Myocardial Gene Expression of Regulators of Myocyte Apoptosis and Myocyte Calcium Homeostasis During Hemodynamic Unloading by Ventricular Assist Devices in Patients With End-Stage Heart Failure

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Background—In patients with end-stage heart failure, characterized by an increased susceptibility to cardiomyocyte apoptosis and a labile cardiomyocyte calcium homeostasis, a ventricular assist device (VAD) is implanted for bridging to cardiac transplantation and results in myocardial unloading. Although phenotype changes in the failing heart are assumed to result from hemodynamic overload, the reversibility of these changes under unloading is unknown.

Methods and Results—By use of quantitative reverse-transcription polymerase chain reaction, mRNA expression analyses were performed on left ventricular specimens obtained from 10 nonfailing donor hearts (from 8 patients with dilated cardiomyopathy and 2 patients with coronary heart disease) at the time of VAD implantation and 36 to 169 days later during VAD removal with subsequent cardiac transplantation. In terminally failing hearts before VAD support, left ventricular mRNA analyses revealed increased Pro-ANP, reduced antiapoptotic Bcl-xL and antiapoptotic Fas isoform FasExo6Del, and a decreased ratio of sarcoplasmic reticulum Ca\(^{2+}\)-ATPase per sarcolemmal Na\(^{+}\)-Ca\(^{2+}\) exchanger in comparison with nonfailing ventricles. After VAD unloading, ventricular transcription of Pro-ANP was immediately normalized, and apoptotic DNA fragmentation was attenuated. In patients with dilated cardiomyopathy, mRNAs of Bcl-xL and FasExo6Del/Fas were enhanced depending on time on VAD. The Bcl-xL mRNA level correlated positively with that of the Bcl-xL protein. Transcription of sarcoplasmic reticulum Ca\(^{2+}\)-ATPase/Na\(^{+}\)-Ca\(^{2+}\) exchanger demonstrated recovery in only 4 of 10 patients.

Conclusions—Mechanical support of the failing heart induces a time-dependent change in myocardial gene expression compatible with a decreased susceptibility to apoptosis. (Circulation. 1999;100[suppl II]:II-216–II-223.)

Key Words: apoptosis ■ calcium ■ heart failure ■ heart-assist device

Currently, ventricular assist devices (VADs) are implanted for mechanical support of a failing human heart and function reliably for weeks or months without complications even outside the hospital.\(^1\) A shortage of organ donors for heart transplantation has led to the increased application of VADs for patients in end-stage heart failure (HF); the VAD serves as a bridge to transplantation. This bridging also improves the outcome after the subsequent heart transplantation.\(^2,3\)

The overload-induced distension of the heart is a critical factor promoting the progression of HF.\(^4\) Therefore, it has been postulated that some myocardial recovery should occur in the failing heart hemodynamically unloaded by an implanted VAD, and several clinical reports indicate some ventricular recovery under VAD support.\(^5,6\)

Experimental investigations revealed that mechanical distension of the myocardium induces apoptotic death of cardiomyocytes.\(^7\) Furthermore, apoptosis is demonstrable in ventricles of patients with a terminally failing, overloaded heart,\(^8,9\) which is associated with mRNA downregulation of antiapoptotic regulators, such as Bcl-x\(_L\),\(^10\) and FasExo6Del.\(^11\) In human myocardium, FasExo6Del is the most abundant of several soluble antagonistic isoforms of the apoptosis-triggering surface receptor Fas,\(^12\) and it is assumed to function by competing with Fas for the Fas ligand. In contrast, the antiapoptotic Bcl-2 family member Bcl-x\(_L\) is an intracellular protein localized at the outer mitochondrial membrane.\(^13\) It prevents the apoptosis-associated disruption of the mitochondrial transmembrane potential,\(^14\) the release of cytochrome \(c\) from the mitochondrial intermembrane space, and the cyto-

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Bcl-xL, Bcl-2 and Mcl-1 exert an analogous apoptosis-blocker; ND, not determined; and L/RVAD, left/right VAD.

Enhanced expression of the sarcolemmal Na\(^+\) channel 1 prevents function, whereas their action is antagonized by a reduction of this distension by VAD unload-

Apart from the apoptosis-sensitive myocardial phenotype, the left ventricular expression of proteins involved in the myocyte calcium homeostasis is altered in failing human myocardium. In detail, a decreased expression of the sarcolemmal Na\(^+\)-Ca\(^{2+}\) exchanger in end-stage HF. Therefore, we additionally analyzed the left ventricular mRNA levels of both calcium-regulatory determinants under VAD support of the overloaded heart.

### Methods

**Patient Population**

We obtained left ventricular specimens from 10 male patients undergoing VAD implantation as a bridge to transplantation at the time of surgery and at removal of the VAD with subsequent orthotopic cardiac transplantation. Eight patients suffered from dilated cardiomyopathy (DCM), and 2 patients demonstrated end-stage coronary artery disease (CAD). At the time of VAD implantation, patients were 53±12 years old and clinically characterized as shown in Table 1. The hemodynamic data in Table 1 result from the date when the last echocardiographic determination of the ejection fraction was performed. For left ventricular unloading, either TCI HeartMate (Thermo Cardiosystems, Inc), Novacor (Baxter Health-care Corp, Novacor Division), or Thoratec (Thoratec Laboratories, Inc) assist devices were used. For biventricular support in case of TCI or Novacor use, an additional right ventricular assist device of Thoratec was implanted.

In addition, explanted left ventricular specimens of 22 male and 2 female patients (54±8 years old) from the Halle and Hamburg Cardiac Transplant Program exhibiting end-stage HF and treated without VAD unloading were investigated. They had a mean ejection fraction of 25±15%. Thirty patients suffered from DCM; 11 patients, from CAD. Left ventricular tissues from 10 organ donors (8 men and 2 women, 39±5 years old) served as controls. These hearts were not transplanted for technical reasons. The local ethics commit-

### Table 1. Clinical Data for 10 Male Patients Before Ventricular Support by the Assist Device and Thereafter, Shortly Before Orthotopic Heart Transplantation

<table>
<thead>
<tr>
<th>Patient No.</th>
<th>Age, y</th>
<th>HF</th>
<th>Ejection Fraction, %</th>
<th>Drug Treatment</th>
<th>Time on VAD, d</th>
<th>Cardiac Index, L·min(^{-1})·m(^{-2})</th>
<th>Pulmonary Capillary Wedge Pressure, mm Hg</th>
<th>Mean Aortic Pressure, mm Hg</th>
<th>Mean DNA Fragmentation, RU</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>62</td>
<td>DCM</td>
<td>26</td>
<td>ACE i</td>
<td>Novacor LVAD</td>
<td>36</td>
<td>1.33</td>
<td>25</td>
<td>67</td>
</tr>
<tr>
<td>2</td>
<td>42</td>
<td>DCM</td>
<td>24</td>
<td>ACE i</td>
<td>TCI LVAD</td>
<td>42</td>
<td>1.75</td>
<td>25</td>
<td>69</td>
</tr>
<tr>
<td>3</td>
<td>63</td>
<td>DCM*</td>
<td>25</td>
<td>ACE i</td>
<td>Novacor LVAD</td>
<td>57</td>
<td>2.08</td>
<td>16</td>
<td>59</td>
</tr>
<tr>
<td>4</td>
<td>45</td>
<td>DCM</td>
<td>28</td>
<td>ACE i</td>
<td>Novacor LVAD</td>
<td>64</td>
<td>2.20</td>
<td>28</td>
<td>62</td>
</tr>
<tr>
<td>5</td>
<td>47</td>
<td>DCM</td>
<td>26</td>
<td>ACE i</td>
<td>TCI LVAD</td>
<td>116</td>
<td>1.53</td>
<td>41</td>
<td>87</td>
</tr>
<tr>
<td>6</td>
<td>49</td>
<td>DCM</td>
<td>25</td>
<td>ACE i</td>
<td>Novacor/Thoratec LVAD</td>
<td>117</td>
<td>2.79</td>
<td>34</td>
<td>67</td>
</tr>
<tr>
<td>7</td>
<td>59</td>
<td>DCM†</td>
<td>18</td>
<td>ACE i</td>
<td>Novacor LVAD</td>
<td>140</td>
<td>1.46</td>
<td>36</td>
<td>69</td>
</tr>
<tr>
<td>8</td>
<td>30</td>
<td>DCM</td>
<td>25</td>
<td>ACE i</td>
<td>Novacor LVAD</td>
<td>169</td>
<td>1.36</td>
<td>30</td>
<td>78</td>
</tr>
<tr>
<td>9</td>
<td>62</td>
<td>CAD</td>
<td>28</td>
<td>AT-1 bi</td>
<td>Thoratec/Thoratec LVAD</td>
<td>143</td>
<td>1.90</td>
<td>30</td>
<td>70</td>
</tr>
<tr>
<td>10</td>
<td>68</td>
<td>CAD</td>
<td>15</td>
<td>...</td>
<td>TCI LVAD</td>
<td>144</td>
<td>2.69</td>
<td>27</td>
<td>57</td>
</tr>
<tr>
<td>Mean±SD</td>
<td>53±12</td>
<td>24±4</td>
<td></td>
<td></td>
<td>103±49</td>
<td>1.9±0.5</td>
<td>2.9±0.5</td>
<td>29±7</td>
<td>14±8§</td>
</tr>
</tbody>
</table>

HfX indicates heart transplantation; RU, relative units (integral area of the 200-bp DNA fragment/\(\mu g\) DNA); ACE i, ACE inhibitor; AT-1 bi, angiotensin-1 receptor blocker; ND, not determined; and L/RVAD, left/right VAD.

†With hypertensive heart disease.
‡With diabetes mellitus II.
**DNA Preparation and Gel Electrophoresis**

Genomic DNA was prepared from human left ventricular specimens by use of the Puregene DNA Isolation Kit (Biozym). DNA (1.5 μg) was electrophoretically separated in a SYBR Green–stained agarose gel (Biozym). Apoptotic DNA fragmentation was determined by scanning with a laser densitometer that incorporated an evaluation system (Molecular Dynamics).

**TUNEL Assay**

Terminal deoxynucleotidyltransferase-mediated dUTP-biotin nick end labeling (TUNEL)-positive cardiomyocytes were detected in left ventricular cryosections by using the protocol described in the Apoptaq Peroxidase Kit (Oncor). Positive and negative control sections were included. Microscopic evaluations were performed by using an IMT-2 inverted research microscope (Olympus).

**Construction of cRNA Standards and Competitive Reverse-Transcription Polymerase Chain Reaction**

The standard cRNA for each human gene under study was constructed by introducing a definite deletion of 100 bp into the respective cDNA target molecules of the polymerase chain reaction (PCR) amplification. Each standard cDNA was cloned into the pCR-Script SK+ plasmid (Stratagene) and in vitro–transcribed (PCR) amplification. Each standard cDNA was cloned into the respective cRNA standard molecules was added to 4 RT reactions containing the same amount of total RNA (Table 2) and simultaneously reverse-transcribed by using SuperScript Plus reverse transcriptase (GIBCO-BRL). Total RNA was isolated from left ventricular tissues was extracted in lysis buffer (10 mmol/L Tris-HCl [pH 7.4], 1% SDS, and complete protease inhibitor [Boehringer]) by homogenization and subsequent incubation at 95°C for 15 minutes. Protein concentration was measured by the Bio-Rad protein assay, and samples containing 80 μg of total protein were mixed with 2× loading buffer (250 mmol/L Tris-HCl [pH 7.4], 20% glycerol, 4% SDS, 40 mmol/L dithiothreitol, 2 mmol/L Na-EDTA, and 0.1% bromophenol blue), boiled for 2 minutes, and loaded onto a 10% SDS polyacrylamide gel. Proteins were electroblotted onto polyvinylidene fluoride membranes (Pall Gelman) in a final volume of 50 μL. After an initial denaturation at 95°C for 2 minutes, PCR cycles were performed in a thermocycler (Biometra Trioblock) corresponding to the following protocol: 30 seconds at 94°C, 30 seconds at the primer-specific annealing temperature (Table 2), and 30 seconds at 72°C. After extension of the cDNA amplification at 72°C for 5 minutes, the PCR products were electrophoretically separated in an agarose gel containing ethidium bromide (Sigma-Aldrich) for detection. The RT-PCR was evaluated by scanning with a laser densitometer and computer-based imaging system (Molecular Dynamics).

The nucleotide sequences of cDNA clones and PCR fragments were identified by cycle sequencing using the ABI Prism Dye Terminator Cycle Sequencing Kit (Perkin-Elmer) and subsequent automated analysis (Perkin-Elmer, Applied Biosystems Division).

**Western Blot**

After mechanical pulverization in liquid nitrogen, total protein from left ventricular tissues was extracted in lysis buffer (10 mmol/L Tris-HCl [pH 7.4], 1% SDS, and complete protease inhibitor [Boehringer]) by homogenization and subsequent incubation at 95°C for 15 minutes. Protein concentration was measured by the Bio-Rad protein assay, and samples containing 80 μg of total protein were mixed with 2× loading buffer (250 mmol/L Tris-HCl [pH 7.4], 20% glycerol, 4% SDS, 40 mmol/L dithiothreitol, 2 mmol/L Na-EDTA, and 0.1% bromophenol blue), boiled for 2 minutes, and loaded onto a 10% SDS polyacrylamide gel. Proteins were electroblotted onto polyvinylidene fluoride membranes (Pall Gelman), blocked with 6% nonfat dry milk in TBST (200 mmol/L Tris-HCl [pH 7.5], 300 mmol/L NaCl, and 0.1% Tween 20) at room temperature for 1 hour, and incubated with 5 μg/mL of the primary human antibodies for Bcl-xL (rabbit polyclonal, H-62; Santa Cruz Biotechnology) and for Bcl-2 (mouse monoclonal, L24; DAKO) at room temperature for 4 hours. Blots were subsequently washed in TBST and incubated with peroxidase-conjugated antibodies (anti-rabbit or anti-mouse IgG, respectively; Amersham). Bound antibodies were detected with 12 μmol/L of each dNTP, and 2 U of TaqDNA polymerase (Pharmacia) in a final volume of 50 μL. After an initial denaturation at 95°C for 2 minutes, PCR cycles were performed in a thermocycler (Biometra Trioblock) corresponding to the following protocol: 30 seconds at 94°C, 30 seconds at the primer-specific annealing temperature (Table 2), and 30 seconds at 72°C. After extension of the cDNA amplification at 72°C for 5 minutes, the PCR products were electrophoretically separated in an agarose gel containing ethidium bromide (Sigma-Aldrich) for detection. The RT-PCR was evaluated by scanning with a laser densitometer and computer-based imaging system (Molecular Dynamics).
TABLE 3. Molecular Phenotype Characterization of the Left Ventricular Myocardium of Organ Donors and of Patients With End-Stage HF

<table>
<thead>
<tr>
<th>mRNA/RNA</th>
<th>Organ Donors</th>
<th>No VAD Support</th>
<th>Pre-VAD</th>
<th>Post-VAD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Overload-sensitive protein</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pro-ANP, amol/ng</td>
<td>17 ± 20</td>
<td>105 ± 150*</td>
<td>180 ± 207*</td>
<td>28 ± 28§</td>
</tr>
<tr>
<td>Apoptosis-associated proteins</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bak, amol/mg</td>
<td>301 ± 105</td>
<td>170 ± 93*</td>
<td>158 ± 91*</td>
<td>138 ± 52</td>
</tr>
<tr>
<td>Bax, amol/mg</td>
<td>2.1 ± 1.2</td>
<td>2.3 ± 2.3</td>
<td>1.6 ± 0.9</td>
<td>2.1 ± 1.4</td>
</tr>
<tr>
<td>Bcl-2, amol/mg</td>
<td>16.4 ± 9.5</td>
<td>18.1 ± 11.9</td>
<td>11.4 ± 9.0</td>
<td>9.3 ± 5.0</td>
</tr>
<tr>
<td>Bcl-xL, amol/μg</td>
<td>215 ± 92</td>
<td>134 ± 57*</td>
<td>144 ± 47†</td>
<td>176 ± 42§</td>
</tr>
<tr>
<td>Fas, amol/μg</td>
<td>0.9 ± 0.3</td>
<td>1.2 ± 0.5</td>
<td>1.2 ± 0.3†</td>
<td>1.2 ± 0.5</td>
</tr>
<tr>
<td>FasExo6Del, amol/mg</td>
<td>730 ± 420</td>
<td>300 ± 150†</td>
<td>240 ± 120*</td>
<td>250 ± 140</td>
</tr>
<tr>
<td>FasExo6Del/Fas, × 10⁻³</td>
<td>850 ± 420</td>
<td>250 ± 100*</td>
<td>190 ± 90*</td>
<td>230 ± 90</td>
</tr>
<tr>
<td>Mcl-1, amol/ng</td>
<td>3.2 ± 1.5</td>
<td>3.5 ± 1.6</td>
<td>3.5 ± 0.8</td>
<td>4.2 ± 1.1§</td>
</tr>
<tr>
<td>Apoptotic DNA fragmentation, RU</td>
<td>0.13 ± 0.11</td>
<td>0.35 ± 0.23*</td>
<td>0.47 ± 0.14*</td>
<td>0.34 ± 0.21\†</td>
</tr>
<tr>
<td>Heart-specific functional proteins</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SR Ca²⁺-ATPase, amol/pg</td>
<td>91 ± 41</td>
<td>ND</td>
<td>59 ± 10†</td>
<td>63 ± 30</td>
</tr>
<tr>
<td>Na⁺-Ca²⁺ exchanger, amol/ng</td>
<td>533 ± 257</td>
<td>ND</td>
<td>736 ± 247†</td>
<td>652 ± 224</td>
</tr>
<tr>
<td>SR Ca²⁺-ATPase/Na⁻-Ca²⁺ exchanger</td>
<td>198 ± 118</td>
<td>ND</td>
<td>87 ± 33*</td>
<td>103 ± 47</td>
</tr>
<tr>
<td>n</td>
<td>10</td>
<td>24</td>
<td>10</td>
<td></td>
</tr>
</tbody>
</table>

Values are mean ± SD. RU indicates relative units (integral area of the 200-bp DNA fragment/μg DNA); ND, not determined.

*P < 0.01, †P < 0.05, $P < 0.05$ vs organ donors. §P < 0.05, ||P < 0.05 vs Pre-VAD.

Results

Data of HF Patients Without VAD Support

Compared with the myocardium of organ donors, the myocardium of HF patients is characterized by an increased mRNA expression of Pro-ANP and a decrease in FasExo6Del, Bcl-xL, and Bak (Table 3). Furthermore, we observed elevated apoptotic oligonucleosome-sized DNA fragmentation in left ventricles (Table 3, Figure 1A) associated with an increased abundance of apoptotic cardiomyocytes observed by the TUNEL assay (Figure 1B).

Influence of ACE Inhibitor Therapy on Bcl-xL and Bcl-2 Expression

Whereas patients without therapy by inhibitors of the angiotensin-converting enzyme (ACE) showed low mRNA levels of Bcl-xL (92.0 amol/μg RNA) and of Bcl-2 (10.1 amol/mg RNA) (n = 8), patients under ACE inhibitor treatment (n = 16) revealed partial normalization of Bcl-xL mRNA (147.8 amol/μg RNA) and Bcl-2 mRNA (20.9 amol/mg RNA) (P < 0.01 and P < 0.05, respectively). These mRNA data could be confirmed on the protein level, which was observed by Western blot analysis (Figure 2). In patients treated with ACE inhibitors compared with patients with no ACE inhibitor treatment, the Bcl-xL protein level was determined to be 1.0 versus 0.51 relative units (P < 0.001), respectively, and the Bcl-2 protein level was 0.79 versus 0.32 relative units (P < 0.09), respectively. Furthermore, we observed a significantly positive correlation between Bcl-xL or Bcl-2 protein and the mRNA levels in the group of all HF patients and donors (r = 0.488 [P < 0.05] and r = 0.461 [P < 0.05], respectively). However, these HF patients showed no significant left ventricular alterations either in apoptotic DNA fragmentation or in Pro-ANP mRNA expression or ejection fraction depending on the therapy with ACE inhibitors.

Data of HF Patients Supported by VAD

Clinical data of the 10 VAD patients revealed a highly critical hemodynamic state before VAD implantation (Table 1). During the time of ventricular support, a tendency to improved cardiac function could be assumed in these patients from decreasing cardiothoracic x-ray ratios or transthoracic echocardiography. However, these putative improvements could not be exactly quantified because the VAD was not temporarily turned off for measurements of the genuine hemodynamic parameters. This potentially risky procedure was avoided, since this observational study was not designed...
to prove the potential of cardiac recovery under VAD. Nevertheless, shortly before the orthotopic cardiac transplantation, a decrease in pulmonary capillary wedge pressure and an absence of deterioration or even an increase in the cardiac index (determined either by pulmonary artery catheter or transthoracic echocardiography) indicated the beneficial effects on hemodynamic unloading by VAD (Table 1).

**Transcription of Pro-ANP**

In left ventricular specimens, we observed a decrease in Pro-ANP mRNA expression under hemodynamic unloading approaching the mRNA levels of donor ventricles (Table 3 and Figure 3). This downregulation of Pro-ANP mRNA did not depend on time elapsed on VAD: it reached $23 \pm 18$ amol/ng RNA in patients with <100 days ($n=4$) and $31 \pm 30$ amol/ng RNA in patients with >100 days ($n=6$) of VAD support compared with the starting level of $180 \pm 207$ amol/ng RNA.

**Apoptotic DNA Laddering**

No or minimal signs of DNA fragmentation could be observed in left ventricles of donors in accordance with TUNEL-negative cardiomyocytes (Figure 1). The left ventricles of VAD-supported patients showed an intensive DNA fragmentation before VAD unloading. This DNA laddering tended to be attenuated after the time on VAD ($P=0.07$, Tables 1 and 3 and Figure 3).

**Transcription of Apoptosis-Associated Genes**

Before VAD support of terminally failing hearts, left ventricular transcription of Bcl-xL, FasExo6Del, and Bak was reduced compared with that in nonfailing left ventricles of organ donors. These data are comparable to those of HF patients without VAD support (Table 3). After VAD unloading, the Bcl-xL mRNA expression was enhanced by $29 \pm 30\%$ ($P<0.05$), and this improvement revealed a significant dependence on duration of VAD support (Figure 4A). Furthermore, Figure 4A indicates that the left ventricular Bcl-xL mRNA expression reached the mean value of donor ventricles after $\approx 130$ days of VAD unloading.

Similarly, an increase in mRNA expression of FasExo6Del in relation to Fas correlated with the duration of ventricular support in DCM patients (Figure 4B). However, this time-dependent increase in the ratio of FasExo6Del per Fas did not reach the level of increase exhibited by donor ventricles during the period of VAD unloading. In addition, the mRNA level of Mcl-1 was slightly elevated (20 $\pm$ 26%) under VAD support ($P<0.05$). For Bcl-2 and the Bcl-2-related Bak and Bax, we did not observe any transcriptional changes under support by VAD (Table 3). Consequently, the left ventricular Bcl-xL mRNA level was increased in relation to its heterodimerization partner Bak by $42 \pm 46\%$ ($P<0.06$).

The left ventricular mRNA expression of the $\alpha$ subunit of the leukocyte-specific adhesion glycoprotein p150.95, an indicator of inflammatory cell infiltration, did not change during time on VAD (0 $\pm$ 7%).

**mRNA Expression of Na$^+$-Ca$^{2+}$ Exchanger and SR Ca$^{2+}$-ATPase**

mRNA expression of the Na$^+$-Ca$^{2+}$ exchanger tended to be elevated in the left ventricles of HF patients before VAD support compared with the left ventricles of donor hearts, whereas mRNA of the SR Ca$^{2+}$-ATPase tended to be lowered, and the ratio of SR Ca$^{2+}$-ATPase per Na$^+$-Ca$^{2+}$ exchanger was significantly reduced. Under hemodynamic unloading, left ventricular transcription of the SR Ca$^{2+}$-ATPase as well as of the Na$^+$-Ca$^{2+}$ exchanger remained at failing levels despite this mechanical support (Table 3).

However, in the left ventricles of 4 patients (Nos. 3, 4, 6, and 8; see Table 1), an improvement of the mRNA ratio of SR Ca$^{2+}$-ATPase per Na$^+$-Ca$^{2+}$ exchanger ($\approx 40\%$; ie, larger than

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**Figure 1.** Comparison of left ventricular data from patients with end-stage HF and donors. A, DNA gel electrophoresis of 1.5 $\mu$g DNA indicates apoptotic DNA fragmentation in HF patients. M indicates 100-bp DNA marker. B, Apoptotic cardiomyocyte nuclei detected by TUNEL assay appear brown-stained and are indicated by arrows.

**Figure 2.** Left ventricular protein expression of Bcl-xL and Bcl-2 from organ donors and from patients with end-stage HF who were treated with ACE inhibitors (ACE i-pos.). Western blot analysis indicates myocardial Bcl-xL as 30-kDa and Bcl-2 as 25-kDa protein.
the SD of pre-VAD values) could be observed. These 4 patients suffered from DCM, and post hoc comparison with the remaining 4 DCM patients without such improvement of the SR Ca\textsuperscript{2+}-ATPase/Na\textsuperscript{+}-Ca\textsuperscript{2+} exchanger mRNA level under VAD indicated that they also showed more improved gene expressions of antiapoptotic factors. Their myocardial mRNA levels of FasExo6Del were increased by 94±645% (P<0.05), of Mcl-1 by 42±9% (P=0.07), and of Bcl-xL/Bak by 59±49% (P=0.05) during the time on VAD. Similar elevations were not observed in the other subgroup for FasExo6Del (7±80%), Mcl-1 (9±41%), or Bcl-xL/Bak (33±52%), although the duration of VAD support was not significantly different in both subgroups (102±52 versus 84±52 days in patients with versus without increased SR Ca\textsuperscript{2+}-ATPase/Na\textsuperscript{+}-Ca\textsuperscript{2+} exchanger mRNA level). The subgroup without a VAD-induced increase in SR Ca\textsuperscript{2+}-ATPase/Na\textsuperscript{+}-Ca\textsuperscript{2+} exchanger mRNA expression may have been in a worse condition immediately before VAD implantation: their cumulative dosage of phosphodiesterase inhibitors, needed for positive inotropic treatment, was 564±356 mg milrinone equivalents (in case of the use of enoximone, the enoximone dosage in milligrams was divided by 5.5), and the last monitored pulmonary wedge pressure was determined to be 35±5 mm Hg. These values tended to be lowered in the subgroup with an improved expression of the mentioned genes under VAD: the pulmonary wedge pressure was 25±6 mm Hg (P=0.06 versus the other subgroup) and the cumulative milrinone equivalent dosage was 233±191 mg.

Discussion

In the present study, we report that the HF patients who required VAD support exhibited an apoptosis-susceptible myocardial phenotype and an abnormal myocardial mRNA expression of calcium-regulatory proteins before VAD unloading. Compared with donor hearts, the hearts of the HF patients exhibited increased cardiac DNA fragmentation with more abundant TUNEL-positive cardiomyocytes; these data are in agreement with the data of Olivetti et al.\textsuperscript{9} which additionally showed that in overloaded human myocardium apoptosis mainly affects cardiomyocytes. Thus, the reduction in myocardial DNA laddering under VAD (Tables 1 and 3) probably reflects the decrease in the ongoing apoptotic loss of cardiac myocytes, because mRNA expression of the leukocyte adhesion molecule p150.95,\textsuperscript{23} an indicator of inflammatory cell
of the renin-angiotensin system, a decreased expression of antiapoptotic versus proapoptotic Bcl-2-related proteins, and an elevated translation of the Fas receptor. Furthermore, an increase in cardiomyocyte apoptosis and the changed expression of Bcl-2 relatives by experimental overactivity of ACE could be renormalized by angiotensin I receptor blockade or by long-term therapy with ACE inhibitors. Confirming these experimental data, expressions of Bcl-xL and Bcl-2 were partially normalized by ACE inhibitor therapy in the failing human myocardium of our patients. Since nearly all VAD patients were treated with ACE inhibitors, VAD unloading has an additionally stimulatory effect on the mRNA expression of the apoptosis-preventing Bcl-xL, which correlates with the Bcl-xL protein. Furthermore, after ventricular support of patients who had a coronary heart disease, an upregulation of the Bcl-xL protein could be observed, too. However, the ventricular unloading, either VAD-mediated or ACE inhibitor–mediated, cannot be the sole stimulus: the overload-indicating Pro-ANP mRNA level was identical in the non–VAD-supported group of patients with and without ACE inhibitor therapy.

Although our data indicate partial apoptotic phenotype normalization under VAD support with cardiac apoptosis reduction of the failing myocardium, these results cannot yet provide an assessment of the quantitative relevance of myocyte apoptosis. Furthermore, they cannot indicate the therapeutic potential of assist devices in terminal HF for several reasons: First, the analysis could not be designed as a treatment study. Second, we analyzed myocardium only from those patients who experienced a clinical stabilization under assist device unloading. Not all patients in such a deteriorated state of terminal HF can be successfully stabilized by a VAD. Third, it must be remembered that the myocardium of some patients may have lost this hypothetical potential for recovery because of irreversible remodeling processes, like fibrosis and scar formation.

Finally, it remains to be stressed that a reduction in apoptosis is only one of several basic mechanisms required for a functional recovery of the failing heart. Therefore, mRNA expressions of SR Ca2+–ATPase and Na+/Ca2+–exchanger were additionally determined. However, the mRNA ratio of the SR Ca2+–ATPase per Na+/Ca2+–exchanger was normalized in only a few patients. Since a correlation between impaired mRNA and protein levels of the SR Ca2+–ATPase and the decreased SR 45Ca2+ uptake has been demonstrated in the failing human heart, our results are comparable to those of Frazier et al., who observed an enhanced myocardial calcium uptake/binding in the SR from some HF patients under chronic VAD support. As mentioned above, the myocardium of only some patients will be capable of renormalizing the expression of calcium-regulatory determinants under VAD for irreversible injuries. Furthermore, VADs are presently used only in patients who cannot be stabilized otherwise, and up to now, only a few selected patients could be weaned from the VAD without a subsequent heart transplantation. The selective use of VADs is necessary because it seems not to be justified to withhold an organ offer from those patients who need it. It is plausible to postulate that the VAD application in patients with less deteriorated
cardiac function could be more successful. In agreement with this assumption, our observation that the subgroup with VAD-induced improvement of gene expression for antiapoptotic and calcium-handling proteins tends to exhibit a somewhat less deteriorated hemodynamic status immediately before VAD implantation, can as be deduced from a diminished cumulative phosphodiesterase inhibitor dosage and a lower pulmonary wedge pressure.

In conclusion, our findings indicate some potential of VAD as an intervention to interrupt the vicious cycle of distortion-induced proapoptotic phenotype shifts, and in the future, VAD may become a treatment option for recovery in a subgroup of patients otherwise undergoing transplantation.

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Myocardial Gene Expression of Regulators of Myocyte Apoptosis and Myocyte Calcium Homeostasis During Hemodynamic Unloading by Ventricular Assist Devices in Patients With End-Stage Heart Failure

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