Toll-Like Receptor-2 Modulates Ventricular Remodeling After Myocardial Infarction

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Background—Toll-like receptors (TLRs) are members of the interleukin-1 receptor family and transduce similar signals as interleukin-1 receptor in response to exogenous pathogens. Recent studies have demonstrated that TLRs are activated by endogenous signals, such as heat shock proteins and oxidative stress, that may contribute to ventricular remodeling after myocardial infarction. In this study, we determined whether TLR-2 was involved in cardiac remodeling after myocardial infarction.

Methods and Results—Myocardial infarction was induced by surgical left anterior descending coronary artery ligation on wild-type (WT) mice and TLR-2–knockout (KO) mice. The survival rate was significantly higher in KO mice than in WT mice 4 weeks after myocardial infarction (65% versus 43%, P<0.03). Infarct size and degree of inflammatory cell infiltration in infarct area were similar between WT and KO mice. However, myocardial fibrosis in the noninfarct area of KO mice was much less than in WT mice (P<0.01) and was accompanied by reduced transforming growth factor-β1 and collagen type 1 mRNA expressions (P<0.01 and P<0.05, respectively). Left ventricular dimensions at end diastole were smaller in KO mice than in WT mice at 1 week (P<0.05) and 4 weeks (P<0.01) after surgery. Furthermore, fractional shortening was higher (27.7±2.5% versus 21.2±2.6%, P<0.05, at 1 week, and 24.3±2.0% versus 16.6±2.5%, P<0.01, at 4 weeks) in KO mice compared with WT mice.

Conclusions—These data suggest that TLR-2 plays an important role in ventricular remodeling after myocardial infarction.

Key Words: receptors • remodeling • myocardial infarction • heart failure

Toll-like receptors (TLRs) recognize exogenous ligands, such as lipopolysaccharide, peptidoglycan, bacterial lipoprotein, and oligonucleotides, during the inflammatory responses.1,2 TLRs have interleukin-1 receptor–like intracellular signaling pathways that lead to nuclear localization of nuclear factor (NF)-κB/Rel-type transcription factors.1–3 Furthermore, TLRs are expressed in various organs, such as lung, brain, kidney, and heart.2,4 In vitro studies using isolated cardiomyocytes and in vivo studies using C3H/HeJ mice, which are TLR-4 deficient, have suggested that myocardial activated TLR-mediated signaling pathways in response to exogenous ligands induce cardiac dysfunction.5–7 Recent studies have demonstrated that TLR-mediated signaling pathways are also activated by endogenous ligands, such as heat-shock protein and oxidative stress, in isolated ventricular cardiomyocytes.8,9 A large myocardial infarction induces ventricular remodeling, which contributes to cardiac dysfunction and mortality.10,11 The ventricular remodeling process involves several mediators, such as oxidative stress,12 cytokines,13 and neurohormones.14 Several lines of evidence have suggested that oxidative stress alters cell survival and hypertrophic signals that lead to cardiomyocyte apoptosis and/or hypertrophy in isolated cardiomyocytes and in animals.12,15–18 Because oxidative stress after myocardial infarction was identified as playing a pivotal role for ventricular remodeling, we hypothesized that TLR-2 deficiency resulted in derangement of ventricular remodeling and preservation in cardiac function after myocardial infarction. To evaluate the potential role of TLR-2 on ventricular remodeling after myocardial infarction, we examined the effects of TLR-2 deficiency on cardiac function, structural aspects of ventricular remodeling,
and survival rates after myocardial infarction in a mouse model.

Methods

Animals

Male TLR-2–knockout (KO) mice and wild-type (WT) mice with the same C57BL/6 backgrounds were used. Mice were housed in a facility with a 12/12-hour light/dark cycle and were given free access to water and standard rodent chow. The room was kept specific pathogen–free. The animals were handled according to the animal welfare regulations of Yamagata University, and the study protocol was approved by the Animal Subjects Committee of Yamagata University. The investigation conformed to the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health.

Left Coronary Artery Ligation

Induction of myocardial infarction was performed as described previously, with minor modifications. Briefly, mice (25 to 35 g body weight) were anesthetized by intraperitoneal injection with a mixture of ketamine (80 mg/kg) and xylazine (8 mg/kg). Animals were intubated with a 20-gauge polyethylene catheter and were ventilated with a rodent ventilator (Harvard Apparatus). A median sternotomy was performed, the left anterior descending artery was subsequently tied off. Infarction was evident from discoloration of the left ventricle (LV). Finally, the chest wall was closed. The animals remained in a supervised setting until fully conscious.

Echocardiography

Transthoracic echocardiography was performed as described previously, with a FFsonic 8900 (Fukuda Denshi) with a 10-MHz phased-array transducer at 1 or 4 weeks after surgery. LV internal dimensions at end systole and end diastole (LVESD and LVEDD) were measured digitally on the M-mode tracings and averaged from 3 cardiac cycles. LV fractional shortening (%FS) was calculated as ([LVEDD − LVESD]/LVEDD)×100.

Morphological Examination and Infarct Size Measurement

At 3 days, 1 week, and 4 weeks after surgery, the heart was excised and fixed with a 10% solution of formalin in PBS. The heart was embedded in paraffin and cut serially from the apex to the base. The infarct size was expressed as a percentage of the total LV wall circumference from each of the 3 LV sections.

To assess polymorphonuclear neutrophil infiltration, the sections stained with hematoxylin and eosin were observed microscopically, and the numbers of polymorphonuclear neutrophils were counted in 3 to 5 fields for each heart.

To assess the degree of fibrosis in the noninfarct area, the sections were stained with Azan-Mallory stain. In each section, high-power fields at a magnification of ×400 were scanned with computer-assisted videodensitometry, and the images from at least 10 fields for each heart were analyzed as described previously. The collagen volume fraction was obtained by calculating the mean ratio of the positive cells to total nucleated cells in infarct area.

To assess polymorphonuclear neutrophil infiltration, the sections of the infarcted area were evaluated to assess the degree of accumulated polymorphonuclear neutrophils and macrophages, because it has been reported that these are important sources for chemokine and cytokine induction or production. After myocardial infarction, polymorphonuclear neutrophils and activated Mac-3–positive macrophages were infiltrated into the infarct area in WT and KO mice (data not shown). However, morphometric analysis demonstrated

Immunohistochemistry

The sections embedded in paraffin were incubated with primary antibody specific to activated macrophages (Mac-3 monoclonal antibody, BD Pharmingen) for 16 hours, followed by biotinylated secondary antibody (biotinylated anti-rat antibody, Jackson ImmunoResearch Laboratories, Inc) and then by avidin-conjugated horseradish peroxidase. Color development was performed with 3,3′-diaminobenzidine, and slides were counterstained with hematoxylin. The degree of macrophage emigration was expressed as a percentage of Mac-3–positive cells to total nucleated cells in infarct area.

Extraction of Total RNAs and Reverse Transcriptase–Polymerase Chain Reaction

Total RNAs were extracted from noninfarct LV using TRIzol (Invitrogen, Tokyo, Japan) and subjected to reverse transcriptase reverse transcriptase–polymerase chain reaction (RT-PCR) amplification using oligonucleotide primers. PCR primers for transforming growth factor–β1 (TGF-β1) were 5′-ACCGAAACAAGC-CCATCTAT-3′ (forward) and 5′-GTAACGGAGAATT-GTTCG-3′ (reverse), primers for collagen type I were 5′-GTGAACTTGGCAAAAACAGT-3′ (forward) and 5′-CTG-GAGACCGAGAAGGCCAC-3′ (reverse), and primers for GAPDH were 5′-ACTCCACTCACGGAATTCAC-3′ (forward) and 5′-AGGGGCGGAGATGATGACC-3′ (reverse). The PCR products were fractionated on a 1% to 2% agarose gel and visualized by ethidium bromide staining. The intensities of the bands were normalized for GAPDH and were expressed as fold increase over control WT.

Statistical Analysis

All values are expressed as mean ± SEM. Comparisons of the histological analysis between WT and KO mice were made by Student’s unpaired t test. To compare body weight, LV weight, lung weight, LVEDD, LVESD, %FS, and RT-PCR data, 1-way ANOVA followed by post hoc test was performed. Survival curves after myocardial infarction were created by the Kaplan-Meier method and compared by log-lank test. Statistical significance was accepted at a value of P<0.05.

Results

Heart Failure and Ventricular Remodeling After Myocardial Infarction

There were no differences in body weight among 4 groups before and after myocardial infarction. Heart weight and LV weight after myocardial infarction were lower in KO mice than in WT mice (Table). Furthermore, lung weight was also lower in KO mice than in WT mice (Table).

Survival Rates and Infarct Size

The survival rates after myocardial infarction were compared up to 4 weeks between WT (n=107) and KO (n=80) mice, as shown in Figure 1. The rate of death during surgery was not different between WT and KO mice (14% and 15%, respectively). The survival rate up to 4 weeks after myocardial infarction was significantly higher in KO mice than in WT mice (65% versus 43%, P<0.03). However, infarct size at 4 weeks after coronary ligation was similar between WT and KO mice, as shown in Figure 2 (55.6±4.2% and 53.2±3.7%, respectively).

Assessment of Inflammatory Cell Infiltration Into the Infarct Area

At 3 days and 1 week after myocardial infarction, histological sections of the infarcted area were evaluated to assess the degree of accumulated polymorphonuclear neutrophils and macrophages, because it has been reported that these are important sources for chemokine and cytokine induction or production. After myocardial infarction, polymorphonuclear neutrophils and activated Mac-3–positive macrophages were infiltrated into the infarct area in WT and KO mice (data not shown). However, morphometric analysis demonstrated...
that there was no significant difference in the degree of inflammatory cell infiltration between WT and KO mice.

The numbers of polymorphonuclear neutrophil infiltration (per 10² μm²) were 41.2±3.5 and 41.0±2.9 at 3 days after myocardial infarction and 54.3±2.8 and 55.0±3.3 at 1 week after myocardial infarction in WT and KO mice, respectively. The degrees of Mac-3-positive macrophage infiltration were 41.0±4.0% and 41.1±3.4% at 3 days and 45.1±4.0% and 44.2±3.3% at 1 week after myocardial infarction in WT and KO mice, respectively.

**Echocardiographic Measurements**

Echocardiography was performed at baseline control and at 1 or 4 weeks after myocardial infarction in WT and KO mice. Under anesthesia, the heart rate was similar between WT and KO mice at each point (data not shown). The representative M-mode echocardiograms of WT and KO mice at baseline and 1 and 4 weeks after myocardial infarction are shown in Figure 3. Before coronary ligation, LVEDD, LVESD, and %FS were similar between WT and KO mice (baseline). LV dilatation after myocardial infarction was evident in both WT and KO mice; however, KO mice showed less ventricular dilatation. LVEDD and LVESD were smaller in KO mice than in WT mice at 1 and 4 weeks after operation, as shown in Figure 4, A and B. LVEDDs (mm) were 4.18±0.11 (n=13) and 4.61±0.17 (n=12) at 1 week (P<0.05) and 4.64±0.13 (n=15) and 5.13±0.16 (n=13) at 4 weeks (P<0.01) in KO mice and WT mice, respectively. LVESDs (mm) were 3.04±0.15 and 3.74±0.19 (P<0.05) at 1 week and 3.53±0.16 and 4.30±0.23 (P<0.01) at 4 weeks in KO mice and WT mice, respectively. Moreover, %FS was significantly higher at 1 and 4 weeks after surgery in KO mice than in WT mice, as shown in Figure 4C (27.7±2.5% versus 21.2±2.6%, P<0.05, at 1 week and 24.3±2.0% versus 16.6±2.5%, P<0.01, at 4 weeks). Both WT and KO mice showed linear relationships (r=0.68 and r=0.62, respectively, P<0.05) between infarct size and %FS at 4 weeks (Figure 4D).

**Assessment of Myocardial Fibrosis in Noninfarct Area**

Morphological changes at the noninfarct area were similar between WT and KO mice at 3 days after myocardial infarction (data not shown). At 1 and 4 weeks after surgery, cardiomyocytes showed mild architectural alteration in KO mice compared with WT mice (data not shown). Myocyte cross-sectional area was not different between KO mice (331.2±48.8 μm²) and WT mice (407.6±66.2 μm²). In KO mice, the degrees of myocardial fibrosis at the noninfarcted interstitial area (0.34±0.06 versus 1.00±0.22, P<0.01) and perivascular area (0.35±0.04 versus 1.00±0.17, P<0.01) were much less than in WT mice at 4 weeks after myocardial infarction (Figure 5, A–E).

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**Weight Parameters**

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<th>KO Control</th>
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<td>253±28*</td>
<td>185±20†‡</td>
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</table>

BW indicates body weight. All values are expressed as mean±SEM of 8 mice per group.

*P<0.01 and †P<0.05 vs control in same strain.

‡P<0.05 and §P<0.01 vs WT at each time point.

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**Figure 1.** Survival curves after myocardial infarction in WT and KO mice. Survival curves were created by Kaplan-Meier method and compared by log-rank test. Percentages of surviving WT and KO mice were plotted. At 4 weeks after myocardial infarction, survival rates were significantly higher in KO mice than in WT mice.

**Figure 2.** Histological analysis of infarct area after myocardial infarction. Top and bottom, Azan-Mallory-stained LV transverse sections in WT and KO mice, respectively.
To investigate whether these morphological observations were accompanied by alterations in gene expression relevant to fibrotic changes, we measured mRNA levels of TGF-β1 and collagen type 1 at 1 week after myocardial infarction. RT-PCR revealed that TGF-β1 and collagen type 1 mRNA obtained from noninfarcted myocardium at 1 week after myocardial infarction in both WT and KO mice. However, these changes in TGF-β1 and collagen type 1 mRNA expressions after myocardial infarction were attenuated in KO mice compared with WT mice, as shown in Figure 6A. When normalized for GAPDH mRNA, the expressions of TGF-β1 and collagen type 1 in KO mice were reduced by 34.2% (P<0.01) and 38.6% (P<0.05), respectively, compared with WT mice (Figure 6B).

Discussion

In the present study, we examined whether ventricular remodeling after myocardial infarction was altered in TLR-2–KO mice. TLR-2–KO mice demonstrated higher survival rate and less ventricular remodeling evaluated by echocardiography, associated with fewer pathological changes and decreased expression of TGF-β1 and collagen type 1 mRNA. The present study provided the direct evidence for involvement of TLR-2–mediated signaling pathway in ventricular remodeling after myocardial infarction.

Previous studies have documented that TLRs are expressed in many organs, such as spleen, brain, kidney, liver, lung, and heart, and act differentially in a wider pattern of tissue expression. In an in vivo study, Nemoto et al demonstrated that myocardial TLRs are involved in cardiac dysfunc-

**Figure 3.** Representative M-mode traces in WT mice (A, C, and E) and KO mice (B, D, and F) at baseline, 1 week, and 4 weeks after myocardial infarction, respectively. AW, PW, and LVD indicate anterior wall, posterior wall, and LV internal dimension, respectively. After myocardial infarction, LVD was increased in both animals; however, LVEDD and LVESD at 4 weeks after myocardial infarction in KO mice were clearly smaller than in WT mice.

**Figure 4.** Quantified data for echocardiographic measurements. A through C, LVEDD and LVESD at 4 weeks after myocardial infarction in KO mice were reduced significantly, by 9.6% and 18.0%, respectively, compared with WT mice. All data were obtained from 12 to 15 animals in each group. Although %FS was reduced in both WT and KO mice after myocardial infarction, %FS in KO mice was significantly higher than that in WT mice. D, Both WT mice (n=12) and KO mice (n=10) showed linear relationships (r=0.68 and r=0.62, respectively, P<0.05) between infarct size and %FS. At baseline, LVEDD, LVESD, and %FS in WT mice (WT-con) and KO mice (KO-con) were similar. *P<0.05 and **P<0.01 vs WT at each time point; ‡P<0.01 vs control in same strain. 1w and 4w indicate 1 week and 4 weeks after myocardial infarction, respectively. Open symbols and filled symbols indicate WT and KO mice, respectively.
tion during inflammatory shock. However, the potential role of TLR-2 in ventricular remodeling after myocardial infarction is still unknown. In this study, at 1 and 4 weeks after myocardial infarction, LVEDD and LVESD were smaller in TLR-2–KO mice than in WT mice. Furthermore, %FS was markedly higher in TLR-2–KO mice than WT mice, which was associated with a better survival rate in KO mice. These results suggest that TLR-2 plays an important role in ventricular remodeling after myocardial infarction.

Several studies have demonstrated that myocardial infarction is associated with inflammatory responses, which can extend myocardial injury.20 Reduced numbers of neutrophils by antineutrophil drugs17 or neutrophil filters22 can reduce myocardial injury after ischemia. Furthermore, Li et al23 demonstrated that necrotic cells activate NF-κB DNA binding activity through the TLR-2–mediated signaling pathway in cardiomyocytes and Chinese hamster ovary fibroblasts. In this study, cardiac dysfunction evaluated by echocardiography at 4 weeks after myocardial infarction was abrogated in TLR-2–KO mice compared with WT mice. Therefore, ventricular remodeling after myocardial infarction was modulated through TLR-2.

In this study, histological examinations demonstrated that myocardial fibrosis was decreased significantly in KO mice at noninfarct area. Furthermore, gene expressions of TGF-β1 and collagen type 1, which were responsible for cardiac fibrosis,12,25 were lower in KO mice than WT mice after

![Figure 5. Histological changes in noninfarct area at 4 weeks after myocardial infarction in WT and KO mice. A through D, Azan-Mallory–stained sections in noninfarct area. A and B demonstrate interstitial, and C and D perivascular, sections. E, In KO mice, myocardial fibrosis in interstitial and perivascular areas at 4 weeks after myocardial infarction were attenuated compared with WT mice.](image)

![Figure 6. Quantification of gene expression related to fibrotic changes. A, RT-PCR was performed to evaluate TGF-β1 and collagen type 1 mRNA expression after myocardial infarction. B, Group data for relative band intensity of TGF-β1, collagen type 1, and GAPDH. At baseline, TGF-β1, and collagen type 1 mRNA expressions were similar between WT (WT-con) and KO (KO-con) mice. At 1 week after myocardial infarction, myocardial expressions of TGF-β1 and collagen type 1 mRNA were increased in WT (WT-MI) and KO (KO-MI) mice. However, induced gene expressions were attenuated in KO mice compared with WT mice. All data are expressed as mean±SEM. *P<0.05 and **P<0.01 vs WT at each time point. †P<0.05 and ‡P<0.01 vs control in same strain.](image)
myocardial infarction. These data support the concept that modulation of myocardial fibrosis in the noninfarct area is one of the possible mechanisms for attenuation of ventricular remodeling in TLR-2–KO mice.

We reported the preservation of cardiac function, increased survival rate, and attenuation of myocardial fibrosis after myocardial infarction in TLR-2–KO mice. In conclusion, this study demonstrates the first evidence that ventricular remodeling after myocardial infarction occurs, at least in part, through the TLR-2–mediated pathway. These results will allow us a novel approach to investigate the pathogenesis of ventricular remodeling after myocardial infarction.

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References

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