HT) controls a wide range of biological functions. In the brain, its role as a neurotransmitter has been abundantly documented. At the periphery, 5-HT originating from the gastrointestinal tract is stored in blood platelets and participates in coagulation and blood pressure homeostasis. 5-HT has been shown to play an important role in various cardiovascular disorders, most notably in the pathogenesis of valvular heart disease. The carcinoid syndrome, which is caused by release of 5-HT and other vasoactive substances from the tumor, results in endocardial damage followed by right-sided valvular dysfunction.1–3 Long-term treatment with drugs acting on 5-HT receptors, such as ergotamine or methysergide, is also known to induce valvular changes similar to those observed in carcinoid heart disease.4

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partially removed from the circulation by the liver and stored in platelets, but that complete removal of free circulating 5-HT is normally achieved by the lung, thus sparing the left-sided valves from exposure to high 5-HT levels under basal physiological conditions. 5-HT therefore appears as the most likely candidate mediator in the development of valvular heart disease associated with fenfluramine/phentermine treatment. Recent experimental studies showing that long-term 5-HT administration induces heart valve disease in rats is consistent with this hypothesis.6

One key protein involved in 5-HT clearance is the membrane-bound 5-HT transporter (5-HTT), which is responsible for cellular internalization of the bioamine.7,8 5-HTT is highly expressed in platelets and in the lung, notably in pulmonary artery endothelial and smooth muscle cells.9,10 It is presently unknown whether lung and/or platelet 5-HTT activity exerts a critical protective role against the potential deleterious effects of circulating 5-HT on the heart. Although treatment with selective 5-HTT inhibitors does not appear to be associated with valvular heart disease,11 it is not known whether 5-HTT deficiency reproduces some of the cardiac or valvular alterations observed in patients with carcinoid syndrome and in individuals undergoing long-term fenfluramine/phentermine treatment. In the present study, we reasoned that if 5-HTT were directly involved in the pathogenesis of cardiac fibrosis, mice with targeted disruption of the 5-HTT gene (5-HTT-KO)12,13 would be prone to develop structural and functional cardiac abnormalities. We therefore performed hemodynamic and histological investigations aimed at assessing whether abnormal fibrosis and functional alterations occur in the hearts of 5-HTT–deficient mice. Because one of the potential mechanisms of 5-HT–induced valvular heart disease may involve 5-HT1b receptors, similar investigations were also performed in 5-HT1b receptor–deficient mice (5-HT1b-KO) and in double mutants devoid of both 5-HTT and 5-HT1b receptors (5-HTT-KO×5-HT1b-KO). In addition, in vitro experiments with human cardiac myofibroblasts allowed direct evaluation of 5-HT1b receptor involvement in collagen synthesis associated with fibrosis.

Methods

Animals
Mice lacking 5-HTT (5-HTT-KO)12 and the 5-HT1b receptor (5-HT1b-KO)13 were generated by homologous recombination on a mixed genetic background (129/Sv×C57BL/6). Double 5-HTT-KO×5-HT1b-KO mutants were obtained by crossing 5-HT1b-KO with 5-HTT-KO heterozygous mice. Pups were typed by Southern blot analysis of tail biopsy samples as described previously.13,14 The anatomic, histological, and functional studies were performed on 8- to 10-week-old male mice. All animal care and procedures were in accordance with institutional guidelines.

Detection of mRNAs Encoding Tryptophan Hydroxylases, 5-HT Receptors, and the 5-HTT in the Mouse Heart
RNA was extracted from the entire heart with Trizol reagent (Gibco BRL). Total RNA was systemically treated with DNase. RNA concentration and quality were determined by agarose gel electrophoresis and spectrophotometry.13 mRNAs encoding 5-HT receptors, 5-HTT, and tryptophan hydroxylases 1 (Tph1) and 2 (Tph2) were detected by reverse transcription followed by polymerase chain reaction (RT-PCR). In brief, 1 μg of total RNA was reverse-transcribed (45 minutes at 48°C) and amplified with use of the access RT-PCR kit (Promega) with specific primers (Table 1) in the presence of 2.5 mmol/L MgCl2. Cycle amplifications were performed at 94°C, 54°C, and 72°C (1 minute each, 28 cycles). PCR products were electrophoresed in 1% agarose gels stained with ethidium bromide.

Cardiac 5-HTT Immunoblotting
Immediately after removal, the hearts were quickly frozen in LN2 and stored at −80°C. After being thawed at 0°C, tissues were sonicated in 0.1 mmol/L phosphate-buffered saline containing anti-proteases (1 μmol/L leupeptin and 1 μmol/L pepstatin A). Homogenates were subjected to sodium dodecyl sulfate–polyacrylamide gel electrophoresis, and proteins in the gel were transferred to a nitrocellulose membrane by electroblotting with use of a transblot Bio-Rad transfer apparatus (Marnes La Coquette). The 5-HTT protein was detected by incubating the nitrocellulose membrane with goat polyclonal anti–5-HTT antibodies (Santa Cruz Biotechnology) at 1:1000 dilution, as described in detail elsewhere.12

Immunohistochemical Labeling
Paraffin sections (5 μm thick) were mounted on Superfrost Plus slides (Fisher Scientific). 5-HTT immunostaining was performed with the same polyclonal anti–5-HTT antibodies as those used for immunoblotting experiments but diluted to 1:2000, as previously described.12

Hemodynamic Measurements
Mice aged 8 to 10 weeks were anesthetized with ketamine (60 mg/kg IP) and xylazine (10 mg/kg IP). The right carotid artery was exposed and a polyethylene catheter (PE10) was inserted. Arterial pressure was measured with a Gould P10 EZ pressure transducer connected to pressure modules and a Gould TA 550 recorder. The heart rate under these conditions was between 300 and 500 bpm. If the heart rate fell to <300 bpm, measurements were excluded from analysis (this happened twice). Finally, animals were deeply anesthetized with sodium pentobarbital (40 mg/kg IP) and exsanguinated. The thorax was opened and the heart was removed and weighed.

Echocardiography
Mice were anesthetized with ketamine (60 mg/kg IP) and xylazine (10 mg/kg IP). The left side of the thorax was shaved. Echocardiograms were recorded with a Technos echo device (ESAOTE Biomedia) equipped with a 13.8-MHz probe. Echocardiography data (2D, M-mode, and Doppler) were collected on an optical disk and stored for off-line analysis. Left ventricle wall thickness and lumen diameters at end diastole and end systole were measured from the M-mode images, and fractional shortening was calculated. Five consecutive beats were analyzed in triplicate and averaged.

Histological Analysis and Assessment of Cardiac Fibrosis
The hearts were fixed in a bath of 4% aqueous buffered formalin for 1 week and processed for paraffin embedding. Longitudinal sections (5 μm thick) containing both right and left ventricles were stained with the collagen-specific Sirius red and Masson’s trichrome dye.16 Image analysis was performed with a charge-coupled device Iris camera (CDD Iris, Sony) coupled with a light microscope (Laborlux). Total collagen volume fraction (ie, the ratio of collagen surface area to total cardiac surface area, as a percentage) of the stained tissue, was determined separately in the valve area and myocardium with Perfect Image software (Clara Vision).

Culture of Human Cardiac Valve Myofibroblasts
Human aortic and mitral heart valves were collected from recipients during valve replacement surgery for nonrheumatic aortic stenosis or mitral insufficiency. Explants were cut into small pieces (3 to 5 mm), which were transferred to cell-culture flasks (Nunc). To allow the
myofibroblasts to grow out, the valve tissues were maintained in Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 15% fetal calf serum (FCS), 2 mmol/L glutamine, and antibiotics (100 U/mL penicillin and 0.1 mg/mL streptomycin). After 2 weeks, the cells collected in the culture medium and the valve tissues were transferred to new cell-culture flasks. Cell phenotype was determined with monoclonal antibodies against desmin, vimentin, and factor VIII. Cells positive for vimentin and negative for desmin and factor VIII were labeled as myofibroblasts. We found that 95% of cells were myofibroblasts in these experiments, whether the cells were derived from the mitral or the aortic valve. Cells were used for the study between passages 1 and 3.

Measurement of \([3H]\)Proline Incorporation

Myofibroblasts in DMEM supplemented with 15% FCS were seeded into 24-well plates at a density of 2.5\times10^4 cells/well and allowed to adhere. The cells were subjected to 48 hours of growth arrest in medium containing only 0.2% FCS and then incubated in DMEM supplemented with 0.2% FCS, 0.6 mmol/L ascorbic acid, 0.1 mmol/L iproniazid (a monoamine oxidase inhibitor), and 5\times10^{-6} mol/L \([3H]\)proline, with (10\times10^{-6} to 10\times10^{-8} mol/L) or without 5-HT. The effect of 5-HT was also examined in the presence of 10\times10^{-6} mol/L fluoxetine (a specific 5-HTT inhibitor), GR127935 (a 5-HT 1B/1D receptor antagonist), ketanserin (a 5-HT2A receptor antagonist), or SB206553 (a 5-HT2B/2C receptor antagonist). Each of these drugs was added 20 minutes before 5-HT. After incubation for 48 hours, the cells were washed with phosphate-buffered saline, treated with ice-cold 10% trichloroacetic acid, and dissolved in 0.3 N NaOH/0.1% sodium dodecyl sulfate. The incorporated radioactivity was counted. \([3H]\)proline incorporation is reported as counts per minute per well.

Blood 5-HT and 5-HIAA Concentrations

5-HT and its metabolite 5-hydroxyindoleacetic acid (5-HIAA) were determined in whole blood by high-performance liquid chromatography coupled to electrochemical detection, as described in detail elsewhere.18

Chemicals

\([3H]\)proline was from Amersham Pharmacia Biotech. Iproniazid, ascorbic acid, 5-HT, and SB206553 (N-3-pyridinyl-3,5-dihydro-5-methylbenzo[1,2-b:4,5-b']dipyrrolo-[1]-5-carboxamide hydrochloride) were from Sigma. The other compounds were fluoxetine (Eli Lilly), ketanserin (Janssen), and GR127935 {2-methyl-4’-(5-methyl-(1,2,4)oxadiazol-3-yl)-biphenyl-4-carboxylic acid [4-methoxy-3-(4-methyl-piperazine-1-yl)-phenyl]amide; GlaxoSmithKline}.

Statistical Analysis

All results are reported as mean±SEM. The nonparametric Mann-Whitney test was used to compare wild-type (WT) and 5-HTT-KO mice. The Kruskal-Wallis test was used to test the in vitro effects of 5-HT on fibroblasts and for comparisons among WT, 5-HT1B, and 5-HTT mutant mice. When the Kruskal-Wallis test showed a significant difference, the groups were further compared with a nonparametric Student-Newman-Keuls test. Probability values <0.05 were considered statistically significant.

### Table 1. Primers for RT-PCR Analysis of mRNAs Encoding Mouse 5-HT Receptors, 5-HTT, Tph1, and Tph2

<table>
<thead>
<tr>
<th>Receptor</th>
<th>Primer 1</th>
<th>Primer 2</th>
<th>Length, bp</th>
<th>GenBank Accession No.</th>
</tr>
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<tr>
<td>5-HT1A</td>
<td>TGCTATGCTGGTCTCTAT</td>
<td>TCTCAACACTCCCCTGC</td>
<td>118–1137</td>
<td>NM_008308</td>
</tr>
<tr>
<td>5-HT1B</td>
<td>tcggattttgaaacagacacc</td>
<td>tctcagcactgctgcctgc</td>
<td>699–719</td>
<td>NM_010482</td>
</tr>
<tr>
<td>5-HT1D</td>
<td>tcacccagtgaaaaatgaacg</td>
<td>tggttctttcttcagaggg</td>
<td>1783–1803</td>
<td>NM_008309</td>
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<tr>
<td>5-HT2A</td>
<td>AAGACTCGAAGCAATTGAGT</td>
<td>AGATTTTCAGGAAGGTCTTGTT</td>
<td>1215–1238</td>
<td>NM_172812</td>
</tr>
<tr>
<td>5-HT2B</td>
<td>CTCTTTTCAACTGCCCATC</td>
<td>TGTATTTCTAGATGCAGTACG</td>
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<td>NM_008311</td>
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<tr>
<td>5-HT2C</td>
<td>ATGGCTTATGTTGTGGAGGC</td>
<td>AAGAAAGCAGGAGAGAG</td>
<td>1759–1779</td>
<td>NM_008312</td>
</tr>
<tr>
<td>5-HT3A</td>
<td>GGGAGCTCCGGAACATTCTT</td>
<td>CCCCATATCTATCTACCA</td>
<td>458–478</td>
<td>NM_013561</td>
</tr>
<tr>
<td>5-HT4</td>
<td>taatgtgcttcacagggqg</td>
<td>actgctggctaggtcagcagc</td>
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<td>NM_008313</td>
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<tr>
<td>5-HT6</td>
<td>agttcgtctgacagtaagcgc</td>
<td>CAGCCAGTGCAAGAAAGAAT</td>
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<td>AF134158</td>
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<tr>
<td>5-HTT</td>
<td>ACATGTTGGGTTCCTTTCAT</td>
<td>TTTGACTCTCTCTTCAGATGCT</td>
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<td>NM_010484</td>
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<tr>
<td>TPH1</td>
<td>tgatgttttgcagctatcc</td>
<td>cgtgagcagctgaatatcttc</td>
<td>1775–1797</td>
<td>NM_009414</td>
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<tr>
<td>TPH2</td>
<td>CTACAGTACACTTGCTGACATGC</td>
<td>CTACAGTACACTTGCTGACATGC</td>
<td>1604–1627</td>
<td>NM_173391</td>
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</table>

\(\Delta\) indicates hybridization temperature.
Results

Expression of Tph, 5-HT Receptors, and 5-HTT in the Mouse Heart

RT-PCR analysis showed that mouse cardiac tissues contained mRNAs for Tph1, 5-HTT, and various 5-HT receptor types, including 5-HT1 (a, b, and d), 5-HT2 (a, b, and c), 5-HT3A, and 5-HT4 (Figure 1). In contrast, mRNAs encoding 5-HT6 receptor or Tph2 were not detected in the mouse heart. Significant amounts of Tph1 mRNAs were found in human valvular tissues (data not shown). 5-HTT expression in the heart was further confirmed by immunoblotting experiments, which showed high levels of 5-HTT protein in cardiac tissues from WT mice. Immunohistochemical analyses showed predominant localization of 5-HTT in the endocardium and endothelium of coronary arteries and capillaries (Figure 2a). In contrast, 5-HTT protein was not detected in tissues from 5-HTT-KO mice (Figure 2b) or in isolated cardiomyocytes from WT animals (data not shown).

Blood 5-HT and 5-HIAA Concentrations in 5-HTT-KO Versus WT Mice

Whole-blood 5-HT levels were dramatically reduced in 5-HTT-KO mice (n=6) compared with WT mice (n=6), from 29.25±1.10 to 0.35±0.02 nmol/mL (P=0.004), whereas blood 5-HIAA levels did not differ between the 2 groups (1.60±0.30 nmol/mL in 5-HTT-KO mice and 1.15±0.20 nmol/mL in WT mice; n=5 in each group).

Anatomic and Hemodynamic Characteristics of the Heart in 5-HTT-KO Versus WT Mice

As shown in Table 2, 5-HTT-KO and WT mice did not differ with regard to body weight, heart weight, heart weight to body weight ratio, systemic arterial pressure, or heart rate. Moreover, cardiac function evaluated by noninvasive transthoracic Doppler echocardiography revealed both dilated cardiopathy (increased left ventricular lumen diameter) and myocardial hypokinesis (decreased fractional shortening) in 5-HTT-KO mutants compared with paired WT mice (Table 3).

Histological Evidence of Cardiac Fibrosis in 5-HTT-KO Versus WT Mice

Histological analysis showed that 5-HTT-KO mice developed cardiac fibrosis compared with WT mice. Collagen staining predominated in valvular tissues (Figure 3a and 3d) but was also seen in interstitial (Figure 3b and 3e) and perivascular (Figure 3c and 3f) regions in both ventricles. Quantification of collagen staining revealed significantly more collagen in both myocardial and valvular regions in 5-HTT-KO mice compared with littermate controls (Figure 4). Myocardial fibrotic lesions did not appear to affect sections diffusely but instead seemed to occur focally within the myocardium.
valvular regions, histological examination demonstrated leaflet thickening and marked collagen accumulation at the attachment site and in the leaflet itself. These lesions were homogeneously distributed in the mitral, tricuspid, aortic, and pulmonary valves, as illustrated by Figure 5c, showing similar collagen staining of the mitral and aortic valves in the same cardiac section. Interestingly, several areas of chondroid metaplasia were identified within the valvular tissue in 4 of 6 5-HTT-KO mice; these areas were more prominent at the attachment site and base of the leaflets (Figure 5e).

5-HT1B Receptor–Mediated Increase in [3H]Proline Incorporation in Human Cardiac Valve Myofibroblasts

In myofibroblasts cultured in serum-free medium, 5-HT produced a concentration-dependent increase in [3H]proline incorporation. Pretreatment of the cells with GR127935 (10^{-6} mol/L), a 5-HT1B receptor antagonist, significantly inhibited the effect of 5-HT (Figure 6). In contrast, the 5-HT–induced increase in [3H]proline incorporation was not affected by cell exposure to 10^{-7} mol/L of the 5-HT2A and 5-HT2C receptor antagonists ketanserin and SB206553, respectively. Similarly, the effect of 5-HT was unchanged by cell treatment with the 5-HTT inhibitor fluoxetine (Figure 6).

Anatomic, Hemodynamic, and Histological Characteristics of the Heart in 5-HT1B-KO and Double 5-HTT-KO×5-HT1B-KO Mice Compared With WT Mice

In contrast to 5-HTT-KO mutants, 5-HT1B-KO mice were not significantly different from the relevant WT controls with regard to anatomic, histological, and functional characteristics of the heart (data not shown). However, double 5-HTT-KO×5-HT1B-KO mutants exhibited marked alterations in cardiac parameters. Thus, echocardiography revealed similar changes in left ventricular lumen diameter and fractional shortening in double mutants as in 5-HTT-KO mice. Moreover, the extent of cardiac fibrosis was similar in these 2 groups (Figure 7).

**Discussion**

The present results show that 5-HTT gene deficiency in 5-HTT-KO mice led to the development of cardiac myocar- dial and valvular fibrosis, together with left ventricular dysfunction and dilation, supporting a role for 5-HT in the induction of valvular heart disease. Thus, 5-HTT–deficient mice seem highly relevant for studying the mechanisms by which 5-HT may induce valvulopathy. Although 5-HT1B receptors were found to mediate the stimulatory effect of 5-HT on cardiac fibroblast-induced collagen production, double-mutant mice deficient in 5-HTT– and 5-HT1B–encoding genes exhibited the same cardiac alterations as 5-HTT-KO mice, indicating that 5-HT1B receptors do not contribute to 5-HT–induced cardiac fibrosis.

The potential role of 5-HT in cardiac function, as well as its involvement in various cardiovascular disorders, has generated considerable interest in recent years. Thus, disruption of the nonneuronal Tph1 gene encoding tryptophan hydroxylase, the rate-limiting enzyme of 5-HT synthesis in peripheral tissues, was recently shown to induce cardiomyopathy, demonstrating the importance of peripheral 5-HT for normal heart development and function.19 As shown in the present study, several 5-HT receptor types are expressed by cardiac cells, suggesting that 5-HT may exert various direct effects on the heart. Although 5-HT_{1A} receptors have been implicated in the control of cardiac electrical activity,20 previous studies in mice lacking or overexpressing 5-HT_{1B} receptors revealed important roles for this receptor in heart morphogenesis and cardiomyocyte hypertrophy.21 However, the roles for other 5-HT receptor types expressed by the mouse heart, including 5-HT_{1A}, 5-HT_{1B}, 5-HT_{1D}, 5-HT_{2A}, and 5-HT_{2C}, remain to be determined. In addition, we found that cardiac tissues expressed large amounts of 5-HTT. Although 5-HTT has been shown to be expressed by fetal cardiac myocytes and to mediate their proliferation,22 we found, in accordance with other studies,22 that 5-HTT was not expressed by adult cardiomyocytes. This suggests a transient effect of 5-HTT on cardiomyocytes confined to the developmental period. In adult WT mice, cardiac 5-HTT expression predominated in vascular and endocardial endothelial cells, suggesting a high capacity of these cells to internalize 5-HT and presumably to achieve its degradation. This observation therefore suggests that, in addition to pulmonary vascular cells, cardiac endothelial cells may be well equipped to clear 5-HT from the bloodstream, thereby controlling the potential cardiac effects of indoleamine released at remote peripheral sites.

We used 5-HTT-KO mice, which have been well characterized in terms of locomotor activity and behavior.13 As reported previously, we found that blood 5-HT concentrations were dramatically reduced in the mutants compared with WT mice, an expected finding, because platelet 5-HT is known to

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**TABLE 2. Anatomic and Hemodynamic Parameters in WT and 5-HTT KO Mice**

<table>
<thead>
<tr>
<th></th>
<th>WT (n=6)</th>
<th>5-HTT KO (n=5)</th>
<th>P</th>
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</thead>
<tbody>
<tr>
<td>Body weight, g</td>
<td>33.8±0.6</td>
<td>34.4±1.3</td>
<td>0.60</td>
</tr>
<tr>
<td>Heart weight, mg</td>
<td>173.7±13.6</td>
<td>164.0±11.0</td>
<td>0.92</td>
</tr>
<tr>
<td>Heart weight/body wt, mg/g</td>
<td>5.1±0.3</td>
<td>4.8±0.3</td>
<td>0.75</td>
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<tr>
<td>Systolic arterial pressure, mm Hg</td>
<td>105±8</td>
<td>99±11</td>
<td>0.52</td>
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<tr>
<td>Diastolic arterial pressure, mm Hg</td>
<td>75±3</td>
<td>69±8</td>
<td>0.30</td>
</tr>
<tr>
<td>Mean arterial pressure, mm Hg</td>
<td>88±4</td>
<td>84±10</td>
<td>0.37</td>
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<tr>
<td>Heart rate, bpm</td>
<td>327±18</td>
<td>312±15</td>
<td>1.00</td>
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</table>

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**TABLE 3. Echocardiographic Parameters in WT and 5-HTT KO Mice**

<table>
<thead>
<tr>
<th></th>
<th>WT (n=5)</th>
<th>5-HTT-KO (n=6)</th>
<th>P</th>
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<tbody>
<tr>
<td>LVD-ED, mm</td>
<td>3.02±0.10</td>
<td>3.86±0.05</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>LVD-ES, mm</td>
<td>1.48±0.09</td>
<td>2.46±0.18</td>
<td>0.02</td>
</tr>
<tr>
<td>FS, %</td>
<td>51±2</td>
<td>39±4</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>IVS-ED, mm</td>
<td>0.80±0.03</td>
<td>0.90±0.09</td>
<td>0.34</td>
</tr>
<tr>
<td>PW-ED, mm</td>
<td>0.82±0.03</td>
<td>0.87±0.09</td>
<td>0.33</td>
</tr>
</tbody>
</table>

LVD indicates left ventricular lumen diameter; ED, end diastole; ES, end systole; IVS, interventricular septum thickness; PW, left ventricular posterior wall thickness; and FS, left ventricular diameter fractional shortening.

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contribute 99% of whole-blood 5-HT. In contrast, blood 5-HIAA levels did not differ between 5-HTT-KO and WT mice, arguing against adaptive alterations in 5-HT synthesis in the KO animals. Suppression of 5-HT uptake by platelets or vascular cells may have resulted in an increase in the plasma 5-HT concentration, which is usually in the nanomolar range and therefore difficult to assess, because platelet activation occurs during blood sampling. Our results showing Tph1 expression in mouse cardiac tissues and in human valvular tissues also suggest cardiac synthesis of 5-HT. A deficiency in 5-HTT gene activity may therefore result in more indoleamine available for binding to 5-HT receptors on cardiac tissues, either contained in plasma or produced locally within the heart.

In clinical disorders associated with increased 5-HT circulating levels such as carcinoid tumors, cardiac valve dysfunction is common. The pathological characteristics of carcinoid tumor–associated heart disease include diffuse collections of thick white plaques composed of myofibroblasts. These plaques are deposited on the endocardial surfaces of the right heart and valves, where increased extracellular matrix accumulation and increased thickness of the valve leaflets are also found. In 5-HTT-KO mice, we found fibrotic areas to be interstitial and perivascular, with evidence for valvulopathy characterized by hyperplastic valvular lesions. Although we did not observe plaque on the endocardium, the extent and location of the fibrotic lesions in 5-HTT-KO mice differed strikingly from those commonly observed in heart failure. Indeed, cardiac fibrosis as a marker of heart failure is mainly interstitial and homogeneously distributed throughout the myocardium. In 5-HTT–deficient mice, myocardial fibrosis was heterogeneously distributed and associated with marked valvular fibrosis. An interesting feature in these mutants was the presence of several areas of chondroid metaplasia within the valvular apparatus, which were prominent at the attachment site and base of the leaflets, suggesting a contribution of 5-HT to cell metaplasia and chondrocyte development within valvular tissues.

In 5-HTT-KO mice, fibrotic lesions were found in both the right and left parts of the heart, with no apparent predominance on the right side. It is likely that the left-sided lesions contributed to the development of left ventricular dysfunction, as reflected by the increase in left ventricular lumen diameter and the decrease in left ventricular diameter fractional shortening on echocardiography. In humans, such carcinoid involvement of left-sided heart valves has been reported in patients with patent foramen ovale, carcinoid tumor of the lung, or active carcinoid syndrome with high 5-HT levels. Similar findings, including bilateral cardiac involvement and closely related pathological changes, have been reported in patients exposed to fenfluramine or ergot alkaloids. Ergot alkaloids are considered nonselective 5-HT receptor agonists, and the phentermine/fenfluramine combination is believed to increase the availability of free 5-HT as a result of both increased 5-HT release from platelets and inhibition of pulmonary monoamine oxidase activity. Taken together, these observations have led to the suggestion that increased cardiac 5-HT delivery or pharmacological activation of cardiac 5-HT receptors was the mechanism underlying the development of valvulopathy in patients taking these medications. This issue has also been addressed in patients treated with selective 5-HT reuptake inhibitors, although no relation was found between use of those drugs and development of valvular heart disease. A direct relation between 5-HT overload and valvulopathy was recently demonstrated in an in vivo rat model. Both right- and left-sided heart lesions were observed, consisting of thickened valvular cusps and carcinoid-like plaques. The present results obtained in 5-HTT-KO mutants lacking lung, cardiac, and platelet 5-HTT

Figure 4. Myocardial and valvular fibrosis quantification in WT and in 5-HTT-KO mice. Each bar is the mean±SEM of 6 independent determinations (1 per mouse). *P<0.05 compared with corresponding values in WT mice.
provide further support for a direct link between 5-HT and the development of valvulopathy. In addition, they suggest that 5-HTT may play a key role in protecting the heart from the effects of circulating or locally produced 5-HT.

The potential mechanisms leading to the formation of 5-HT–induced carcinoid plaque are still a matter of debate. Although 5-HTT was proposed as a key protein because of its high affinity for fenfluramine derivatives, its involvement seems unlikely, given the present results showing development of cardiac fibrosis in the absence of this transporter. Among potentially involved 5-HT receptors, 5-HT1 and 5-HT2 receptor families are good candidates because (1) they are abundantly expressed by interstitial cells in human heart valves and (2) their stimulation leads to fibroblast hyperplasia. Special attention was given to 5-HT1B receptors, whose stimulation triggers fibroblast proliferation. Furthermore, this receptor type is detected in subendocardial cells, where its stimulation also leads to cell proliferation. In the present study, we used cultured human valvular fibroblasts to examine the potential role of these receptors in 5-HT–induced incorporation of proline as an index of collagen production. Interestingly, only GR127935, an antagonist of 5-HT1B/1D receptors, inhibited the effect of 5-HT, whereas selective blockade of either 5-HT2A or 5-HT2B receptors had no effect. However, double-KO mice (5-HTT-KO/5-HT1B-KO) deficient in both the transporter and the 5-HT1B receptor type did not differ from 5-HTT–deficient mice with regard to the extent of cardiac fibrosis. Thus, 5-HT1B receptors do not seem

Figure 5. Histological staining (Masson’s trichrome) of valvular fibrosis in a WT (a, b) and a 5-HTT-KO (c, d) mouse and chondroid metaplasia in the aortic valve apparatus of a 5-HTT-KO mouse (e). Scale bar=500 μm in a and c, 125 μm in b and d, and 25 μm in e.

Figure 6. [3H]proline incorporation into cultured human cardiac valve myofibroblasts in response to increasing concentrations of 5-HT (10^-8 to 10^-6 mol/L). The response to 10^-6 mol/L 5-HT was also measured in the presence of ketanserin (KT), SB206553 (SB), GR 127935 (GR), or fluoxetine (Fluox) at 10^-6 mol/L each. Values are mean±SEM of 6 independent experiments. *P<0.05, †P<0.01, compared with [3H]proline incorporation with no 5-HT added (0); ‡P<0.01 compared with 10^-6 mol/L 5-HT alone.
than in 5-HTT-WT, 5-HT1B-WT, and 5-HT1B-KO mice. Each bar

$P<0.05$ than in 5-HTT-WT, 5-HT1B-WT, and 5-HT1B-KO mice. Each bar

Figure 7. Valvular fibrosis quantification in 5-HTT-KO, 5-HT1B-

KO, and double-mutant 5-HTT-KO×5-HT1B-KO mice and the

respective WT controls with the same genetic background. Colla-

gen accumulation did not differ significantly between

5-HTT-KO and double-mutant mice. However, collagen accumu-

lation in these 2 groups was significantly greater ($P<0.05$)

in 5-HTT-KO, 5-HT1B-WT, and 5-HT1B-KO mice. Each bar

is the mean±SEM of 6 independent determinations (1 determi-

nation per mouse).

to play a role in cardiac fibrosis, at least in mice. Until now,

most of the studies were performed in vitro to evaluate the

response of fibroblasts or cardiac interstitial cells to 5-HT,

ergot-related drugs (ergotamine, methysergide), and fenflur-

amine derivatives. Depending on the studies, activation of

5-HT2A or 5-HT2B receptors was suggested as the potential

mechanism.25 Indeed, norfenfluramine, the fenfluramine

metabolite, has been shown to exhibit high affinity for

$\\text{5-HT}2\text{C}$, but not for $\\text{5-HT}2\text{A}$ receptors.25,26 In

contrast, studies of sheep valvular interstitial cells showed

that 5-HT acted mainly through $\\text{5-HT}2\text{A}$ receptors.27,28

Interestingly, $\\text{5-HT}2\text{A}$ receptor activation leads to increased

synthesis and function of transforming growth factor-\(\beta\),

suggesting that 5-HT may also have indirect fibrotic

effects mediated by transforming growth factor-\(\beta\). Because

the present studies were performed on cells from diseased

human valves, whereas the aforementioned studies investi-
gated cells from normal animal heart valves, we cannot

exclude the possibility that the action of 5-HT may vary

to the species or the presence of underlying

valve disease. It is also possible that no direct relation

exists between in vitro results obtained with cultured cells

and in vivo results in 5-HTT-KO mice. The data reported

herein suggest that the experimental model of 5-HTT-KO

mice may prove useful for identifying the 5-HT receptors

involved in the cardiac alterations caused by 5-HT or ergot

and fenfluramine derivatives and for elucidating the un-

derlying mechanisms.

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Disclosures

None.

References


1985;7(suppl 7):S79–S85.

2. Alegret JM, Subirana MT, Marcuello E, Bayes-Genis A, Ferres P, Torner

M. Tricuspid and pulmonary valve in a case of carcinoid syndrome. Rev


3. Robbiolio PA, Rigolin VH, Wilson JS, Harrison JK, Sanders LL, Bashore

TM, Feldman JM. Carcinoid heart disease: correlation of high 5-

hydroxytryptamine levels with valvular abnormalities detected by cardiac catheterization and echocardiography. Circulation.


4. Hendrix M, Van Dorpe J, Flameng W, Daenen W. Aortic and mitral

valve disease induced by ergotamine therapy for migraine: a case report


5. Gardin JM, Schumacher D, Constantine G, Davis KD, Leung C, Reid CL.

Valvular abnormalities and cardiovascular status following exposure to
dexfenfluramine or phentermine/fenfluramine. JAMA. 2000;283:

1703–1709.

6. Gustafsson BI, Tømmerås K, Nordrum I, Loennechen JP, Brunsvik A,

Solligård E, Fossmark R, Bakke I, Syversen I, Waldum H. Long-term

serotonin administration induces heart valve disease in rats. Circulation.


7. Lesch KP, Wolozin BL, Estler HC, Murphy DL, Rieder P. Isolation of a
cDNA encoding the human brain serotonin transporter. J Neural Transm


AS, Ganapathy V, Blakely RD. Antidepressant-and cocaine-sensitive

human serotonin transporter: molecular cloning, expression, and chro-


9. Lesch KP, Fanburg BL. Serotonin uptake by bovine pulmonary endothelial

cells in culture: stimulation by hypoxia. Am J Physiol. 1986;250:

C766–C770.

10. Eddahibi S, Hubert M, Fadel E, Raffestin B, Darmon M, Capron F,

Simonneau G, Darteville P, Hamon M, Adnot S. Serotonin transporter

overexpression is responsible for pulmonary artery smooth muscle hyper-

plasia in primary pulmonary hypertension. J Clin Invest. 2001;108:

1141–1150.


between selective serotonin-reuptake inhibitor therapy and heart valve


12. Eddahibi S, Hanoun N, Lantumey L, Lesch K, Raffestin B, Hamon M,

Adnot S. Attenuated hypoxic pulmonary hypertension in mice lacking the


1555–1562.


Mossner R, Westphal H, Lesch K. Altered brain serotonin homeostasis

and locomotor insensitivity to 3,4-methylenedioxymethamphetamine


Homozygous mice deficient in serotonin 5-HT1B receptor and antide-

pressant effect of selective serotonin reuptake inhibitors. C R Seances Soc


S. Induction of serotonin transporter by hypoxia in pulmonary vascular

smooth muscle cells: relationship with the mitogenic action of serotonin.


The reliability of the tracing-method of fine cardiac fibrosis at a magni-

fication of ×10: preliminary study for quantitative analysis of fibrosis in

large tissue sections of hearts with cardiomyopathy. Heart Vessels. 1985;


17. Barnes NM, Sharp T. A review of central 5-HT receptors and their


Alterations of central serotonin and dopamine turnover in rats treated with

ipsapirone and other 5-hydroxytryptamine1A agonists with potential


19. Cote F, Thévenot E, Fligny C, Fromes Y, Darmon M, Ripoche MA,


J, Vodjani G. Disruption of the nonneuronal tph1 gene demonstrates the

importance of peripheral serotonin in cardiac function. Proc Natl Acad


20. Pau D, Workman AJ, Kane KA, Rankin AC. Electrophysiological effects

of 5-hydroxytryptamine on isolated human atrial myocytes, and the


140:1434–1441.


Maroteaux L. Overexpression of the serotonin 5-HT2B receptor in heart

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**CLINICAL PERSPECTIVE**

At present, a role for 5-HT in valvulopathy has been established in only a small number of conditions, including carcinoid disease and long-term fenfluramine/phentermine treatment. Here, we show that a 5-HTT deficiency leads to the development of valvulopathy in mice, probably because of increased bioavailability of 5-HT, which is no longer internalized by vascular cells or platelets and cannot be degraded. Because 5-HTT is expressed by cardiac and lung tissues, these findings suggest a key role for this transporter in protecting the cardiac valves against the toxic effects of 5-HT released from platelets. This possibility is of considerable clinical relevance because 5-HTT expression is governed by many factors, including polymorphism of its gene promoter, hypoxia, inflammatory cytokines, and drugs. Although treatment with selective 5-HTT inhibitors given for mood disorders does not seem to induce valvular heart disease, some individuals may be more susceptible to these drugs because of a 5-HTT genotype associated with low expression of the protein. Whether 5-HT contributes to the pathogenesis of aortic sclerosis is also an open question. Examining 5-HTT gene polymorphism as a risk factor for aortic sclerosis is one way to examine this possibility. Another important finding from our study is that tryptophan hydroxylase, the rate-limiting enzyme of 5-HT synthesis, is expressed in the heart. This raises the intriguing possibility that dysregulation in local 5-HT synthesis may also play a role in valvular disorders, in addition to alterations in the activity or expression of 5-HTT.
Deficiency of the 5-Hydroxytryptamine Transporter Gene Leads to Cardiac Fibrosis and Valvulopathy in Mice

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