Muscle Sympathetic Nerve Activity Averaged Over 1 Minute Parallels Renal and Cardiac Sympathetic Nerve Activity in Response to a Forced Baroreceptor Pressure Change

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Background—Despite the accumulated knowledge of human muscle sympathetic nerve activity (SNA) as measured by microneurography, whether muscle SNA parallels renal and cardiac SNAs remains unknown.

Method and Results—In experiment 1, muscle (microneurography, tibial nerve), renal, and cardiac SNAs were recorded in anesthetized rabbits (n=6) while arterial pressure was changed by intravenous bolus injections of nitroprusside (3 μg/kg) followed by phenylephrine (3 μg/kg). In experiment 2, the carotid sinus region was vascularly isolated and vagotomized, and aorta-denervated rabbits (n=10). The 3 SNAs were recorded while intracarotid sinus pressure was increased stepwise from 40 to 160 mm Hg in 20-mm Hg increments maintained for 60 seconds each. Muscle SNA averaged over 1 minute was well correlated with renal (r=0.96±0.01, mean±SE) and cardiac (r=0.96±0.01) SNAs in experiment 1 (baroreflex closed-loop condition) and also with renal (r=0.97±0.01) and cardiac (r=0.97±0.01) SNAs in experiment 2 (baroreflex open-loop condition).

Conclusions—Muscle SNA averaged over 1 minute parallels renal and cardiac SNAs in response to a forced baroreceptor pressure change. (Circulation. 2005;112:384-386.)

Key Words: catecholamines ■ muscles ■ nervous system, autonomic ■ nervous system, sympathetic
Co) was inserted into the left tibial nerve to record muscle SNA, based on human\textsuperscript{2-5} and animal\textsuperscript{6} microneurography. We identified muscle SNA by the following discharge characteristics: (1) afferent activity induced by tapping of the calf muscles but not by gently touching the skin and (2) excitatory and inhibitory responses to a decrease and an increase in baroreceptor pressure, respectively. The nerve fibers peripheral to the electrodes were ligated securely to eliminate afferent signals. The preamplified signals of SNAs were bandpass filtered at 150 to 1000 Hz except those of muscle SNA in experiment 2 (480 to 5000 Hz). These signals were full-wave rectified and lowpass filtered (cutoff frequency, 30 Hz) to quantify nerve activity.

**Experiment 1: Baroreflex Closed-Loop Condition**

The rabbits were maintained in a supine position (n=6). All baroreceptor afferents and vagal nerves were intact. Three SNAs and AP were recorded at a 200-Hz sampling rate with a 12-bit analog-to-digital converter. After 2 minutes of baseline recording, nitroprusside (3 μg/kg) and, after a 2-minute delay, phenylephrine (3 μg/kg), was injected as a bolus via the right femoral vein. The data were averaged over 1 minute, the relations of muscle SNA against renal or cardiac SNAs (Figure 1A). When presented as SNA (time point P). Fine and bold lines indicate SNA signals resampled at 10 Hz and those averaged over 1 minute, respectively.

**Experiment 2: Baroreflex Open-Loop Condition**

To strictly control baroreceptor pressure (n=10 rabbits), a baroreflex loop was opened by vascular isolation of the carotid sinuses.\textsuperscript{14} Bilateral intracarotid sinus pressure (CSP) was controlled by a servo-controlled piston pump.\textsuperscript{14} Bilateral vagal and aortic depressor nerves were sectioned at the middle of the neck to eliminate reflexes from the cardiopulmonary region and the aortic arch. After surgical preparation, CSP was increased stepwise from 40 to 160 mm Hg in increments of 20 mm Hg. Each pressure step was maintained for 60 seconds. The 3 SNAs were recorded and stored as in protocol 1.

**Data and Statistical Analysis**

We averaged SNAs over 1 minute and generated scatterplots for muscle SNA against renal or cardiac SNA. For each type of SNA, 100 and 0 arbitrary units (AU) were assigned to the maximum 1-minute SNA value and the noise level determined by intravenous infusion of hexamethonium bromide (6 mg/kg),\textsuperscript{16} respectively. The other SNA signals were then normalized to these values. The correlation coefficients (r) for muscle SNA versus renal or cardiac SNA were determined.

In protocol 2, the relation between CSP and SNA was characterized by a 4-parameter logistic equation model: $y = P_2 + (P_4 - P_2)/(1 + \exp(P_1(x - P_3)))$, where $y$ is SNA and $x$ is CSP; $P_1$ is the response range of SNA; $P_2$ is the coefficient for calculation of gain; $P_3$ is the CSP corresponding to the midpoint of the operation; and $P_4$ is minimum SNA. All data are presented as mean±SD, and $P<0.05$ was considered significant.

**Results**

In experiment 1 (baroreflex closed-loop condition), nitroprusside injection decreased AP by 16±3 mm Hg while muscle SNA was increased. Subsequent phenylephrine injection increased AP by 41±9 mm Hg while muscle SNA was decreased. Thereafter, as AP gradually decreased, muscle SNA was again increased. These responses of muscle SNA were similar to those of renal and cardiac SNAs (Figure 1A). When presented as SNA averaged over 1 minute, the relations of muscle SNA against renal SNA and cardiac SNA were both close to the line of identity (Figure 1B). All animals showed strong correlations between 1-minute muscle and renal SNAs ($r=0.97±0.01$; range, 0.96 to 0.99) and between 1-minute muscle and cardiac SNAs ($r=0.97±0.01$; range, 0.95 to 0.99). The baroreflex relation of muscle SNA against CSP was almost superimposable on that of renal or cardiac SNA in individual animals. The parameters in a reverse-sigmoid logistic function fitted in muscle SNA were similar to those in renal or cardiac SNA:

**Figure 1.** Experiment 1. A, Representative integrated signals of renal, cardiac, and muscle SNA during intravenous bolus injections of nitroprusside (time point N) followed by phenylephrine (time point P). Fine and bold lines indicate SNA signals resampled at 10 Hz and those averaged over 1 minute, respectively. B, Scatterplots of 1-minute muscle SNA against 1-minute renal and cardiac SNAs of same data shown in A. C, Representative original (upper panels) and integrated (lower panels) signals of 3 SNAs before pharmacological injection from 1 animal. R-M indicates renal vs muscle SNAs; C-M, cardiac vs muscle SNAs.

**Figure 2.** Experiment 2. A, Representative integrated signals of renal, cardiac, and muscle SNA during 1-minute stepwise increases in CSP from 1 animal. Fine and bold lines indicate SNAs resampled at 10 Hz and those averaged over 1 minute, respectively. B, Scatterplots of 1-minute muscle SNA against renal and cardiac SNAs. C, Sigmoidal baroreflex relation between each SNA and CSP. B and C used same data as shown in A. R-M indicates renal vs muscle SNAs; C-M, cardiac vs muscle SNAs.
Discussion
Despite accumulated data of muscle SNA as measured by microneurography in human studies, whether muscle SNA parallels other SNAs controlling cardiovascular organs remains unclear. The major new finding in this study is that 1-minute muscle SNA was correlated strongly with both renal and cardiac SNAs, with ρ at nearly unity, in both baroreflex closed- and open-loop conditions. This finding supports our hypothesis that muscle SNA averaged over 1 minute parallels renal and cardiac SNAs in response to baroreflex forcing. Our finding suggests that microneurographic muscle SNA is a useful proxy for renal and cardiac SNA in addressing baroreflex control of SNA.

Earlier human studies\(^3\)\(^4\) reported that microneurographic muscle SNA was correlated with noradrenaline spillovers in the kidney (ρ=0.58) and heart (ρ=0.49) at rest, suggesting a correlation between muscle SNA and cardiac or renal SNA. However, because spillover values are affected by neurotransmitter kinetics in synapses (release and uptake) and circulating noradrenaline independent of SNA,\(^7\) these results are not definitive. The present study complemented and extended the human studies by recording these SNAs directly and demonstrated stronger correlations (ρ>0.95) between muscle SNA and cardiac or renal SNA than earlier studies of spillover technique.

Previous studies reported a greater response of splenic SNA to baroreceptor pressure changes than those of cardiac and renal SNAs in cats, suggesting regional differences in SNAs,\(^18\) but those studies did not investigate muscle SNA. Additionally, these regional differences were detected in faster SNAs averaged over 4 to 8 seconds.\(^18\)\(^19\) The present study investigated 1-minute SNA and hence did not address the relation between faster muscle SNA and renal or cardiac SNA.

The present study does not contradict earlier findings that indicated regionally different SNA responses to physiological stresses other than baroreceptor pressure changes. For example, the human cold pressor test increased muscle SNA but not heart rate.\(^20\)

Limitations
The anesthetic, artificial respiration, and surgical procedures used in this study may affect SNAs. In addition, experiment 2 was performed under a nonphysiological condition and did not investigate baroreflex hysteresis. We bandpass filtered all SNAs at the same condition (150 to 1000 Hz) except muscle SNA in experiment 2 (480 to 5000 Hz, human study condition).\(^2\) However, this did not affect the interpretation of data, because both experiments 1 and 2 showed strong correlations between muscle SNA and renal or cardiac SNA.

Conclusion
Muscle SNA averaged over 1 minute parallels renal and cardiac SNAs in response to a forced baroreceptor pressure change.

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References
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