Display of Fas Ligand Protein on Cardiac Vasculature as a Novel Means of Regulating Allograft Rejection

Nadir Askenasy, MD, PhD*; Esma S. Yolcu, PhD*; Zhiliang Wang, MD; Haval Shirwan, PhD

Background—Fas ligand (FasL) is a potent death-inducing molecule with important functions in immune homeostasis and tolerance to self-antigens. The complex biological activities of FasL and its inefficient expression using conventional gene transfer approaches limit its use for immunomodulation to prevent allograft rejection. We have recently generated a chimeric FasL with core streptavidin (SA-FasL) with potent apoptotic activity and designed a novel approach to display it on the surface of several cell types via biotinylation. We herein tested whether SA-FasL can also be displayed on vascular endothelial cells in the heart and examined its effect on graft survival after transplantation into syngeneic and allogeneic hosts.

Methods and Results—SA-FasL was efficiently displayed on the vasculature of BALB/c hearts with a half-life of 9 days in vivo. Transplantation of hearts displaying SA-FasL into syngeneic hosts resulted in indefinite graft survival without detectable toxicity to the grafts and hosts. In contrast, transplantation of allogeneic C57BL/10 hearts displaying SA-FasL into BALB/c recipients resulted in graft rejection, but in a delayed fashion as compared with control hearts (mean survival time=17.4±5 versus 9.6±1 days). Allograft survival was further extended to 21±2.6 and 24±3 days (P<0.05) by intravenous treatment of graft recipients with 1 dose of SA-FasL–decorated donor splenocytes on days 2 and 6 after transplantation, respectively.

Conclusions—This study shows for the first time that exogenous proteins can be displayed on the endothelium of solid organs for therapeutic purposes. This approach provides a convenient and rapid means of displaying exogenous proteins on the surface of cells, tissues, and solid organs, with broad research and therapeutic implications. (Circulation. 2003;107:1525-1531.)

Key Words: immunology ■ proteins ■ apoptosis ■ transplantation ■ gene therapy

The critical role that the Fas/Fas ligand (FasL) interaction plays in immune homeostasis, self-tolerance, and immune privilege in vital organs led to attempts to use these molecules to prevent allograft rejection and induce tolerance to solid organ grafts. Conceptually, the presentation of antotgens to T cells in the presence of FasL may lead to the apoptosis of activated cells and may block alloreactive responses. The process of activation-induced cell death (AICD) involves interaction of FasL with the Fas receptor expressed on the surface of activated lymphocytes, which initiates a cascade of events that culminates in the activation of caspases and cell apoptosis. Several studies demonstrated effective blockade of alloreactive responses and survival of allogeneic liver, kidney, thyroid, and pancreatic islets using FasL expressed via genetic modifications as an immunomodulatory approach. In contrast, transgenic expression of wild type FasL in pancreatic β-cells and hearts resulted in their accelerated rejection by allogeneic hosts. The hyperacute rejection of these allografts was accompanied by heavy inflammatory infiltrates, which were likely recruited into the grafts by the soluble FasL, generated from the membrane-bound form by matrix metalloproteinases. Soluble FasL has dual functions that include an anti-apoptotic effect, which is caused by competition with the membrane-bound FasL for Fas binding, and chemotaxis of neutrophils, which initiates and perpetuates inflammation. Additional factors, such as the expression level of FasL, the kinetics of expression, the nature of the microenvironment where FasL is expressed, and the lack of mechanisms that regulate FasL function in various tissues may contribute to the opposing effects of FasL on the immune responses, ie, immune activation versus immune blockade.

We reasoned that the transient cell-surface display of a non-cleavable form of FasL that lacks the metalloproteinase...
cleavage site and exerts a more potent apoptotic function than wFasL might facilitate its effective use as an immunomodulatory molecule to regulate allograft rejection. Furthermore, because the clinical application of gene therapy is limited by the low efficiency of gene delivery systems, the modest capacity to control gene expression, difficulties in targeting to the desired tissues, and safety issues, we have recently developed a novel method, designated as ProtEx, that allows for rapid and durable display of exogenous proteins on the surface of isolated cells. This technique relies on the generation of chimeric proteins with core streptavidin/avidin and their attachment to biotinylated cells under physiological conditions. In this study, we tested whether a chimeric molecule composed of the extracellular portion of rat FasL lacking the metalloproteinase site and of core streptavidin (SA-FasL) can be effectively displayed on the heart graft vasculature, and tested the effect of SA-FasL on graft survival in syngeneic and allogeneic hosts. Selective display of FasL on the surface of vascular endothelium may be particularly relevant to the efficient use of this molecule to prevent heart rejection because the endothelium is resistant to Fas/FasL-mediated apoptosis, whereas cardiomyocytes are extremely sensitive to this apoptotic pathway. We report here for the first time that cardiac vasculature can be modified with an exogenous chimeric protein under conditions of hypothermic extracorporeal preservation without detectable toxicity, and such grafts showed indefinite survival in syngeneic and prolonged survival in allogeneic hosts.

**Methods**

**Animals**

B10 (C57BL/10Sn, H-2°) and BALB/c (H-2°) mice were purchased from Jackson Laboratories (Bar Harbor, Me), housed in a pathogen-free facility, and treated in accordance with the National Institutes of Health Guide for the Care and Use of Animals under supervision of the Institutional Animal Care and Use Committee.

**Heart Preparation**

Mice (8 to 10 weeks old) were anesthetized by intraperitoneal injection of ketamine/xylazine (100/10 μg/g), and 20 μL of heparin sulfate was administered. Aortas of the donor hearts were cannulated and hypothermic cardiac arrest was induced by infusion of ice-cold medium. Retrograde coronary perfusion at normothermia (37°C) was performed in a Langendorff preparation using double-walled, water-jacketed containers and tubing to maintain constant temperature.

The hearts were paced at 8 Hz, diastolic pressure was set at 5 to 10 mm Hg, and the left ventricular developed pressure (LVPD) was measured as the difference between the peak systolic and diastolic pressures (PowerLab, ADInstruments). Cold cardiac arrest was induced by infusion of cardioplegic solution at 6°C concomitant with topical cooling, and at hypothermia the hearts were perfused with a miniperistaltic pump (P720, Instech) at a rate of 0.05 mL/min. The perfusion medium was a modified Krebs Henseleit (KH) solution, containing (in mmol/L): 121 NaCl, 5.5 KCl, 1.2 MgSO₄, 2.5 CaCl₂, 23 NaHCO₃, 11 glucose, 5 pyruvate, and 0.05 hexanoate (Sigma Chemical Co). The solution was supplemented with 16 mmol/L MgSO₄ for cardioplegia (KH-Mg), 5 μmol/L EZ-Link Sulfo-NHS-LC-Biotin (KH-biotin, Pierce), 0.5 μg/mL streptavidin labeled with fluorescein isothiocyanate (FITC) (KH-SA, Zymed), 350 ng/mL chimeric SA-FasL protein, or with 350 ng/mL S2 supernatant containing a dysfunctional form of FasL (KH-S2). Media were aerated with O₂/CO₂ (95/5) to obtain a measured pH of 7.3.

**Construction of SA-FasL Chimeric Gene and Expression in Drosophila S2 Cells**

A recombinant gene composed of core streptavidin and the extracellular portion of FasL lacking the metalloproteinase cleavage site (SA-FasL) was constructed using standard molecular biology techniques as recently described. Briefly, genomic DNA was isolated from *Streptomyces avidinii* and amplified using specific primers for the 5'- and 3'-end of core streptavidin in polymerase chain reaction. The extracellular domain of rat FasL lacking the metalloproteinase cleavage site was cloned using a similar strategy. The DNA inserts were then subcloned into the DES expression system (Invitrogen). Supernatant from S2 cells transfected with an expression vector containing FasL cDNA with a 5' frame-shift mutation (S2 proteins) or core streptavidin served as negative controls.

**Display of SA-FasL on Vascular Endothelium**

At normothermia (protocol 1), perfusion included 20 minutes with KH to allow stabilization of cardiac function, 20 minutes KH-biotin, 10 minutes KH for biotin washout, 20 minutes KH-SA, or 350 ng/mL SA-FasL, or 350 ng/mL S2 proteins, and 10 minutes KH for washout. At hypothermia (protocol 2), the hearts were arrested with KH-Mg and perfused at a rate of 50 μL/min as follows: 4 minutes KH, 8 minutes KH-biotin, 4 minutes KH for biotin washout, 8 minutes KH-SA, 350 ng/mL SA-FasL, or 350 ng/mL S2 proteins, and 4 minutes KH for washout.

**Decoration of Splenocytes With SA-FasL**

Spleens were harvested from B10 donors and processed for single cell suspension. Splenocytes were then biotinylated in 15 μmol/L of freshly prepared EZ-Link Sulfo-NHS-LC-Biotin for 30 minutes at room temperature. Cells were washed twice and resuspended in PBS supplemented with 50 to 100 ng SA-FasL per 10⁶ cells/mL. After incubation on ice for 20 minutes with gentle mixing, the cells were washed extensively and analyzed in flow cytometry for the cell surface SA-FasL protein using MFL-4 mAb conjugated to FITC. Four million irradiated cells (2000 rad) were injected intravenously into graft recipients on days 2 or 6 after transplantation.

**Statistical Analysis**

Data are presented as means±SD for each experimental protocol. The experimental variables were evaluated for reproducibility by 1-way ANOVA. Differences between the experimental protocols were evaluated with a post hoc Scheffe test, with P<0.05 selected for level of statistical significance.

**Results**

**Modification of Cardiac Endothelium With Biotin and Exogenous Proteins Without Detectable Toxicity**

Modification of organs with exogenous proteins for therapeutic purposes is contingent on the preservation of organ function after manipulation. We first evaluated the effect of
modification with biotin and decoration with streptavidin or SA-FasL on the function of B10 hearts perfused at normothermia (protocol 1). Control hearts (n=5) had baseline LVDP of 89±6 mm Hg and coronary flow rates of 1.4±0.1 mL/min. Biotinylation and decoration with streptavidin-FITC (Figure 1), S2 proteins, or SA-FasL (n=5 in each group) resulted in an insignificant decrease in LVDP to 81±5 mm Hg, without detectable changes in coronary flow rates.

We next assessed the feasibility of displaying SA-FasL on vascular endothelium under hypothermic extracorporeal heart preservation conditions. Four groups of B10 mouse hearts (control, biotin-SA-FITC, biotin-S2 proteins, and biotin-SA-FasL; n=5 in each group) were perfused at 6°C (protocol 2). There were no major differences between control and test groups with respect to LVDP and coronary flow rates on reperfusion at normothermia. Perfusion with biotin resulted in effective biotinylation of the heart vascular endothelium as demonstrated by binding of streptavidin-FITC (n=5; Figure 1). The biotinylated endothelium was effectively decorated with SA-FasL, as detected by a FITC-MFL-4 mAb using fluorescence microscopy (n=5; Figure 2). In contrast, hearts modified with biotin and treated with S2 proteins scored negative for the binding of MFL4 mAb (n=4; Figure 2d). Taken together, these data demonstrate that SA-FasL can be displayed on the surface of heart vascular endothelium without detectable acute toxicity under hypothermic organ preservation conditions.

**Decoration of Vascular Endothelium With SA-FasL Prolongs Graft Survival**

We next tested whether hearts manipulated ex vivo at hypothermia can function after transplantation into syngeneic hosts. Hearts that were perfused with KH solution only survived indefinitely in syngeneic BALB/c mice without detectable acute or chronic toxicity (n=7; mean survival time [MST] >100 days; Table 1). Control hearts that were modified with biotin and decorated with the SA-FasL protein (protocol 2) also survived indefinitely in syngeneic recipients (n=7; MST >100 days). Importantly, SA-FasL on the graft vasculature was not toxic to the recipient liver as determined by the normal levels of alanine aminotransferase (25±9 IU/mL) and alkaline phosphatase (48±13 IU/mL) measured on days 7 and 21 after transplantation. Furthermore, livers from these animals did not show significant immunostaining for apoptosis as compared with naïve animals (data not shown).

**Figure 1.** Modification of heart endothelium with biotin. The images present the surface of tissue cryosections as observed by fluorescence microscopy (Axioplan, C. Zeiss) at 10×. Hearts (n=5) were perfused at hypothermia with Krebs-Henselelt solution (KH) at a rate of 0.05 mL/min in 5 stages: 4 minutes KH, 8 minutes KH containing 5 μmol/L biotin, 4 minutes KH, 8 minutes KH containing 0.5 μg/mL streptavidin-FITC, and 4 minutes KH. Tissues were frozen and then sectioned. a, Brightfield image of the frozen tissue surface. b, Fluorescence of streptavidin-FITC bound to the biotinylated endothelium is demonstrated on the surface of the section. The image was pseudo-colored to simulate the real hues. A cutoff arteriole shows no fluorescence (blue arrow), and reflected endothelium injured by the sectioning process shows bright fluorescence (yellow arrowheads). Measurements of the fluorescence intensity showed 70±3 arbitrary units (at the location demarcated 4 to 6) as compared with a background fluorescence of 7.5±3 U (at the location demarcated 1 to 3). c, The trajectory of the coronary artery is demarcated by superposition of the fluorescence (b) image over the brightfield image (a). d, Unmodified hearts perfused with streptavidin-FITC served as controls without a detectable signal.

**Figure 2.** The display of SA-FasL on mouse heart endothelium. Hearts were modified with biotin by perfusion and decorated with 350 ng/mL SA-FasL (n=5) or S2 proteins (n=4) as detailed in the legend to Figure 1. FITC-labeled MFL-4 mAb against rat FasL was used for detection. Tissues were snap-frozen and sequential cryosections were imaged with an Axioplan fluorescence microscope. a, Brightfield image (10×) presents the surface of the sectioned muscle. b, Fluorescence image (10×) of an oblique section of the cardiac apex after biotinylation and decoration with SA-FasL protein demonstrates staining of the left descending coronary artery endothelium with FITC-MFL4 mAb (blue arrows). An arteriole and its territory of supply are demarcated in red. c, An enlarged fluorescence image (acquired at 20×) of a section through the coronary artery decorated with SA-FasL. d, Fluorescence image (20×) of a section of heart modified with biotin, decorated with S2 proteins, and stained with FITC-MFL4 mAb served as the background control without detectable fluorescence.
Extended Endothelial Display of SA-FasL In Vivo in the Absence of Neutrophil Infiltration

The rejection of allogeneic grafts displaying SA-FasL might have been caused by the loss of protein from the endothelium, which resulted in the restoration of alloreactive immune responses over time, and/or by the recruitment of inflammatory infiltrates. To assess the persistence of SA-FasL on graft vasculature, hearts were excised at different times after transplantation and stained with FITC-MFL-4 mAb. The fluorescence intensity ratio (versus background) decreased from 9.3 ± 2.2 before transplantation to 5.7 ± 1.4 and 3.2 ± 0.7 on days 6 and 12 after transplantation, respectively (Figure 4). A linear curve fit showed a half-life of 9 days for SA-FasL on vascular endothelium, suggesting that the shear stress of blood flow in the graft vasculature did not significantly contribute to the turnover of biotin-streptavidin complex from the endothelial surface. Persistence of SA-FasL protein for weeks on the endothelium is consistent with our previous observations that SA-FasL may persist on the cell surface for weeks in vitro and in vivo.

To provide evidence that the observed graft prolongation was due to FasL-induced apoptosis in Fas-expressing alloreactive lymphocytes, aortic endothelial cells decorated with SA-FasL were co-incubated with A-20 cells (a Fas+ mouse B lymphoma line) for 18 hours. Apoptosis was determined using propidium iodide and annexin V-FITC in flow cytometry in 4 independent cultures. There was >40% cell death of A-20 cells in cultures containing SA-FasL–decorated endothelial cells as compared with <5% cell death in cultures containing biotinylated endothelial cells with and without core streptavidin as control (Figure 3). Furthermore, we did not observe significant endothelial cell death in cultures containing SA-FasL as compared with control cultures (data not shown). This is consistent with previous studies demonstrating that endothelial cells are resistant to FasL-mediated killing.

<table>
<thead>
<tr>
<th>Group</th>
<th>N</th>
<th>Recipient</th>
<th>Donor</th>
<th>Treatment</th>
<th>Mean Survival</th>
<th>Time, d</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>7</td>
<td>BALB/c</td>
<td>BALB/c</td>
<td></td>
<td>100</td>
<td></td>
</tr>
<tr>
<td>B</td>
<td>7</td>
<td>BALB/c</td>
<td>BALB/c</td>
<td>Perfusion*</td>
<td>100</td>
<td></td>
</tr>
<tr>
<td>C</td>
<td>7</td>
<td>BALB/c</td>
<td>BALB/c</td>
<td>Biotin-FasL†</td>
<td>&gt;100</td>
<td></td>
</tr>
<tr>
<td>D</td>
<td>7</td>
<td>BALB/c</td>
<td>B10</td>
<td></td>
<td>&gt;100</td>
<td></td>
</tr>
<tr>
<td>E</td>
<td>6</td>
<td>BALB/c</td>
<td>B10</td>
<td>Biotin-S2‡</td>
<td>8.6 ± 1.4</td>
<td></td>
</tr>
<tr>
<td>F</td>
<td>7</td>
<td>BALB/c</td>
<td>B10</td>
<td>Biotin-FasL†</td>
<td>17.4 ± 5‡</td>
<td></td>
</tr>
</tbody>
</table>

*Cardiac grafts were transplanted after perfusion at hypothermia with Krebs Henseleit solution.
†Hearts perfused with biotin and SA-FasL.
‡Hearts perfused with biotin and S2 proteins.

Figure 3. Endothelial cells decorated with SA-FasL induce apoptosis in Fas+ A-20 lymphoma cell line. A-20 cells were co-incubated with biotinylated cells (a, None), cells decorated with core streptavidin (b, SA), and cells decorated with SA-FasL (c, FasL) for 18 hours. The cells were then harvested and analyzed for apoptosis using PI and annexin V-FITC, and A-20 cells were selected by gating on B220 positive cells (anti-B220 mAb-APC) with the use of flow cytometry.

Immunomodulation of Graft Recipients With Donor Splenocytes Decorated With the SA-FasL Protein Extends Graft Survival

The apparent late onset of acute rejection in the presence of SA-FasL on vascular endothelium led us to test whether
additional immunomodulation of graft recipients with splenocytes expressing SA-FasL further delayed rejection. Injection of naïve (n=5) or biotinylated donor splenocytes treated with S2 proteins (n=5) on day 2 after transplantation into recipients of unmanipulated heart allografts did not affect the tempo of graft rejection (MST=8±1.4. days and 9.5±2 days, respectively; Table 2). Acute allograft rejection was also unaffected by injection of donor splenocytes decorated with SA-FasL on day 2 after transplantation of unmanipulated hearts (n=5; MST=10.2±1.3 days), implying that the direct display of SA-FasL on the heart endothelium may be critical to the prolonged survival. Consistent with this notion, injection of SA-FasL decorated splenocytes into the recipients of SA-FasL–decorated hearts on day 2 after transplantation resulted in prolonged allograft survival to 21±2.6 days (n=7). This rejection time was further extended to 24±3 days when donor splenocytes decorated with SA-FasL were injected 6 days after transplantation (n=7; P<0.05 versus MST=17.4±5 days for SA-FasL–decorated hearts). By contrast, injection of the recipients of SA-FasL–decorated hearts with donor unmanipulated splenocytes 2 days after transplantation abolished the SA-FasL–mediated prolonged survival (n=5, MST=10.8±2 days, P<0.05 versus MST=17.4±5 days for SA-FasL–decorated hearts only).

**Discussion**

We present a novel approach that exploits the direct display of exogenous proteins on vascular endothelium of solid organs for immunotherapeutic purposes. This approach is unique and differs in several aspects from conventional methods that utilize gene transfer or systemic administration of biologics for immunomodulation. First, recombinant proteins are directly displayed on the vascular endothelium to avoid the difficulties associated with gene transfer approaches, such as the inefficiency of gene transfer, limited control over the levels of expression, and lack of safety.

Second, the display of proteins is efficiently performed within a short period of time under conditions of extracorporeal organ preservation similar to those used in the clinic. Third, the proteins are selectively introduced into the target tissues or organs to achieve a localized effect, which may prove more efficacious as compared with systemic immunomodulation.

Fourth, the high selectivity of direct protein display on the graft endothelium may avoid toxicities that otherwise would be associated with the introduction of proteins into the parenchymal tissue. For example, the toxicity of FasL for smooth and cardiac muscles is negated when the expression of this death molecule is restricted to the endothelium.

Using FasL as an immunomodulatory molecule, we have demonstrated that the direct display of a non-cleavable isoform on cardiac vascular endothelium results in statistically significant prolongation of graft survival in allogeneic hosts. We did not observe infiltration of neutrophils in hearts

---

**Figure 4.** In vivo kinetics of SA-FasL display on heart graft endothelium as determined by fluorescence imaging of the surface of tissue cryosections. The images (Axioplan, exposure 5 seconds) demonstrate the bifurcation of the left main coronary artery (yellow arrow) into the left anterior descending (LAD) and circumflex (CX) coronary arteries. Hearts displaying SA-FasL were excised 6 (a) and 12 (b) days after transplantation and perfused with FITC-MFL-4 mAb. There was a decrease in fluorescence intensity in the coronary arteries over time. Tissues from biotinylated hearts perfused with S2 proteins and stained with FITC-MFL-4 mAb served as negative controls without detectable fluorescence (data not shown).

**Figure 5.** Histopathological analysis of heart allografts. Sections were stained with hematoxylin and eosin and images were acquired with an Olympus microscope at 10×. a, Syngeneic heart on day 20 after transplantation with normal histology. b, Allogeneic heart modified with biotin and treated with S2 proteins showing heavy inflammatory infiltrates (blue nuclei), edema, and disruption of the myocardial sarcomeres on day 6 after transplantation. c, Allogeneic heart decorated with SA-FasL, reveals scarce inflammatory infiltrates and a well-preserved myocardial structure on day 6 after transplantation. d, Allogeneic heart decorated with SA-FasL shows infiltration of inflammatory cells (blue nuclei) and onset of edema and muscle injury on day 12 after transplantation.
displaying the chimeric protein, which suggests that SA-FasL is not chemotactic for polymorphonuclear cells, as shown in studies that used wFasL21,23 and other forms of non-cleavable FasL.32 Furthermore, the selective display of SA-FasL on the graft vascular endothelium may also prevent intimal hyperplasia, and may therefore ameliorate the development of transplant-associated vasculopathy. Previous reports showed that the expression of wFasL in vascular endothelial cells results in T cell apoptosis33 and reduces inflammatory cell infiltrates,34,35 and both wt and non-cleavable FasL limit intimal hyperplasia.36,37

The studies presented herein mainly provide proof-of-concept for protein-based therapy. Optimization of SA-FasL concentration and the timing of injection of decorated donor splenocytes remain to be further evaluated for the effective use of SA-FasL as a tolerogenic molecule. The inability of SA-FasL to completely prevent graft rejection may be due to the emergence of alloreactive memory cells from non-lymphoid tissues when the concentration of SA-FasL on graft vasculature decreased below the effective threshold.35,36–40 Although SA-FasL in our chimeric protein lacks the putative metallocproteinase site and as such persists on the graft vasculature with a half-life of 9 days in vivo, this may not be sufficient for effective elimination of a large pool of peripheral alloreactive lymphocytes. This notion is consistent with our observations that additional post-transplant immunomodulation of graft recipients with donor splenocytes decorated with SA-FasL further prolongs graft survival. In contrast, immunomodulation with unmanipulated donor splenocytes negates the protective effect exerted by SA-FasL on graft vasculature, plausibly by effectively activating a large pool of peripheral alloreactive lymphocytes that shifts the immunological balance toward rejection. These observations suggest that repeated injection of SA-FasL–decorated donor cells into graft recipient before or after transplantation may serve as an effective approach to induce tolerance. T cells become extremely sensitive to apoptosis by Fas/FasL interaction after going through several rounds of antigenic challenge, approximately 3 days after the initial antigenic stimulation.38 Consistently, repeated pretransplant manipulation of graft recipients with donor dendritic cells or macrophages genetically modified to express wFasL was shown to lead to donor-specific immune non-responsiveness.16,17

Immunomodulation by FasL expression involves selective depletion of alloreactive immune cells, predominantly of CD4+ and CD8+ T cell phenotypes, that express Fas on activation31–33 and are prone to AICD10–12 both in vivo and in vitro.16,17 A similar proapoptotic effect was shown for wFasL expressed on the surface of endothelial cells and vascular endothelium,33–36 and we demonstrate that endothelial cells and splenocytes decorated with non-cleavable FasL are even more potent as a trigger of apoptosis in Fas-positive cells.25 The display of FasL either on the graft itself or in its immediate vicinity aims to create a site of immune privilege that simulates a physiological defense mechanism present in vital organs.1–3 Importantly, recent studies have demonstrated the critical role of localized responses in immune regulation and raise the question whether such responses are not only more efficacious, but also specific, the 2 important characteristics of the adaptive immune system.39,40 We have recently demonstrated that co-transplant of pancreatic islets and SA-FasL–decorated splenocytes under the kidney capsule of allogeneic hosts results in the prevention of graft rejection by non-systemic immunoregulatory mechanisms.25 Our present findings demonstrate the importance of local defense by expression of the FasL chimeric protein on the surface of vascular endothelium to achieve prolonged graft survival.

This study demonstrates for the first time that heart grafts can be directly manipulated to display exogenous proteins on the vascular endothelium under extracorporeal preservation conditions used in the clinics, with no significant toxicity. This method of cell membrane modification with biotin and decoration with proteins chimeric with streptavidin is rapid, practical, and safe as compared with protein expression by gene transfer approaches. Furthermore, localized immune protection with protein display not only avoids adverse side effects associated with systemic therapy, but may also prove more efficacious because the target tissue is the site where the effector immunological reactions take place. Hearts modified to display SA-FasL on the vascular endothelium had prolonged survival in allogeneic hosts, demonstrating the immunomodulatory potential of this novel approach. Inasmuch as FasL–mediated apoptosis primarily affects the proinflamma-
tory Th1 cells and spares the antiinflammatory Th2 cells and Fas/Fasl interaction is critical to the development of immunoregulatory cells in selected systems. This molecule has the capacity to induce tolerance. Additional studies, however, are needed to optimize conditions for the use of Fasl, a tolerogenic molecule and study underlying mechanisms of tolerance. This approach, therefore, provides a novel means of effectively and rapidly displaying exogenous proteins with desired functions on the vasculature of solid organs, and as such may serve as an alternative to DNA-based gene therapy with broad research and therapeutic implications.

Acknowledgments
This study was supported in part by grants from American Heart Association (9650229N, 9606386V), the Juvenile Diabetes Research Foundation (1-2001-328), the National Institutes of Health (R21 DK61333, R01 AI47864), the Commonwealth of Kentucky Research Challenge Trust Fund, and the Leah and Edward M. Frankel Bone Marrow Transplantation Trust. The authors thank D. D. Frank for critical review of the manuscript and O. Grimany for his technical assistance in protein purification and Western blot analysis.

References
44. Askenasy et al Fas Ligand Protein Extends Heart Graft Survival 1531

Downloaded from http://circ.ahajournals.org/ by guest on July 25, 2017
Display of Fas Ligand Protein on Cardiac Vasculature as a Novel Means of Regulating Allograft Rejection
Nadir Askenasy, Esma S. Yolcu, Zhiliang Wang and Haval Shirwan

Circulation. 2003;107:1525-1531; originally published online March 17, 2003;
doi: 10.1161/01.CIR.0000064893.96179.7E
Circulation is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2003 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the
World Wide Web at:
http://circ.ahajournals.org/content/107/11/1525

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Circulation can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Circulation is online at:
http://circ.ahajournals.org/subscriptions/