In Vivo Cell Seeding With Anti-CD34 Antibodies Successfully Accelerates Endothelialization but Stimulates Intimal Hyperplasia in Porcine Arteriovenous Expanded Polytetrafluoroethylene Grafts

Joris I. Rotmans, MD, PhD; Jan M.M. Heyligers, MD; Hence J.M. Verhagen, MD, PhD; Evelyn Velema, BSc; Machiel M. Nagtegaal, BSc; Dominique P.V. de Kleijn, PhD; Flip G. de Groot, PhD; Erik S.G. Stroes, MD, PhD; Gerard Pasterkamp, MD, PhD

**Background**—The patency of AV expanded polytetrafluoroethylene (ePTFE) grafts for hemodialysis is impaired by intimal hyperplasia (IH) at the venous outflow tract. The absence of a functional endothelial monolayer on the prosthetic grafts is an important stimulus for IH. In the present study, we evaluated the feasibility of capturing endothelial progenitor cells in vivo using anti-CD34 antibodies on ePTFE grafts to inhibit IH in porcine AV ePTFE grafts.

**Methods and Results**—In 11 pigs, anti-CD34–coated ePTFE grafts were implanted between the carotid artery and internal jugular vein. Bare ePTFE grafts were implanted at the contralateral side. After 3 (n=11) or 28 (n=9) days, the pigs were terminated, and the AV grafts were excised for histological analysis and SEM. At 3 and 28 days after implantation, 95% and 85% of the coated graft surface was covered by endothelial cells. In contrast, no cell coverage was observed in the bare graft at 3 days, whereas at 28 days, bare grafts were partly covered with endothelial cells (32%; P=0.04). Twenty-eight days after implantation, IH at the venous anastomosis was strongly increased in anti-CD34–coated grafts (5.96±1.9 mm²) compared with bare grafts (1.70±0.4 mm²; P=0.03). This increase in IH coincided with enhanced cellular proliferation at the venous anastomosis.

**Conclusions**—Autoseeding with anti-CD34 antibodies results in rapid endothelialization within 72 hours. Despite persistent endothelial graft coverage, IH at the outflow tract is increased profoundly at 4 weeks after implantation. Further modifications are required to stimulate the protective effects of trapped endothelial cells. (Circulation. 2005;112:12-18.)

**Key Words:** arteriovenous anastomosis ■ endothelium ■ grafting ■ hyperplasia ■ renal dialysis

The patency of AV expanded polytetrafluoroethylene (ePTFE) grafts for hemodialysis is severely compromised by intimal hyperplasia (IH) at the venous outflow tract, ultimately leading to graft thrombosis.1 The clinical relevance of AV graft failure is illustrated by the limited 1- and 2-year primary patency rates of 50% and 25%, respectively.1,2 No effective intervention currently is available to improve graft patency. Successful development of new strategies requires closer insight into the pathogenesis of IH formation. IH formation is thought to reflect an accumulation of several separate entities, including inflammatory, coagulatory, and hemodynamic factors.3–5 A crucial factor in both activation of coagulation and inflammation is the lack of a functional endothelial monolayer on the prosthetic graft, because the endothelium constitutes the first-line homeostatic defense mechanism by exerting anticoagulatory and antiinflammatory effects.6–8 Consequently, endothelial cell (EC) seeding at the luminal surface of prosthetic vascular grafts is a valuable strategy to improve graft patency.9 Indeed, graft seeding with autologous ECs has been shown to increase patency rates of prosthetic bypass grafts in clinical trials.10 However, implementation of in vitro EC seeding is hampered by the laborious procedures for harvesting, expansion, and application of ECs obtained from autologous veins or adipose tissue.

Bone marrow–derived endothelial progenitor cells (EPCs) have emerged as a promising source of autologous ECs. EPCs are a subset of CD34(+) cells with the potential to proliferate and differentiate into mature ECs.11 Recent studies have emphasized that circulating EPCs have the capacity to home to sites of vascular injury, thus promoting the process of

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From the Laboratory of Experimental Cardiology (J.I.R., E.V., D.P.V.d.K., G.P.), Department of Vascular Surgery (J.M.M.H., H.J.M.V.), and Department of Haematology (J.M.M.H., F.G.d.G.), University Medical Center, Utrecht, and Department of Vascular Medicine, Academic Medical Center, Amsterdam (J.I.R., M.M.N., E.S.G.S.), the Netherlands.

Reprint requests to Gerard Pasterkamp, MD, PhD, Laboratory of Experimental Cardiology, University Medical Center, Heidelberglaan 100, Room G02.523, Utrecht, The Netherlands. E-mail g.pasterkamp@hli.azu.nl

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reendothelialization. Moreover, in vitro seeding of prosthetic vascular grafts with CD34(+) cells markedly increased graft endothelialization in animal models.

In the present study, we evaluated the feasibility of capturing EPCs in vivo using immobilized anti-CD34 antibodies on ePTFE grafts. If proved valid, this novel technique circumvents the laborious in vitro procedures mandatory for handling ECs. We hypothesized that in vivo “autoseeding” with immobilized anti-CD34 antibodies establishes a confluent, mature EC monolayer that may attenuate IH formation in the outflow tract of porcine AV ePTFE grafts.

**Methods**

**Study Design**

Eleven female Landrace pigs weighing 50.8±0.7 kg were used. In each pig, an anti-CD34–coated ePTFE graft was implanted between the carotid artery and the internal jugular vein at a randomly determined side. The same commercially available bare ePTFE graft was implanted at the contralateral side and served as an internal control. To study graft coverage, 2 pigs were terminated 72 hours after surgery. The remaining 9 pigs were terminated at 28 days of follow-up to evaluate cell retention on the graft and IH in the venous outflow tract. The study protocol was approved by the Ethics Committee on Animal Experimentation of the University Medical Center Utrecht, and animal care conformed to established guidelines.

**Graft Coating**

Anti–human CD34 monoclonal antibodies (IgG2a, epitope class III) were immobilized to the ePTFE graft material (Orbus Medical Technologies) with a proprietary multistep process (Ssens). The first step involves functionalization of the surface. To this functionalized surface, a biopolymer is covalently coupled in a reaction that leaves immobilized polymer without adding any new species to the graft or the polymer. Next, the anti–human CD34 monoclonal antibody is immobilized on the polymer-coated graft through stable peptide linkages. The anti-CD34–immobilized ePTFE grafts have been shown to exhibit cross-reactivity in porcine graft explants, which were observed to have a rich population of EC marker–positive cells after only 4 hours.

**Anesthesia**

Before operation and termination, the animals were fasted overnight and premedicated with intramuscular ketamine hydrochloride 10 mg/kg, midazolam 0.5 mg/kg, and atropine 0.5 mg and intravenous thiopental sodium 4 mg/kg. They were then intubated and ventilated with a mixture of O2 and air (1:2). An ear vein was used for continuous administration of midazolam 0.3 mg · kg⁻¹ · h⁻¹, sufentanil 2.5 μg · kg⁻¹ · h⁻¹, and pancuronium 50 μg · kg⁻¹ · h⁻¹.

**AV Graft Implantation**

Starting 6 days before the operation, the pigs received acetylsalicylic acid 80 mg/d. Clobidroglue 225 mg was given 1 day before operation and continued at a dose of 75 mg/d until termination. The ePTFE AV grafts were created bilaterally between the carotid artery and internal jugular vein as described previously. In short, heparin 5000 IU IV was administered before manipulation of the vessels. After dissection of the common carotid artery and the internal jugular vein, papaverin 5 mg/mL was applied locally to prevent vascular spasm. Next, the carotid artery was clamped, and a standardized 8-mm arteriotomy was performed. An end-to-side anastomosis was created at an angle of 45° by use of a continuous suture of 8-0 polypropylene (Ethicon). All ringed ePTFE grafts were 5 mm in diameter and 7 cm in length. The venous anastomosis was created in a similar fashion.

**Tissue Preparation and Histological Analysis**

After 72 hours (2 pigs) or 4 weeks (9 pigs) of follow-up, pigs were anesthetized as described previously. Heparin 10 000 IU was administered before manipulation of the vessels. Next, the carotid artery was cannulated, and the grafts and adjacent vessels were perfused with saline for 3 minutes. Subsequently, the pigs were euthanized, and the grafts and adjacent vessels were excised. The ePTFE grafts were then cut into 2 pieces: the first 2 cm from the arterial anastomosis was fixed in 2% glutaraldehyde for SEM, and the remainder of the ePTFE graft and the adjacent jugular vein was immersed in formalin for histological analysis. After 24 hours of formalin fixation, the graft and jugular vein were cut into 5-mm blocks and embedded in paraffin. Histological analysis was performed on 5-μm-thick sections obtained from 4 different locations to determine intimal and medial areas: at the center of the ePTFE graft with an in-between distance of 5 mm to determine graft coverage (2 sites), at the center of venous anastomosis, and 5 mm proximal (ie, caudal) to the venous anastomosis. For morphometric analysis, sections were stained with elastin van Gieson (EvG). With the highest magnification that allowed visualization of the entire vein section in 1 field, the intimal and medial areas were manually traced at the venous anastomosis and proximal to the anastomosis. The intima was defined as the tissue area encompassed by the internal elastic lamina. Thrombus formation was discriminated from IH through the use of hematoxylin-eosin– and EvG-stained sections. In the sections of the venous anastomosis, 2 parts of the intimal area were distinguished: the intima covering the graft (shoulder region) and the intima located at the venous part of the anastomosis (cushioning region). For immunohistochemical analysis, sections were incubated in 1.5% hydrogen peroxide in methanol to block endogenous peroxidases. Next, sections were incubated in boiling 10 mmol/L citrate acid for 15 minutes and subsequently preincubated with 10% horse serum (Vector Laboratories). Serial sections were stained with murine antibodies against α-smooth muscle actin (Sigma) for vascular smooth muscle cells (VSMCs) at 1:1500 dilution, Ki-67 (Immunotech) for cellular proliferation at 1:100 dilution, and MAC387 (Serotec) for macrophages at 1:10 dilution. Subsequently, sections were incubated with a biotinylated anti-mouse IgG antibody (Vector Laboratories) for 1 hour. To visualize ECs, sections were incubated with lectin from bandeiraea simplicifolia BS-1 at 1:100 dilution (Sigma) for 1 hour. Immunoreactive materials were visualized by use of streptavidin-labeled HRPO, diaminobenzidine in 0.05 mol/L Tris-CI mixed with 0.01 mol/L imidazole, and 0.1% hydrogen peroxide. Sections were counterstained with hematoxylin. Ki-67–stained sections from each venous anastomosis were used to measure the amount of proliferating cells divided by the sum of Ki-67–negative and –positive cells and expressed as a percentage.

**Scanning Electron Microscopy**

The integrity of the cellular coverage of the ePTFE grafts was also visualized by SEM. For this purpose, the ePTFE grafts were fixed in 2% glutaraldehyde and then dehydrated through increasing concentrations of ethanol (80% to 100%). The samples were dried with hexamethyldisilazane. Next, the ePTFE disks were sputter coated with a thin layer of platinum/palladium and analyzed with an SEM (Philips XL30).

**Statistical Evaluation**

Data are presented as mean±SEM. SPSS 11.0 was used for all statistical calculations. To ascertain the significance of differences, we performed the Wilcoxon test. A value of P<0.05 was considered significant.

**Results**

In total, 22 grafts were successfully implanted in 11 Landrace pigs. In the 2 pigs terminated after 3 days, all grafts (n=4)
were patent at time of harvest. In the 9 pigs terminated at 4 weeks, grafts were patent in 6 pigs, whereas bilateral occlusion was observed in 3 pigs. Histological analysis of the occluded grafts showed recent thrombotic occlusion on top of extensive IH in the venous outflow tract. These occluded grafts were excluded for further analysis because it is impossible to reliably perform morphometric analysis of IH in occluded grafts. The remaining patent grafts (n=16) were included for final analysis.

**Scanning Electron Microscopy**

Three days after graft implantation, the anti-CD34–coated grafts showed a confluent monolayer of cells. These attached cells had an inhomogeneous phenotype, ranging from round to spreading and flattened cells. In contrast, the bare grafts showed only little cellular adherence. Twenty-eight days after graft implantation, most of the attached cells on the coated graft were flattened. In some areas, cells with a plateletlike phenotype were observed on top of the cellular layer on the coated graft. At this time point, partial coverage of cells was observed in the bare grafts also (Figure 1).

**Histological Analysis**

**Graft Coverage**

Histological analysis of the grafts confirmed complete cellular coverage of the anti-CD34–coated grafts 3 days after graft implantation. The adhered cells were identified as ECs by lectin staining (Figure 2). EC coverage at 3 days after implantation was ≈95% in the coated graft and <5% in the bare grafts. Four weeks after implantation, 88±5% of the surface in the coated grafts was covered by lectin-positive cells, which was significantly higher than in the bare grafts (32±8%; P=0.04). (Figure 3A and 3B).

**Intimal and Medial Areas**

EvG-stained sections from the venous anastomosis revealed prominent intimal thickening at 4 weeks after graft implantation (Figure 3C and 3D). At this location, a strong increase
in intimal area was observed in coated grafts compared with bare grafts (the Table). Overall, the amount of microvessels in the intima was relatively low in both groups. The vast majority of microvessels were located in the adventitia of the recipient vein. In this vessel layer, microvessel formation also was comparable between groups. Approximately 90% of the intimal cells expressed the VSMC marker \( \alpha \)-actin (Figure 3E), whereas at the shoulder region of the venous anastomosis, a small amount of macrophages was observed in both groups. Thus, except for the size of the intimal area, no differences in composition of the IH lesions at the venous anastomosis were observed between groups. At the proximal vein (5 mm proximal to the toe of the venous anastomosis), the intimal and medial areas did not differ significantly between the 2 groups (the Table).

### Cellular Proliferation

Ki-67–stained sections obtained from the venous anastomosis showed prominent cellular proliferation in the intima (Figure 3F). At the venous anastomosis, the proliferation index was significantly higher in the coated grafts (20.9±2%) compared with the bare grafts (13.3±2%; \( P=0.027 \)). The higher proliferation rate in the coated grafts was due mainly to increased proliferation at the shoulder region (Figure 4).

### Discussion

In the present study, we show that anti-CD34 coating of AV ePTFE grafts results in a rapid and almost complete coverage of the graft with lectin-positive cells within 72 hours after implantation in pigs. This cellular coverage of coated grafts persists for at least 4 weeks. Despite cellular coverage of the

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**Figure 2.** Representative hematoxylin-eosin–stained sections of bare (A) and CD34-coated (B) grafts obtained from graft center at 72 hours after implantation. C, Adhered cells on CD34-coated grafts were identified as ECs by lectin staining.

**Figure 3.** Representative sections obtained at 4 weeks after graft implantation. Lectin-stained sections of bare (A) and CD34-coated (B) grafts obtained from graft center. EvG-stained sections of venous anastomosis of bare (C) and CD34-coated (D) grafts. E, Detail of \( \alpha \)-actin smooth muscle cell–stained section of venous anastomosis. F, Detail of Ki-67–stained section of venous anastomosis. Extensive proliferation is observed at shoulder region.
Graft Coverage With Endothelial (Progenitor) Cells

The concept of EC seeding of grafts is based on the assumption that these cells will constitute a biologically active lining that will attenuate activation of blood passing through the graft. Unfortunately, the complicated procedures for harvesting, expansion, and seeding of ECs in vitro have prevented this method from finding broader application in humans. EPCs are bone marrow–derived CD34(+) cells with the potential to proliferate and differentiate into mature ECs.11,20 Circulating EPCs have recently been shown to actively provide new ECs from the circulating blood to sites of endothelial denudation and/or injury.12,13 Promoting adherence of these “endogenous” circulating endothelial progenitor cells could in theory eliminate the need for in vitro seeding procedures. In the present study, we observed rapid coverage with lectin(+) cells from 3 days onward. Four weeks after implantation, the cellular coverage of anti-CD34–coated grafts remained almost confluent, whereas the histological appearance of the cells gradually changed to flattened. The degree of cellular coverage compares favorably to previous studies on in vitro graft seeding techniques with CD34(+) cells in which graft coverage did not exceed 65%.15 The efficiency of in vitro seeding techniques using mature ECs has also been limited by low levels of EC retention.21,22 Combined, these data illustrate that antibody-assisted auto-seeding of CD34(+) cells is associated with more confluent coverage of grafts, whereas it simultaneously offers a safer and easier technique for clinical use.

Graft Coverage and IH

Increased graft coverage should result in less activation of circulating blood cells and potentially contribute to paracrine mediators conveying antiinflammatory and antiproliferative effects downstream at the venous outflow tract. This line of events assumes that the attached cells are able to exert effects similar to the physiological actions of a normal endothelial lining. In contrast, despite complete graft endothelialization in the anti-CD34–coated grafts, we observe a 3-fold increase in IH at the venous anastomosis.

Concomitantly, a strong increase in cellular proliferation was present, predominantly at the shoulder region of the venous outflow tract. These data imply a proliferative rather than antiproliferative effect of the cells covering the graft. Our results are consistent with those of Kang et al.,23 who showed aggravated restenosis after intracoronary infusion of peripheral blood stem cells in patients who underwent coronary stenting. Several factors may have contributed to the adverse effects in the anti-CD34–coated grafts. First, it has previously been reported that CD34(+) cells are able to

### Summary of Morphometric Analysis of EvG-Stained Sections Obtained From the Center of the Venous Anastomosis and the Proximal Jugular Vein

<table>
<thead>
<tr>
<th></th>
<th>Anti-CD34–Coated Grafts (n=6)</th>
<th>Bare Grafts (n=6)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Center of venous anastomosis</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total intimal area, mm²</td>
<td>5.96±1.9</td>
<td>1.70±0.4</td>
<td>0.03</td>
</tr>
<tr>
<td>Shoulder region, mm²</td>
<td>2.47±0.8</td>
<td>0.84±0.4</td>
<td>0.03</td>
</tr>
<tr>
<td>Cushioning region, mm²</td>
<td>3.48±1.6</td>
<td>0.86±0.3</td>
<td>0.08</td>
</tr>
<tr>
<td>Medial area, mm²</td>
<td>2.95±0.8</td>
<td>1.49±0.2</td>
<td>0.05</td>
</tr>
<tr>
<td>Intima-to-media ratio</td>
<td>1.95±0.4</td>
<td>1.23±0.4</td>
<td>0.03</td>
</tr>
<tr>
<td>Jugular vein (5 mm proximal to venous anastomosis)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intimal area, mm²</td>
<td>2.49±0.6</td>
<td>2.39±1.0</td>
<td>0.75</td>
</tr>
<tr>
<td>Medial area, mm²</td>
<td>2.75±0.3</td>
<td>2.29±0.3</td>
<td>0.25</td>
</tr>
<tr>
<td>Intima-to-media ratio</td>
<td>0.85±0.2</td>
<td>0.88±0.3</td>
<td>0.75</td>
</tr>
</tbody>
</table>

All values are expressed as mean±SEM.
differentiate into various cell types, including ECs.
Thus, VSMCs can originate from bone marrow–derived CD34(+) progenitor cells, implying that differentiation of CD34(+) captured cells into VSMCs can contribute to the increased proliferation index in CD34-coated grafts. Where differentiation of CD34(+) progenitor cells into macrophages has been reported, we observed no significant difference in macrophage numbers between coated and bare grafts. Second, EPCs have the capacity to release potent proangiogenic growth factors such as vascular endothelial growth factor (VEGF) and hepatocyte growth factor. The proliferative and migratory effects of these cytokines are not restricted to ECs but also include VSMCs. Enhanced secretion of proliferation factors from cells adhered to anti-CD34-coated grafts may have contributed to increased IH at the venous anastomosis. Third, interaction of ECs with CD34–coated grafts may have contributed to increased IH at the venous anastomosis.

**Study Limitations**

The adhered cells were characterized as endothelial-like cells by their morphological appearance on SEM and by immunohistochemistry using lectin from Bandeiraea simplicifolia, a marker for porcine ECs. However, extensive characterization of the adhered cells was not feasible because of the unavailability of monoclonal antibodies for porcine endothelial (progenitor) cells markers such as VE-cadherin in EPCs. Therefore, the absence of a physiological microenvironment may also contribute to the apparent lack of protective effects. Fourth, laminar shear stress has been shown to upregulate EC-specific markers such as VE-cadherin in EPCs. Therefore, the turbulent flow pattern in AV grafts may have deteriorated differentiation into ECs, especially in the anastomatic region. Finally, adequate maturation of captured CD34(+) cells may have been hampered by the binding of the immobilized antibody to the CD34 epitope.

**Clinical Implications and Future Directions**

The present findings indicate that autoseeding with EPCs offers a potential strategy to establish graft endothelialization in AV grafts. It remains to be established whether and to what extent these captured cells can be stimulated to regain their functional capabilities. In this respect, it is interesting to note recent developments demonstrating the regulatory role of vascular endothelial growth factor, erythropoietin, and angiopoietins on maturation of endothelial progenitor cells. In addition, capturing a mixture of both CD34(+) and CD14(+)CD34(–) cells also holds promise for optimal EC differentiation and functionality of EPCs. These options need to be addressed in future studies.

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**References**

The patency of arteriovenous grafts for hemodialysis is severely compromised by intimal hyperplasia at the venous outflow tract. The concept of endothelial cell seeding of expanded polytetrafluoroethylene (ePTFE) grafts is based on the assumption that these cells will constitute a biologically active lining that will attenuate activation of blood passing through the graft, thereby reducing the release of mitogens for vascular smooth muscle cells (VSMCs). Endothelial progenitor cells are bone marrow–derived CD34(+) cells with the potential to proliferate and differentiate into mature endothelial cells. In the present study, we evaluated the feasibility of “autoseeding” with endothelial progenitor cells using immobilized CD34(/H11001) coating grafts. The present findings show that anti-CD34 coating is successful in promoting autoseeding of ePTFE grafts, whereas the trapped cells aggravate neointimal formation. Several factors can contribute to these adverse effects of stem-cell seeding, including differentiation of CD34(+) into VSMCs and the release of potent mitogens for VSMCs by the trapped cells. Further modifications of the graft coating are required to stimulate the protective effects of trapped endothelial cells.
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