Novel Cardioprotective Role of a Small Heat-Shock Protein, Hsp20, Against Ischemia/Reperfusion Injury

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Background—Heat-shock proteins (Hsps) have been shown to render cardioprotection from stress-induced injury; however, little is known about the role of another small heat-shock protein, Hsp20, which regulates activities of vasodilation and platelet aggregation, in cardioprotection against ischemia injury. We recently reported that increased expression of Hsp20 in cardiomyocytes was associated with improved contraction and protection against β-agonist–induced apoptosis.

Methods and Results—To investigate whether overexpression of Hsp20 exerts protective effects in both ex vivo and in vivo ischemia/reperfusion (I/R) injury, we generated a transgenic (TG) mouse model with cardiac-specific overexpression of Hsp20 (10-fold). TG and wild-type (WT) hearts were then subjected to global no-flow I/R (45 minutes/120 minutes) using the Langendorff preparation. TG hearts exhibited improved recovery of contractile performance over the whole reperfusion period. This improvement was accompanied by a 2-fold decrease in lactate dehydrogenase released from the TG hearts. The extent of infarction and apoptotic cell death was also significantly decreased, which was associated with increased protein ratio of Bcl-2/Bax and reduced caspase-3 activity in TG hearts. Furthermore, in vivo experiments of 30-minute myocardial ischemia, via coronary artery occlusion, followed by 24-hour reperfusion, showed that the infarct region–to–risk region ratio was 8.1 ± 1.1% in TG hearts (n = 7), compared with 19.5 ± 2.1% in WT hearts (n = 11, P < 0.001).

Conclusions—Our data demonstrate that increased Hsp20 expression in the heart protects against I/R injury, resulting in improved recovery of cardiac function and reduced infarction. Thus, Hsp20 may constitute a new therapeutic target for ischemic heart diseases. (Circulation. 2005;111:1792-1799.)

Key Words: ischemia ■ reperfusion ■ myocardial infarction ■ proteins
sis. To further define the functional significance of Hsp20 in vivo and its potential protective mechanisms, we generated a TG mouse model with cardiac-specific overexpression of this protein. Our findings indicate that increased Hsp20 expression in the heart protects against IR injury, resulting in full functional recovery and reduced infarction, which implicates this protein as a potential therapeutic target for ischemic heart disease.

**Methods**

**Generation of TG Mouse Model**

TG mice were generated by using mouse cardiac Hsp20 cDNA (mcHsp20) under control of α-myosin heavy chain promoter (α-MHCp), a 600-bp DNA fragment containing human growth hormone polyadenylation sequences (hGHpA) ligated 3’ to 500-bp mcHsp20 cDNA open reading frame. Fragment containing mcHsp20/hGHpA was then ligated downstream from 5.5 kb of mouse cardiac-specific α-MHC promoter. Positions of PCR primers used for genotyping are indicated by arrows. b, Genotypic analysis of genomic DNA from WT (−) and TG (+) mice. Control PCR was set up to amplify a 350-bp fragment of TSH-β. c, Ventricular sections from 3-month-old mice stained with hematoxylin and eosin (magnification, ×400) indicated no apparent morphological/pathological abnormalities in TG hearts. d, Aliquots (20 μg) of different tissue homogenates from WT and TG mice were subjected to Western blotting with an anti-Hsp20 antibody. e, Quantitative immunoblotting analysis showed that there was a 10-fold increase in Hsp20 protein level in TG hearts relative to WT hearts (1.0). f, Increased Hsp20 expression did not alter Hsp25 and αB-crystallin protein levels.

**Immunoblotting**

Heart homogenates were analyzed by standard Western blotting to compare Hsp20, Hsp25, αB-crystallin, Bcl-2, Bax, actin, and α-actin levels. Binding of the primary antibody was detected by peroxidase-conjugated secondary antibodies and enhanced chemiluminescence (Amersham), and bands were quantified with densitometry. Antibodies Hsp20 (Research Diagnostics Inc), Hsp25 (Affinity Bioreagents Inc), αB-crystallin (Calbiochem), Bcl-2(C-2), Bax (B-9), actin, and α-actin (Santa Cruz Biotechnology Inc).

**Immunoprecipitation**

Association of Hsp20 with Bax and Bcl-2 was studied using Dynabeads Protein G (Dynal Biotech) according to the manufacturer’s instructions. Briefly, cardiac homogenate from wild-type (WT) heart was precleared by incubating with the beads for 1 hour at 4°C to minimize nonspecific binding. Fresh beads were washed with 0.1
mol/L sodium phosphate buffer and then coated with anti-Hsp20 or anti-Bax antibody. The bound antibody was crosslinked to the beads using 20 μmol/L dimethyl pimelidate in a 0.2 mol/L triethanolamine solution. The precleared homogenate was then added to the crosslinked beads, and binding was mediated at 4°C for 1 hour. Finally, the proteins were eluted off (using 0.1 mol/L citrate), and their identity was determined by immunoblotting.

Global Ischemia Ex Vivo
The cellular and functional responses to I/R were assessed in mice by using an isolated perfused heart model as previously described.24,25 Male adult mice (12 to 14 weeks old) were anesthetized intraperitoneally (IP) with pentobarbital sodium (50 mg/kg). Hearts were rapidly excised and mounted on a Langendorff apparatus, perfused with Krebs-Henseleit buffer (noncirculating), and stabilized for 30 minutes, and then the hearts were subjected to 45 minutes of no-flow global ischemia and 2 hours of reperfusion. A fluid-filled balloon made of plastic film was inserted into the left ventricle via the mitral valve and inflated to yield a left ventricular end-diastolic pressure of 10 mm Hg. The balloon was attached via polyethylene tubing (PE50) to a pressure transducer connected to a Heart Performance Analyzer (Micro-Med), and continuous left ventricular pressure was measured. A bipolar electrode (NuMed) was inserted into the right atrium, and atrial pacing was performed at 400 bpm with a Grass S-5 stimulator. Pacing was stopped during ischemia and restarted at reperfusion. After reperfusion, the hearts were weighed, frozen, and cut into 8 sections, with 1 section made at the site of the ligature. Infarct sizes were determined and expressed as a percentage of the region at risk. Perfusion effluent was collected every 15 minutes of preischemia and also during measuring lactate dehydrogenase (LDH) release. Perfusion effluent was rapidly excised and mounted on a Langendorff apparatus, perfused with 5% phthalo blue. Hearts were transversely cut into 5 to 6 sections, with 1 section made at the site of the ligature. Infarct sizes were determined and expressed as a percentage of the region at risk.

Lactate Dehydrogenase Release and TUNEL Assays
In addition to cardiac function, cardiac injury was assessed by measuring lactate dehydrogenase (LDH) release. Perfusion effluent was collected every 15 minutes of preischemia and also during reperfusion. Total LDH released from the heart was determined using a CytoTox 96 assay (Promega) and expressed as units per gram of wet heart weight. For terminal dUTP nick end-labeling (TUNEL) observation (cyanosis) and continuous ECG monitoring. After 24 hours of reperfusion, the aorta was cannulated, and the heart was perfused with 1% TTC (37°C, 60 mm Hg) as previously described.28 The occluder, which had been left in place, was repositioned, and the heart was perfused with 5% phthalo blue. Hearts were transversely cut into 5 to 6 sections, with 1 section made at the site of the ligature. Infarct sizes were determined and expressed as a percentage of the region at risk.

Statistical Analysis
Data are expressed as mean±SEM. Statistical analysis was performed using a 2-tailed Student t test for unpaired observations and ANOVA for multiple comparisons. Values of P<0.05 were considered statistically significant.

Results
Generation of Hsp20 TG Mice
We generated TG mice that carry the mouse cardiac Hsp20 cDNA under the control of the α-MHC mouse promoter (Figure 1, a and b). All Hsp20 TG mice were healthy and showed no apparent cardiac morphological or pathological abnormalities (Figure 1c). Western blot analysis (Figure 1d) of tissue homogenates revealed that Hsp20 was expressed predominantly in the cardiac and in much lower abundance in skeletal and smooth muscles of WT mice. Transgenesis resulted in a 10-fold increase of the Hsp20 protein level in the

Baseline Functional Parameters in Langendorff-Perfused Isovolumically Contracting Mouse Hearts

<table>
<thead>
<tr>
<th></th>
<th>HR, bpm</th>
<th>BW, g</th>
<th>HW, mg</th>
<th>LVDP, mm Hg</th>
<th>+dP/dt, mm Hg/s</th>
<th>−dP/dt, mm Hg/s</th>
<th>Outflow, ml/min</th>
</tr>
</thead>
<tbody>
<tr>
<td>TG, n=8</td>
<td>354±15</td>
<td>27.4±1.2</td>
<td>174±14</td>
<td>83±6*</td>
<td>4429±149*</td>
<td>2370±128*</td>
<td>1.3±0.2</td>
</tr>
<tr>
<td>WT, n=9</td>
<td>363±20</td>
<td>27.8±0.9</td>
<td>171±12</td>
<td>69±4</td>
<td>3311±185</td>
<td>1891±100</td>
<td>1.4±0.3</td>
</tr>
</tbody>
</table>

HR indicates heart rate; BW, body weight; HW, heart weight; LVDP, left ventricular developed pressure; +dP/dt, rate of contraction; and −dP/dt, rate of relaxation.

*P<0.05 vs corresponding WT group.
TG hearts (Figure 1e). Cardiac overexpression of Hsp20 did not alter the expression of other small Hsps, such as Hsp25 and αB-crystallin, in the heart (Figure 1f).

Improvement in the Postischemic Recovery of Function

Because acute expression of Hsp20 in rat cardiomyocytes is protective against apoptosis,23 we examined whether increased in vivo expression of Hsp20 protects against postischemic injury. To exclude the involvement of inflammatory components on reperfusion, we used an isolated perfused-heart preparation. Body weights and heart weights of the mice used in these studies were similar between the TG and WT groups (Table). Hearts were stabilized for 30 minutes, and baseline function was measured. Hsp20 overexpression resulted in an increased contractile function under basal conditions (Table), consistent with our previous report using adenovirus-mediated Hsp20 gene transfer in cardiomyocytes.20 Hearts were then subjected to 45 minutes of global ischemia and 2 hours of reperfusion. During reperfusion, the TG hearts exhibited significantly better functional recovery than the WT hearts (Figure 2). It is noteworthy that recovery of left ventricular developed pressure and the rates of contraction (+dP/dt) and relaxation (−dP/dt) were 54±5% and 59±4%, respectively, in the TG hearts and only 20±3% and 21±3%, respectively, in WT hearts after 10 minutes of reperfusion. Most importantly, these parameters were completely recovered after 1 hour of reperfusion in TG hearts, whereas WT hearts recovered to 69±6% and 71±4%, respectively, of preischemic values.

Attenuation of IR-Induced Necrosis and Apoptosis

To determine the degree of necrosis in these IR hearts, we assessed the level of LDH released during the first hour of reperfusion after global ischemia. The total LDH was twice as high in WT hearts compared with TG hearts (Figure 3a), which indicated reduced necrosis in Hsp20-overexpressing hearts. Therefore, we examined whether the functional pro-
Protection of the Hsp20 TG hearts was related to the antiapoptotic property of small Hsps. Heart lysates from a subset of experimental WT and TG animals were assayed for DNA fragmentation by use of a quantitative nucleosome assay. WT hearts exhibited a 3-fold increase over Hsp20 TG hearts (Figure 3b). To substantiate the protective effect of Hsp20 via its antiapoptotic action, we also performed TUNEL assays of the hearts. TUNEL-positive cardiomyocytes from the left ventricles of WT animals were 4-fold higher than those in TG animals (Figure 3c). Thus, both methods of detecting apoptosis (DNA fragmentation and TUNEL assay) demonstrated significant attenuation in TG hearts.

Decreased Myocardial Infarct Size Ex Vivo and In Vivo

After 45 minutes of no-flow, global ischemia, followed by 120 minutes of reperfusion ex vivo, we determined myocardial infarct size by histochemical and TTC staining. Histochemical examination of post–ischemic/reperfused WT hearts revealed contraction bands (hypercontracted myofibers) and vacuolizations (Figure 4a and c), which are both indicative of cardiomyocyte damage; however, the myofibers were well preserved in Hsp20 TG hearts (Figure 4b and d), suggesting that priming the heart with Hsp20 maintains the integrity of the muscle during IR injury. Myocardial infarct size as a percentage of the area at risk was reduced by 6-fold in TG compared with WT hearts (Figure 4e).

These studies were extended to an in vivo model of 30-minute myocardial ischemia, via coronary artery occlusion, followed by 24-hour reperfusion. Importantly, the infarct region–to–risk region ratio was 8.1 ± 1.1% in TG hearts (n = 7), compared with 19.5 ± 2.1% in WT hearts (n = 11, P < 0.001), under in vivo conditions (Figure 4f).

Mechanism(s) of Hsp20 Cardioprotective Effects in IR

To elucidate the potential mechanism(s) of Hsp20 cardioprotection, we first assessed Bcl-2 and Bax protein expression levels in hearts before or after IR, because alterations of Bcl-2 and Bax protein levels have been shown in isolated cardiomyocytes after hypoxia/reoxygenation and in hearts during IR. Overexpression of Hsp20 did not alter the expression levels of either Bcl-2 or Bax (Figure 5a and b); however, on IR, the levels of Bcl-2 were significantly reduced, whereas the levels of Bax were greatly increased, in WT hearts, compared with their preischemic samples (Figure 5a and b). Consequently, the relative ratio of Bcl-2 to Bax expression in the TG hearts was more than 2-fold higher, compared with WT hearts after IR (Figure 5c). Moreover, immunoprecipitation of homogenates from WT hearts using the Hsp20 antibody revealed that Hsp20 interacted with Bax but not Bcl-2 (Figure 5d). A reciprocal immunoprecipitation approach using the Bax antibody also demonstrated an association of Bax with Hsp20 (Figure 5e). Furthermore, the activity of caspase-3 was significantly reduced in TG compared with WT hearts on IR (Figure 5f).

Hsp20 has been shown previously to be phosphorylated under β-adrenergic stimulation of isolated cardiomyocytes. To determine whether the protective effects of...
Figure 5. Effects of Hsp20 overexpression on apoptosis-related proteins. In ex vivo WT hearts, protein levels of Bcl-2 (a) were significantly decreased, whereas Bax (b) levels were significantly increased, after 45 minutes of no-flow ischemia, followed by 120 minutes of reperfusion. These proteins showed no alterations in TG hearts. c, Ratio of Bcl-2/Bax protein levels in post–ischemic/reperfused TG hearts was significantly higher than in WT hearts. d and e, Hsp20 coimmunoprecipitated with Bax but not with Bcl-2. Mouse heart homogenates were immunoprecipitated with anti-Hsp20 (d) or anti-Bax antibodies (e). Proteins were separated on SDS-PAGE gels and probed with anti-Bax (d1) or anti-Hsp20 antibodies (e1). Then, membrane (d1) was stripped and reprobed with anti-Bcl-2(d2); subsequently, this membrane (d2) was restriped and reprobed with anti-Hsp20 (d3). Similarly, membrane (e1) was stripped and reprobed with anti-Bax antibodies (e2). Preimmunoprecipitated heart homogenate was used as positive control (H11001), and immunoprecipitate without Hsp20 or Bax antibodies was used as negative control (H11002). f, Caspase-3 activity was significantly lower in TG hearts than in WT hearts after IR. *P<0.05 vs post–ischemic/reperfused WT hearts; n=6 in each group. g–j, Hsp20 phosphorylation was analyzed by use of 2D gel electrophoresis. Homogenates were obtained from pre–ischemic/reperfused (g and i) and post–ischemic/reperfused (h and j) WT and TG hearts that were subjected to occlusion of LAD artery for 30 minutes and reperfusion for 24 hours. Homogenates were extracted by use of a solubilization solution of 7 mol/L urea, 2 mol/L thiourea, 4% CHAPS, 20 mmol/L DTT, 20 mmol/L spermine, and 1 mmol/L PMSF. Extract was centrifuged (100 000 g, 1 hour, at 10°C), and 500 µg from each preparation was separated by 2D electrophoresis, using isoelectric focusing strips (immobiline Drystrips, 18 cm, pH 4 to 7). 2D gels were Sypro Ruby–stained, and protein spots corresponding to phosphorylated (pI of 5.5) or nonphosphorylated (pI of 5.7) Hsp20 were further confirmed by Western blotting and LC-MS/MS analysis (data not shown).
Hsp20 against IR injury in vivo may be associated with increases in its phosphorylation levels, we applied 2D gel electrophoresis. Our findings indicated that 16±1% of total Hsp20 was phosphorylated in TG hearts (Figure 5i). This phosphorylation was significantly increased to 37±2% after 24 hours of reperfusion (Figure 5j), suggesting that Hsp20 phosphorylation plays an important role in cardioprotection against IR injury. Similar assessment of altered Hsp20 phosphorylation in WT hearts was not feasible because of the low levels of endogenous Hsp20 expression (Figure 5, g and h).20,23

Discussion

In experimental IR models, although apoptosis may be initiated during ischemia, the number of apoptotic cells is increased during reperfusion.3 These observations suggest that the apoptotic component contributes to the death of the myocardium particularly during reperfusion. Thus, unraveling these apoptotic mechanisms becomes very important in developing new therapeutic approaches to attenuate cell death resulting from IR injury.

Increasing evidence has shown that several Hsps have anti-apoptotic roles, and overexpression of Hsp27, αB-crystallin, Hsp32 (HO-1), and Hsp70 in the heart can attenuate the IR injury and improve cardiac function9–15,31; however, the Hsp20 studied here is different from the other Hsps in its unique protein kinase A/protein kinase G phosphorylation site RRAS20,21,23 (phosphorylation of the Ser-16 site significantly increases the contractility in cardiomyocytes; unpublished data), regulatory activities of vasorelaxation,21 and platelet aggregation.22 Furthermore, previous studies have shown that Hsp20 is translocated to actin filaments on stress, suggesting its cytoskeletal stabilizing function in cardiomyocytes.23,32 Taken together, these properties of Hsp20 suggest that it may benefit the ischemic heart at multiple levels.

In the present study, we investigated the role of Hsp20 on cardioprotection during myocardial ischemia. Interestingly, cardiac-selective overexpression of Hsp20 was associated with full functional recovery and decreased infarct size both ex vivo and in vivo on IR injury. There are at least 2 mechanisms that underlie cardioprotection in Hsp20 TG mice from IR injury. The first involves stabilization of the apoptosis-related proteins Bcl-2 and Bax by Hsp20 overexpression (Figure 5, a–c), whereas WT hearts exhibited decreases in Bcl-2 and increases in Bax levels after IR, similar to recent findings in vivo.30 It has been shown previously that overexpression of Bcl-224 or ablation of Bax25 was associated with improved cardiac function, which correlated with a reduction of cardiomyocyte apoptosis, after IR. In addition, delayed ischemic preconditioning, on phenylephrine treatment of rabbit hearts, resulted in an increased Bcl-2/Bax ratio and a reduction of apoptosis.33 Thus, preserved Bcl-2 and Bax protein levels may be an important mechanism of inhibiting IR–induced apoptosis by overexpression of Hsp20. Importantly, we showed that Hsp20 complexed with the protein Bax (Figure 5, d and e), which possibly prevented the translocation of Bax from the cytosol into the mitochondria during IR. As a result, Hsp20 may preserve the integrity of mitochondria, restrict release of cytochrome c, and repress activation of caspase-3. In fact, our data demonstrated decreased caspase-3 activity in post–ischemic/reperfused TG hearts (Figure 5f), suggesting that cardioprotection from IR–induced apoptosis may involve inhibition of the conversion of procaspase-3 (p24) to active caspase-3 because of inhibition of Bax translocation (data not shown). It has been shown that caspase-3 expression is increased in association with heart failure and apoptosis in experimental animals.34 Several groups have reported that direct blockade of caspase activation with a peptide inhibitor protects the myocardium against lethal reperfusion injury.34,35 In contrast, heart-targeted overexpression of caspase-3 in mice has been shown to increase infarct size and depress cardiac function.36 Therefore, the decreased activation of caspase-3 might also be one of the cardioprotective mechanisms against IR-induced injury.

Recently, Hsp60 and αB-crystallin have also been shown to be complexed with the proapoptotic protein Bax.20,27,39 Under stress conditions, these Hsps and Bax dissociate, whereupon Bax translocates to the mitochondria to participate in apoptosis, suggesting a role for Hsps upstream of caspase activation; however, Hsp70 was found to protect cells from death induced by enforced expression of caspase-3, suggesting that protection by Hsp70 may occur downstream of caspase activation.40 In addition, Hsp70 has been reported to interfere with apoptosis by a direct interaction with Apaf-1.41,42 Moreover, Hsp27 has been reported to directly bind to cytosolic cytochrome c and sequester it from Apaf-1.43,44 Thus, different Hsps may act via different mechanisms to prevent cell death. The data presented here demonstrate that Hsp20 prevents IR-induced apoptosis, possibly through the Bax-caspase pathway.

Another cardioprotective mechanism of Hsp20 may involve the maintenance of muscle integrity during IR injury, as evidenced by the presence of less damaged myofibril in the TG hearts after IR. These cardioprotective effects may be associated with increased Hsp20 phosphorylation in the TG hearts after IR, consistent with previous reports on enhanced interaction of phosphorylated Hsp20 with actin and actinin, which further stabilizes the microfilaments.23,45 Because there is strong evidence that cytoskeletal injury plays a crucial role in the pathogenesis of myocardial ischemic injury,46 it is plausible that Hsp20 translocates from the soluble fraction of cardiomyocytes to the insoluble fraction after IR, leading to protection of the collapsed intermediate filament network or cytoskeletal protein damage.

In summary, our findings demonstrate that increased expression of Hsp20 protects the heart from IR-induced injury, leading to restoration of cardiac function and reduced infarction. Thus, Hsp20 may provide a new potential therapeutic target for heart disease. Future studies using adenoviral or adenosivirus-associated viral gene transfer in vivo after myocardial infarction may further elucidate the potential clinical benefits of Hsp20.

Acknowledgments

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References


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